ENTERED

Department of Energy
Carlsbad Field Office
P. O. Box 3090
Carlsbad, New Mexico 88221

DEC 14 2011

Mr. John Kieling, Acting Chief
Hazardous Waste Bureau
New Mexico Environment Department
2905 Rodeo Park Drive East, Building 1
Santa Fe, New Mexico 87502-6303

Subject: Waste Isolation Pilot Plant Contract Laboratory Standard Operating Procedure Updates

Dear Mr. Kieling:

This letter transmits a summary of the standard operating procedures (SOPs) used by the Permittees' contract laboratories for groundwater, volatile organic compound, and hydrogen & methane analyses that were updated during calendar year 2011. Enclosed is a CD-ROM containing updated electronic copies for the Waste Isolation Pilot Plant 2011 Lab SOP Portfolios.

These revised SOPs are being provided to comply with Permit Attachment L, Section L-4c (3) and Attachment N, Section N-4e.

We certify under penalty of law that this document and all attachments were prepared under our direction or supervision in accordance with a system designed to assure that qualified personnel properly gather and evaluate the information submitted. Based on our inquiry of the person or persons who manage the system, or those persons directly responsible for gathering the information, the information submitted is, to the best of our knowledge and belief, true, accurate, and complete. We are aware that there are significant penalties for submitting false information, including the possibility of fines and imprisonment for knowing violations.

If you have any questions regarding these procedure updates, please contact Ms. Susan McCauslin at (575) 234-7349.

Sincerely,

Edward Ziemianski, Interim Manager
Carlsbad Field Office

M. F. Sharif, General Manager
Washington TRU Solutions LLC

Enclosure

cc: w/enclosure

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SUMMARY

Contract Laboratory Standard Operating Procedures Used for the WIPP Groundwater Detection Monitoring, Volatile Organic Compound Monitoring, and Hydrogen & Methane Monitoring Programs, December 2011

Part 1. Hall Environmental Analysis Laboratory (Groundwater)

The Standard Operating Procedures (SOPs) for the contract laboratory, Hall Environmental Analysis Laboratory (HEAL), are listed below in the index table. HEAL subcontracts two analytical processes to Anatek Labs, Inc. (Anatek) and Columbia Analytical Services, Inc.(Columbia). The SOPs associated with these analyses are also included in the index table. The SOP from Anatek has changed and is summarized in an independent summary sheet with the attached electronic procedures for Method 6020. Attached SOPs from HEAL do not itemize the changes within the procedures or highlight these changes; however, the HEAL SOP change summary table summarizes the modifications. Procedures with changes are submitted in compliance with Permit Attachment L, Section L-4c(3).

INDEX TO CONTRACT LABORATORIES STANDARD OPERATING PROCEDURES

Hall Environmental Analysis Laboratory

HEAL WIPP SOP List		
Test	Latest SOP Revision	Revised since last submittal to NMED?
Specific Conductance	S-120.1-6	No
Mercury	S-245.1_7470-3	No
Specific Gravity	S-SM2710F-0	No
Volatiles	S-8260-10	Yes
Semi-Volatiles	S-8270C-10	Yes
Total Recoverable Metals, Dissolved Metals	S-6010-8	Yes
pH	S-SM4500H+ 6	Yes
Total Organic Carbon	S-TOC-5	Yes
Total Dissolved Solids	S-SM2540C-6	Yes
Total Suspended Solids	S-SM2540D-3	Yes
Alkalinity	S-SM2320B-6	Yes
Anions	S-300.0-10	Yes

Anatek WIPP SOP List		
Test	SOP Revision	
Trace Metal Analysis	ALI-A-315.06	Yes
Columbia WIPP SOP List		
Test	SOP Revision	
Total Organic Halogen	GEN-9020, Rev.8	No

Hall Environmental Analysis Laboratory SOP Change Summary

Standard Analytical Method	Current SOP No.	Effective Date	Supersedes	Summary of Changes
EPA SW-846 Method 8260 (VOCs)	S-8260-10	8/26/2011	S-8260-9	Sections 1.2, 9.2, 9.2.1 and Table 1 were updated to replace the previous internal standard Fluorobenzene with Pentafluorobenzene, to include the new standard concentration and the analyte references. Section 1.3 was updated to reference the Laboratory Information Management System (LIMS) for the current Practical Quantitation Limit (PQLs). The annual requirement for all analytes to be spiked in the Laboratory Control Sample (LCS) has been extended from annually to every two years in section 6.15.4. The sample storage requirements were updated to clarify above freezing to 6°C in section 7.2. Appendix 1 and a reference to the appendix in Section 10.5 were included to detail the procedure for low-level soil analysis. Section 11.3.1 has been updated to clarify the qualification ion requirements for review and interpretation, including corrective action requirements. Appendix 2 and a reference to the Appendix in section 12.2.2 have been added to include a calibration curve summary form. Section 12.2.6.6 has been updated to increase control chart frequency from annually to twice per year.
EPA SW-846 Method 8270 (SVOCs)	S-8270C-10	8/26/2011	S-8270C-9	Section 1.3 has been condensed and no longer includes the step-by-step instructions for reviewing limits in the LIMS. The sample storage requirements were updated to clarify above freezing to 6°C in section 7.4. Section 11.1.1 was updated to add the requirement for calculating the tailing factor for Pentachlorophenol and Benzidine every twelve hours. Section 11.3.1 has been updated to clarify the qualification ion requirements for review and interpretation, including corrective action requirements. The Appendix, as well as a reference to it in section 12.2.2, has been added to include a calibration curve summary form.

Standard Analytical Method	Current SOP No.	Effective Date	Supersedes	Summary of Changes
EPA SW-846 Method 6010 (Metals)	S-6010-7	12/1/2010	S-6010-6	Section 3.2 references a PQL summary in Appendix A. Matrix specific Quality Control (includes matrix spike, matrix spike duplicate, post digestion spike, and dilution test) added in 4.9, 10.3, 16.3.C.3, 17.3.C and Appendix F. 7.1.A updated to reference the maintenance logbook for the current hardware and software in use. 7.4 updated to include re-pipettors. 8.3-8.5 details how to make and document the 2% and 5% Nitric acid solutions. Details were added to 8.6.E for the U, Sn, Sr and Si spike solution. 8.7.A, 8.9.A, 8.10.A and 8.10.B added the documentation requirements for spikes and standards. 8.7.C and 8.8.B outline the calibration and verification procedures for Sr. In 9.6 and 9.7 the hold time was corrected to 180 days. Secure login added to 12.2.A. 13.5 added to include the formulas for Hardness and Sodium Absorption Ration (SAR). The Uncertainty formula was removed from 14.2 and now references the Quality Assurance Plan (QAP) for the formula. The Annual Documentation of Continued Proficiency (ADOCP) details were added to 14.3.B. 16.3.A.2 and 16.3.B.1 clarify that an Relative Standard Deviation) of 5% is acceptable. 17.1.E discusses corrective action for iron interferences. 17.1.F details the procedure for reprocessing Boron and Cobalt using alternative Multi-component Spectral Fitting models. Added additional references to 20.0. Appendix H details the SAR procedure. There were minor grammatical corrections throughout the document.
EPA SW-846 Method 6010 (Metals)	S-6010-8	8/31/2011	S-6010-7	Section 2.3 clarifies that results are never blank corrected. Section 3.2 was updated to refer to the QAP for the Method Detection Limit (MDL) procedure and calculations. Laboratory Control Sample Duplicate (LCSD) definition was added to section 4.9. Updated materials section 7.0 to include flasks up to 1000 mL, and an analytical balance and glass syringes have been added. Details added to section 8.7 and 8.8 for the cal/spike and secondary standards. Section 9.2 clarifies that preservation is to have a pH <2. Sections 10.3.A.1 and 10.3.B require the analysis of an LCSD when volume is insufficient for an Matrix Spike Duplicate (MSD). The Exchangeable Sodium Percentage formula has been added to section 13.7. Section 14.1 updated Performance Test frequency from routinely to every 6 months. Uncertainty information was removed from section 14.2. Negative blank limits were added into the procedure in sections 16.3.A.3, 16.3.B.2 and 16.3.C.1. A note was added after section 16.3.C.2 to clarify that the instrument LCS recovery should not be used as a final determination of pass/fail. In section 17.1 the allowable numbers of re-analyses for Quality Control (QC) failures was reduced from 2 to 1. The corrective action Procedure Summary in Appendix I is referenced in section 17.3.E.

Standard Analytical Method	Current SOP No.	Effective Date	Supersedes	Summary of Changes
				There were minor grammatical corrections throughout the document.
Standard Methods SM4500-H^+ (pH)	S-SM4500-H^+-6	8/29/2011	S-SM4500-H^+-5	The sample storage requirements were updated to clarify above freezing to 6°C in Section 9.1 and the hold time for compliance samples was changed to 15 minutes and non-compliance samples to 24 hours. Section 10.1.1.3.1 was updated to list an acceptable calibration slope up to 102% from the previous 101%. Section 11.1.7.6 clarifies Initial Calibration Verification instrument programming requirements. The uncertainty formulas have been removed from Section 132.3 and a reference to their location in the QAP has been included.
Standard Methods SM5310B (TOC)	S-TOC-4	12/14/2010	S-TOC-3	Section 3.1 was updated to reference the LIMS and not as an addendum for the current MDLs. Section 7.6 removed the requirement for amber sample vials. Section 7.9 was updated to include the use of a 500-mL syringe. The procedure for making the second source standard in Section 8.3.2 was updated to state that the standard is now purchased pre-made from a vendor.
Standard Methods SM5310B (TOC)	S-TOC-5	6/3/2011	S-TOC-4	Sections 4.2, 4.3, 10.2.1, 11.2.1 and 16.1.3.1 were updated to require calibration verification after every tenth analysis instead of every fifteen analyses. Section 16.1.1 was included to detail the criteria the instrument uses for determining replicate acceptability. Sections 16.1.2.3 and 16.1.4.3 were updated to establish negative blank requirements.
Standard Methods SM2540C (TDS)	S-SM2540C-5	3/4/2011	S-SM2540C-4	Removed the allowance in section 3.0 to use Total Suspended Solids (TSS) filters if TSS is also to be determined. Section 7.0 updated to include a 100-mL graduated cylinder and forceps as materials. Section 9.1 updated to clarify that samples must be stored above freezing to 6°C. Sections 11.2 and 11.3 have been reworded to clarify the batch preparation procedures. Section 11.4.1 added to state the filtration apparatus is cleaned between each sample and to allow the reuse of filters until they are clogged or pose a risk of tearing. There were minor grammatical corrections throughout the document.
Standard Methods SM2540C (TDS)	S-SM2540C-6	9/14/2011	S-SM2540C-5	Section 13.2 was updated to require that the MDL study be performed over a minimum of three days. Section 15.5 was changed from an acceptable MSD Relative Percent Difference of less than 20% to less than or equal to 5%.

Standard Analytical Method	Current SOP No.	Effective Date	Supersedes	Summary of Changes
Standard Methods SM2540D (TSS)	S-SM2540D-3	3/4/2011	S-SM2540D-2	Section 9.1 was updated to clarify that samples must be stored above freezing to 6°C. Section 11.2 extended the oven calibration frequency from monthly to quarterly. Added the HEAL Balance Calibration SOP to the References in section 20.0. There were minor grammatical corrections throughout the document.
Standard Methods 2320B (Alkalinity)	S-SM2320-5	6/15/2011	S-SM2320-4	Sections 1.2.2, 7.2, 11.1 were updated for the new pH meter. The procedure and calculations for CO2 were added to 1.4 and 12.3. PQL updated to 20 and low level procedure removed from Sections 1.3 and 2.1. 11.1.3 details the low-level procedure should it be needed. Materials added and updated in Section 7.0. Temperature storage requirements updated in 8.1 from </=6°C to 0-6°C. Details were added to 10.1.9.1 and 10.1.10.1 for the preparation of batch QC. 10.1.11 was updated to require that the LIMS Identification (ID) for spikes and standards be entered into the ID2 line of the automated instrument. 10.1.15.1and 11.1.2.3.1 were updated for dilutions being made using 20 mL and to clarify how to document the reduced volume. 11.1.2.2.1.1 and 11.1.2.4.1.1 refer to the Variable Volume Pipette Calibration Check SOP for the proper handling of pipettes. 13.3.1 and 17.4.2 added the use of control chart limits for the LCS. The uncertainty formula was removed from 14.1 and a reference to the QAP was included. Details were added to 17.0 and 18.0 to clarify data reduction procedures, and the Method Blank acceptable limits were updated from </= 2 to 20mg/L. Additional references were added in section 21.0.
Standard Methods 2320B (Alkalinity)	S-SM2320-6	8/29/2011	S-SM2320-5	Sections 3.3 and 13.2 were updated to require MDLs to be conducted annually in place of PQL studies, and that they must be completed over a minimum of three days. Section 8.1 was updated to clarify that samples must be stored above freezing to 6°C. Section 9.3 was added to require that a bracketing pH Continuing Calibration Verification (CCV) be analyzed every 20 samples and at the end of each batch. Section 17.5.1 establishes the acceptable CCV limit of +/- 0.1 and Section 18.3 defines contingencies for CCV failures. The blank correction procedure for PQL studies was removed from the calculations in section 12.0.
EPA Method 300.0 (Anions)	S-300.0-9	4/20/2011	S-300.0-8	Section 3.1 was corrected to state that MDL studies are done every six months. Section 7.1 was updated to include the Ion Chromatography System (ICS)-1500 and all of its associated hardware. Section 8.4 has been updated to include mobile phase preparation for the ICS-1500. Details have been added to section 8.5.1 for the calibration standard mixes. Sections 9.1.2 and 9.2.1 have been updated to clarify that samples must be stored above

Standard Analytical Method	Current SOP No.	Effective Date	Supersedes	Summary of Changes
				freezing to 6°C. Sections 11.1.2 and 11.3.2 reference the 2-mL autosampler vials for the ICS-1500. Section 11.3.7 has been added to include requirements for the AS-50 autosampler. Section 13.3.2 has been updated to require that for LCSs to be used for ADOCPs must be prepared from a second source. The justification for not conducting the bi-annual Linear Calibration Range in section 13.4.2 has been replaced by a reference to the method for the procedure. Section 15.3.3.3 has been updated to reference the LIMS for the current limits instead of an SOP addendum. Section 16.3.3.5 clarifies that Matrix Spike/Matrix Spike Duplicate results are not to be reported for analytes with values that exceed the range of the calibration curve.
EPA Method 300.0 (Anions)	S-300.0-10	8/31/2011	S-300.0-9	Section 3.1 was corrected to reference the LIMS for the current MDLs and not the addendum to the SOP. Sections 8.4.1.2.8 and 8.4.2.2.9 have been added to require documentation on the eluent bottle. Section 8.4.2.3 has been added to improve eluent traceability. Section 8.6.1 has been updated to require calibration standards be made fresh on the day of use. The procedure for evaluating calibration fits has been added to section 10.1.5. Section 11.3.2 references the IC Analysis Summary sheet that has been added to Appendix H. Sections 11.3.7 and 11.3.17 have been added to clarify that the instrument sequence list is the original documentation of sample dilution. Section 12.3 was updated to reference the QAP for the calibration calculations.

Contract Laboratory Standard Operating Procedures Used for the WIPP Groundwater Detection Monitoring, Volatile Organic Compound Monitoring, and Hydrogen & Methane Monitoring Programs, December 2011

(Continued)

Part 2. Carlsbad Environmental Monitoring and Research Center (Volatile Organic Compound Hydrogen and Methane Monitoring)

The Standard Operating Procedures and Quality Assurance Project Plan used by Carlsbad Environmental Monitoring and Research Center (CEMRC), to provide contract analytical laboratory services are listed below in the following table. All of the procedures either had significant changes to the process or minor editorial changes. Procedures with changes are submitted in compliance with Permit Attachment N, Section N-4e. Changes are noted on the procedures in the Temporary Procedural Deviation Form, Revision History, and are highlighted.

INDEX TO CONTRACT LABORATORIES STANDARD OPERATING PROCEDURES

CEMRC, 2011

Procedure #	Revision # _ Date	Effective Date	# of temp. dev.	Revised since last submittal to NMED?
OC-PLAN-001-003-020106 QAPP for VOCs_EAS_	3 _ 05/05/10	May 5, 2010	1	Yes
OC-PROC-002-005-021506 Prep Canisters Sample Trains	5 _ 09/08/11	September 8, 2011	0	Yes
OC-PROC-003-003-021706 GCMS of VOC_EAS	3 _ 05/03/10	May 3, 2010	2	Yes
OC-PROC-004-004-021606 Prep Cal Stds	4 _ 04/19/10	April 19, 2010	2	Yes
OC-PROC-005-005-022106 data validation	5 _ 05/17/11	May 17, 2011	0	Yes
OC-PROC-006-003-022006 Gas Samples in passivated canisters	3 _ 05/03/10	May 3, 2010	2	Yes
OC-PROC-009-002-110907 Hydrogen Methane.	2 _ 09/08/11	September 8, 2011	0	Yes

INDEX TO CONTRACT LABORATORIES STANDARD OPERATING PROCEDURES

CEMRC, 2011

Procedure #	Revision # _ Date	Effective Date	# of temp. dev.	Revised since last submittal to NMED
OC-PLAN-001-003-020106	3 _ 05/05/10	May 5, 2010	1	+1 temp dev
OC-PROC-002-005-021506	5 _ 09/08/11	September 8, 2011	0	Yes
OC-PROC-003-003-021706	3 _ 05/03/10	May 3, 2010	2	+1 temp dev
OC-PROC-004-004-021606	4 _ 04/19/10	April 19, 2010	2	+1 temp dev
OC-PROC-005-005-022106	5 _ 05/17/11	May 17, 2011	0	Yes
OC-PROC-006-003-022006	3 _ 05/03/10	May 3, 2010	2	+1 temp dev
OC-PROC-009-002-110907	2 _ 09/08/11	September 8, 2011	0	Yes

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TEMPORARY PROCEDURAL DEVIATION FORM

Carlsbad Environmental Monitoring & Research Center

Title: Quality Assurance Project Plan for Analysis of Volatile Organic Compounds and/or Hydrogen and Methane in Canister Samples	
Document No: OC-PLAN-001	**Implementing Program: Organic Chemistry**

Deviation:

On page 4, section 5.0, read 'The VOC/HM analysis methods used at CEMRC for the VOCMP/HMMP are based on concepts contained in two EPA documents: (1) EPA, Compendium Method TO-15, 1999 and (2) EPA Draft Contract Laboratory Program Statement of Work (CLP-SOW; Volatile organics Analysis of Ambient Air Canisters, 1994). Hydrogen and Methane analyses are conducted based on methods established for use in Headspace Gas Analysis for WIPP waste'.

It should be read as follows, 'The VOC analysis methods used at CEMRC for the VOCMP are based on concepts contained in US EPA, Compendium Method TO-15, 1999 and EPA Draft Contract Laboratory Program Statement of Work (CLP-SOW; Volatile organics Analysis of Ambient Air Canisters, 1994). Hydrogen and Methane analyses are conducted based on the concepts described in EPA Compendium Method TO-15, EPA CLP-SOW, EPA SW-846 8260B, and ASTM D-1946 (ASTM 2006)'.

Justification: The aforementioned deviation is being implemented to update the reference for Hydrogen and Methane analyses.

Valid for: ___**Until Next Revision**_____

Approved by: _____ **Responsible Manager** 08/08/11 **Date**

STAFF TRAINING

I have reviewed and I understand the procedure deviation described above.

Print Name	Signature	Date

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CARLSBAD ENVIRONMENTAL MONITORING & RESEARCH CENTER (CEMRC)

QUALITY ASSURANCE PROJECT PLAN FOR ANALYSIS OF VOLATILE ORGANIC COMPOUNDS AND/OR HYDROGEN AND METHANE IN CANISTER SAMPLES

OC-PLAN-001-003-020106-_ _ _

Effective Date: May 5, 2010

CARLSBAD ENVIRONMENTAL MONITORING & RESEARCH CENTER

NEW MEXICO STATE UNIVERSITY
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QUALITY ASSURANCE PROJECT PLAN FOR ANALYSIS OF VOLATILE ORGANIC COMPOUNDS AND/OR HYDROGEN AND METHANE IN CANISTER SAMPLES

1.0 PREFACE

This document describes the implementation of quality assurance plans specific to the analysis of volatile organic compounds (VOC) and hydrogen and methane (HM) for the "Volatile Organic Compound Monitoring Plan" and "Hydrogen and Methane Monitoring Plan," hereinafter abbreviated as VOCMP and HMMP, respectively, issued as Attachment N and Attachment N1 of the Waste Isolation Pilot Plant Hazardous Waste Facility Permit.

This document is an addendum to the Quality Assurance Plan for the Carlsbad Environmental Monitoring & Research Center (CP-QAP-004). As stipulated in the CP-QAP, a project specific addendum such as this lists and describes all quality-related requirements that are over and above or in variance with the minimum requirements specified by the CP-QAP.

The purpose of the VOCMP/HMMP is to confirm that the concentrations of VOC/HM in the WIPP underground do not exceed permit-specified action levels. The Organic Chemistry (OC) group at CEMRC will be responsible for the VOC/HM analyses as specified in the Statement of Work (SOW) issued by WTS. As outlined in the SOW, the VOC/HM analyses done at CEMRC must be done in accordance with selected Quality Assurance (QA) Requirements from the American National Standards Institute, ANSI/ASME NQA-1 "Quality Assurance Program Requirements for Nuclear Facilities," specified in Table 1.

Table 1. List of Applicable NQA-1 Requirements for VOC/HM Analyses

Quality Item	Element
1	Organization
2	Quality Assurance Program
4	Procurement Document Control
5	Instructions, Procedures, and Drawings
6	Document Control
7	Control of Purchases Items and Services
8	Identification and Control of Items
12	Control of Measuring and Test Equipment
13	Handling, Storage, and Shipping
15	Control of Nonconforming Items
16	Corrective Action
17	Quality Assurance Records

Many of these NQA-1 requirements are addressed in the CP-QAP. In those cases where the requirements are covered, references to the appropriate sections of the existing documents are

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given below, in some cases with additional clarification. The main purpose of this implementation plan is to provide information relating to the NQA-1 requirements not explicitly addressed in existing documents.

1.1 Acronyms

ANSI - American National Standards Institute
CEMRC - Carlsbad Environmental Monitoring and Research Center
CLP - Contract Laboratory Program
DOT - Department of Transportation
EPA - Environmental Protection Agency
HM - Hydrogen and Methane
HMMP - Hydrogen and Methane Monitoring Program
M&TE - Measuring and Testing Equipment
NMSU - New Mexico State University
OC - Organic Chemistry
QA - Quality Assurance
QAP - Quality Assurance Plan
QC - Quality Control
SOP - Standard Operating Procedure
SOW - Statement of Work
VOC - Volatile Organic Compound(s)
VOCMP - Volatile Organic Compound(s) Monitoring Program
WIPP - Waste Isolation Pilot Plant
WTS – Washington True Solutions

2.0 ORGANIZATION

The organizational structure for the CEMRC is presented in Section 2.0 Management of CP-QAP-004: this includes information on general responsibilities (Section 2.1), program oversight (Section 2.2), stop work and resume work authority (Section 2.3; see also CP-PROC-012, "Nonconformances and Non-Routine Events"), and resolution of disputes (Section 2.5).

Briefly, the Director of CEMRC is responsible for ensuring that all work performed at CEMRC is done in accordance with all applicable professional, quality, and client standards. The responsibility for briefing the Director on quality assurance matters is one of the main duties of the Center's QA Manager. The QA Manager is also responsible for briefing the laboratory manager with regards to any relevant findings, issues, or concerns related to QA and QC procedures. Analysts are responsible for informing the laboratory manager of any trends in analyte recoveries, instrument performance, or any other significant factors that may affect quality.

For the VOC/HM analyses, OC analysts are responsible for reviewing all batch quality control data against default or client-specified criteria. Data are subjected to peer or supervisory review for general compliance with routine QA criteria, and the laboratory manager conducts periodic

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in-depth review of data submittals from the analysts. Both internal and external QA program reviews are performed as part of the managerial process.

3.0 QUALITY ASSURANCE PROGRAM

CEMRC has in place a quality assurance program that was established under ANSI/ASME NQA-1. The CEMRC "Quality Assurance Plan" (CP-QAP-004) was approved by the Carlsbad Area Office of the US DOE on 26 September 2002.

Additionally, the VOCMP/HMMP analyses comply with applicable requirements in "Quality Assurance Project Plan for Volatile Organic Compound Monitoring", WP 12-VC.02, current revision.

4.0 PROCUREMENT DOCUMENT CONTROL

Descriptions of the procedures used for procurements and procurement records may be found in CEMRC document CP-PROC-017, "Procurement of Items and Services", document CP-PROC-023, "Assignment of a Quality Level for Procured Items and Services", and document CP-PROC-024, "Assessment of Supplier Qualifications and Procedure for the Addition of Suppliers to the Qualified Suppliers List".

5.0 INSTRUCTIONS, PROCEDURES, AND DRAWINGS

The VOC/HM analysis methods used at CEMRC for the VOCMP/HMMP are based on concepts contained in two EPA documents: (1) EPA, Compendium Method TO-15, 1999 and (2) EPA Draft Contract Laboratory Program Statement of Work (CLP-SOW; Volatile Organics Analysis of Ambient Air Canisters, 1994). Hydrogen and methane analyses are conducted based on methods established for use in Headspace Gas Analysis for WIPP waste. All work for the VOCMP/HMMP shall be performed in accordance with controlled, validated, and approved procedures that were developed based on the concepts in these documents. Validation of the procedures prior to routine analysis is required to establish applicability and to ensure that the established quality objectives are met prior to program initiation.

SOPs for six unique activities related to the VOCMP/HMMP have been developed; these are to be consistent with the quality assurance objectives (precision, accuracy, sensitivity and completeness) and requirements established in the WTS SOW. The SOPs are to be maintained as controlled documents at the Center and they are listed in Table 2.

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Table 2. List of Standard Operating Procedures used for the Confirmatory Volatile Organic Compound Monitoring Plan and Hydrogen and Methane Monitoring Plan

Controlled Document Identifier	Title
OC-PROC-002	Preparation of Canisters and Sample Trains for Ambient Air Sampling
OC-PROC-003	Gas Chromatography-Mass Spectrometry Analysis of Volatile Organic Compounds (VOC) in Ambient Air from Canisters at ppbv Concentration Levels
OC-PROC-004	Preparation of Calibration Standards in Specially Prepared Canisters for Analysis by Gas Chromatography/Mass Spectrometry
OC-PROC-005	Data Validation and Reporting of Volatile Organic Compounds from Gas Chromatography/Mass Spectrometry Analysis of Ambient Air in Canisters for the WIPP Volatile Organic Compound Monitoring Plan
OC-PROC-006	Receipt, Control, and Storage of Gas Samples in Passivated Canisters
OC-PROC-009	Analysis of Hydrogen and Methane in Passivated Canisters Using Gas Chromatography with Thermal Conductivity Detection

6.0 DOCUMENT CONTROL

Documents and files related to the VOCMP/HMMP are organized in such a way as to allow rapid and routine access to information regarding samples and data. The files are designed with the intention of assembling and archiving information on sample receipt, sample storage, sample processing, analysis, and QC/QA. Much of this information also is included in the hard copy or electronic data deliverables.

The universal aspects of document control at CEMRC are covered in the following documents: CP-PROC-013, "Records Maintenance, Storage, and Establishment of a Filing System" and DC-PROG-001, "Document Control Program".

7.0 CONTROL OF PURCHASES, ITEMS, AND SERVICES

CEMRC scientists are required to use materials, equipment and services that have acceptable quality characteristics for all related work conducted at the Center. The purchase of items and services by the Center staff is controlled by procurement processes in accordance with NMSU procedures and regulations and with CEMRC documents CP-PROC-017, "Procurement of Items and Services", CP-PROC-023, "Assignment of a Quality Level for Procured Items and Services", and CP-PROC-024, "Assessment of Supplier Qualifications and Procedure for the Addition of Suppliers to the Qualified Suppliers List".

Particular attention is given to the following items which are considered to be quality related (specifications for these items are normally required for procurement purposes):

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- calibration reference materials;
- chemicals and reagents;
- analytical instruments;
- measurement and test equipment such as volumetric labware, analytical pipettes, balances, thermometers, pH meters; and
- computer software

Quality-related services include such critical functions as the calibration and repair of analytical balances and development of computer software.

The laboratory manager or their designee shall be responsible for evaluating whether procurement requires quality-related specifications, and when appropriate, this requirement is noted on the Center's Purchase Request Forms.

Items and services can be categorized into standard or special categories. Standard items and services are those that are typically considered to be "off-the-shelf" items or services, which are normally obtained from a vendor's stock or catalog of services. Specifications and descriptions for such items or services are typically supplied by the vendor when needed. Such items and services often require minimal procurement control.

Special items and services are those that are critical, complex, or costly and are procured in accordance with specifications supplied by the analytical requirements for specific projects. The procurement of special items and/or services is reviewed by the laboratory manager or their designee, in addition to the QA Manager or their designee. When appropriate, the specifications are also reviewed by the CEMRC Director for compliance with applicable regulatory, contractual, and financial requirements. The director also gives the final approval for the procurement of special items and services.

Certain quality related materials can be supplied with certificates that document testing results or other critical specifications from manufacturer or vendor. When procuring such materials, the OC group shall request any relevant certificates of content or verification of specifications. Similarly, during the procurement process, certificates for reagents, pipettes, secondary-standard balance weights, etc. may be requested.

Purchase requisitions generated by the technical staff shall be reviewed and authorized for transmittal to the NMSU Purchasing Department by the laboratory manager. The review process shall include the review of quality-related specifications, delivery time requirements, chemical safety limitations, adherence to DOT regulations, NMSU policies, and special care, packaging and receipt requirements.

Copies of all material/product certificates and certificates of conformance shall be kept in the CEMRC procurement documentation files.

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8.0 IDENTIFICATION AND CONTROL OF ITEMS

Identification and control of items at CEMRC is described in detail in Section 6.5 (Identification and Control of Items) of the CP-QAP. As noted above, the identification of quality-related items begins with the purchasing process, see Section 8.1 "Initiate Procurement" in CP-PROC-017, "Procurement of Items and Services." The next step in ensuring quality of items is in the receipt process, which is described in Section 8.4 "Receiving Standard Items" of CP-PROC-017.

Chemicals and reagents are routinely checked indirectly through the analysis of procedural blanks. Standards are other commonly used items requiring identification and control, and specific quality-related procedures for standards are described in CP-PROC-009, "Standards Log-in and Preparation." When questions about the purity of chemicals, reagents or standards arise, they are re-tested and if they are found to be outside acceptable control parameters, they either are replaced or purified.

9.0 CONTROL OF MEASURING AND TEST EQUIPMENT

Control of measuring and test equipment at CEMRC is described in detail in Section 6.4 of the CP-QAP. Quality-related measurement and testing equipment (M&TE) is identified in the SOPs for the VOCMP/HMMP, and these documents also contain information on calibrations and procedures to be followed when M&TE is found to be out of calibration. Log books are appropriately annotated with respect to the use of M&TE. Records of analysis include instrument and sample identifiers along with references to SOPs, and notes of any deviations. This information is routinely recorded in log books for the project. Control of M&TE equipment is accomplished by limiting access to the laboratories through the Center's security system as described below. Additional measures to prevent unauthorized access to M&TE equipment include password protection on computers that control the instruments.

The CEMRC is housed in Light Hall, a 26,000 ft^2 laboratory and office facility adjacent to the NMSU-Carlsbad campus. All activities for the VOCMP/HMMP are conducted in laboratories where the security and integrity of the equipment, samples and data can be controlled. Light Hall was built with an advanced system of electronic locks and key-card access, and the OC laboratories are kept locked at all times, limiting access to authorized personnel. The building is a closed building and all visitors are escorted. Samples, chemical reagents, standards, etc. are securely stored in the locked laboratories.

The laboratories are cleaned regularly, with precautions taken to minimize the possibilities of contaminating samples. Temperature is monitored and recorded in a logbook in the room used for sample storage. Fluorescent lighting from the ceiling illuminates the bench tops and meets applicable OSHA standards. Provisions for the proper storage and disposal of chemical wastes are coordinated through the CEMRC Chemical Hygiene Officer.

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10.0 HANDLING, STORAGE, AND SHIPPING

The CEMRC OC laboratory will maintain the VOCMP/HMMP samples under chain-of-custody and will account for all samples and supporting records of sample handling, preparation, analysis, and disposal. The CEMRC OC laboratory shall maintain sample identity, sample custody, and all sample- and analysis-related records according to the requirements in CP-PROC-001, "Sample Tracking and Chain of Custody." These requirements are designed to ensure the following:

- The custody, traceability and integrity of samples are maintained while in the possession of the OC laboratory
- All sample-related activities (e.g. receipt, storage, preparation, analysis, reporting, and disposal) are recorded in notebooks or through some other acceptable means
- All laboratory records for each specified sample group will be accounted for when the project is completed
- All laboratory records directly related to VOCMP/HMMP samples are assembled and either delivered or made available to WTS

The OC laboratory shall implement the following requirements for sample receiving, sample identification, sample security, sample storage, and sample tracking to ensure accountability of VOCMP/HMMP sample chain-of-custody as well as control of all WTS sample-related records.

Sample Receiving. VOCMP/HMMP samples are received by the OC laboratory in accordance with the specifications in the client SOW and in the project related SOPs. The OC laboratory will make available a sample custodian for receiving VOCMP/HMMP samples. At the time of sample receipt, OC personnel shall sign and date any accompanying chain of custody or related forms in indelible ink to acknowledge receipt and accept custody.

Upon receipt, the condition of shipping containers and sample containers shall be inspected and any problems recorded on a sample receiving and tracking log by the sample custodian or his/her designee. The OC laboratory sample custodian shall contact the appropriate WTS contact person to resolve any problems or discrepancies including but not limited to, absent documents, conflicting information, absent or broken custody seals, and unsatisfactory sample condition (e.g., leaking sample container, improper sample preservation). The OC laboratory shall record the resolution of problems and discrepancies by WTS. All sample discrepancies and their resolutions shall be recorded on a Sample Discrepancy Form, which shall be included in the batch data package.

Sample Identification. The OC laboratory shall maintain the identity of VOCMP/HMMP samples and any prepared samples throughout the laboratory. Each sample and sample preparation container shall be uniquely labeled with this information, which also is recorded in the sample receiving and tracking log.

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Sample Security. The OC laboratory shall designate and control storage areas for VOCMP/HMMP samples and any related samples. Requirements for monitoring of storage conditions and security specified in the SOW shall be appropriately documented.

Sample Tracking. The OC laboratory shall record all activities performed on VOCMP/HMMP canisters and samples including, but not limited to, canister cleaning, canister certification, sample receipt, sample storage, sample preparation, and sample analysis. Documentation of sample tracking activities shall follow the procedures listed in Table 2.

11.0 CONTROL OF NONCONFIRMING ITEMS

"Nonconformances and Non-Routine Events" are the subject of CP-PROC-012. Records of instrument performance are maintained in instrument logs, and nonconforming items are tagged "Out of Service" as specified in the SOPs. Additional information on this issue may be found in Section 6.5, Identification and Control of Items, of the CP-QAP. As noted in the CP-QAP, nonconforming items shall be managed to prevent their inadvertent use and to prevent their recurrence.

12.0 CORRECTIVE ACTION

QC failures require implementation of the corrective actions specified in CP-PROC-012, "Nonconformances and Non-Routine Events" unless other criteria or considerations apply. For example, the analyst may believe that the standard corrective actions should not or cannot apply because the sample has been consumed or has in some way been compromised. In such cases, the laboratory manager must review and approve the data before they are forwarded for reporting. If non-conforming data are reported under such circumstances, a narrative explaining the reasons for not following the standard corrective actions must accompany the data.

13.0 QUALITY ASSURANCE RECORDS

QC samples and performance check standards are routinely utilized to ensure that analytical methods and equipment performance are within pre-established limits. The OC group has implemented a series of QC procedures to verify that the quality of measurements comply with specified performance requirements. The procedures include the use of reference standards, calibration and performance checks of measurement systems, computational checks, evaluation of quality control data, review of results, and verification and validation of data. Information on standards, calibrations, replicate analyses, laboratory performance blank(s), laboratory fortified blank(s), laboratory fortified sample matrix spike(s) duplicate analyses as defined in the SOPs will be incorporated in the hard copy data deliverables as required in the WTS SOW.

CEMRC has instituted a document control and records management system to ensure that documents and records are legible, retrievable, traceable to the phase of work when they were recorded, and defensible. CEMRC procedures for management of Quality Assurance Records are specified in the CP-QAP-004, Section 11.3, "Data Documentation, Control, and Validation", and

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in CP-PROC-013, "Records Maintenance, Storage, and Establishment of a Filing System" and DC-PROG-001, "Document Control Program".

14.0 TRAINING

Training is part of the routine operations at CEMRC, and mechanisms have been established to organize training and to maintain training records. Descriptions of the CEMRC training requirements can be found in CP-PROC-011, "Training and Qualification".

15.0 REFERENCES

EPA Method TO-15 "Determination of Volatile Organic Compounds (VOC) in Air Collected in Specially–Prepared Canisters and Analyzed By Gas Chromatography/Mass Spectrometry (GC/MS)", *Compendium of Methods for the Determination of Toxic Organic Compounds in Ambient Air*, Second Edition, January 1999.

"Volatile Organics Analysis of Ambient Air in Canisters", USEPA Contract Laboratory Program, VCAAO 1.0, Publication 9240.1-18, EPA/540/r/085, PB95-963524, December 1994.

EPA Method 8260B "Volatile Organic Compounds by Gas Chromatography/Mass Spectrometry (GC/MS)", *Test Methods for Evaluating Solid Waste, Physical/Chemical Methods*, SW-846, Third Edition.

16.0 REVISION HISTORY

Revision #	Date	Description
0	11/20/02	Initial Document, released as EC-PLAN-026.
1	02/01/06	Review and revision to reflect transfer of analysis responsibility to the Organic Chemistry group; Document released as OC-PLAN-001.
2	04/18/08	Review and revision to incorporate requirements of current SOW, including hydrogen and methane analysis.
3	05/05/10	Review and revision to incorporate requirements of current SOW; title of plan was revised.

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CARLSBAD ENVIRONMENTAL MONITORING & RESEARCH CENTER (CEMRC)

PREPARATION OF CANISTERS AND SAMPLE TRAINS FOR AMBIENT AIR SAMPLING

OC-PROC-002-005-021506- _ _ _

Effective Date: September 8, 2011

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PREPARATION OF CANISTERS AND SAMPLE TRAINS FOR AMBIENT AIR SAMPLING

1.0 GENERAL DISCUSSION

1.1 Purpose of Procedure

Prior to the sampling of volatile organics in ambient air into passivated canisters, the canisters must be certified to be leak free and conducive to the stability/recovery of compounds of interest. Canisters must be cleaned using a cleaning process certified as effective in cleaning canisters. Traces of volatile organics left in canisters from previous sampling events can lead to erroneously high concentrations of volatile components in subsequent samples. All canisters must be certified as leak-free before use for sampling or standard preparation. A pressurized leak test will be used to certify sample canisters as leak-free. Canister leaks or the retention of target compounds on internal surfaces can lead to erroneously low values for volatile components.

Any new canisters not certified by the manufacturer may also be certified as passive to target compounds held within the container over a one-week period. This is accomplished by preparing a spiked sample in the canister and analyzing once on the day of preparation and again seven days later. The stability/recovery of the spiked sample is determined by comparing the two analysis results. Canisters are proven to be inert to target compounds when the stability/recovery of the spiked sample has not significantly changed over the one-week time period.

The cleaning process for canisters involves a series of pressurization/evacuation cycles, during which the canisters can be heated. Following cleaning, a canister is certified as clean by analyzing an aliquot of ultra high purity nitrogen from the canister for VOC, hydrogen, and methane. The certification blank analysis is used to determine whether significant levels of analytes are present in the canister. Canisters cleaned with a cleaning process that is proven effective that have been certified as leak-free and passive to target compounds are then evacuated to below 50 mtorr to prepare them for use/storage.

Canisters that have undergone recent maintenance (such as valve changes, etc.) must be re-certified as clean and leak free. Such canisters may require testing to determine the stability/recovery of target compounds.

Prior to use of passivated sample train equipment in collection of field samples, sample trains must be cleaned and certified. Sample trains are cleaned by a flush of humidified, UHP nitrogen while the sample trains are heated. Following cleaning, one sample train per cleaning batch must be certified as clean. One train per batch is selected for collection of a certification sample, in which UHP nitrogen is pulled into a clean canister (usually 0.4L canister is used) through the sample train. The sample must be analyzed by GC/MSD and levels of each target VOC must be lower than 0.2 ppbv.

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1.2 Measurement Principle

System pressure may be monitored by use of a 0-50 psia pressure transducer attached to the system controller manifold. Typical canister pressurization during cleaning cycles is 25 to 30 psia. The output of the transducer may be monitored through the system controller software. System vacuum may be monitored by use of a Pirani gauge attached to the system controller manifold. The output of this gauge is monitored through the system controller software. Typical evacuation during operation of the molecular drag-pump is 10 to 50 mtorr.

Canister cleanliness is determined through the measurement of the concentrations of compounds of interest in a blank nitrogen sample via gas chromatography with mass spectrometry for VOC and thermal conductivity detection for hydrogen and methane. One canister from each cleaning batch may be analyzed to demonstrate batch cleanliness.

1.3 Measurement Interferences and Their Minimization

Interferences can be introduced through improperly cleaned and certified canisters being used either for preparation of calibration standards or for sample collection. These interferences can be minimized through careful performance of the written procedures presented in this document. Interferences may also be introduced by contamination of the internal standard with a compound of interest.

1.4 Personnel Responsibilities

Laboratory personnel should verify that this procedure is the latest available revision by checking it against the version in H:\document control\oc. The latest revision of this document shall be in the VOC laboratory at all times. If this procedure is found to be unsatisfactory in any way and a procedure revision is required, notify the Lab Manager or designee.

1.5 Definitions

Blank Sample: An aliquot from a cleaned canister that has been filled with ultra-high purity nitrogen or zero air to a pressure of ~15 psig.

Canister: A stainless steel pressure vessel of a desired volume (e.g. 6 L or 0.4 L) with an interior surface passivated by a specialized polishing and/or coating process.

Contract-Required Quantitation Limit (CRQL): The lowest concentration of a target analyte, which will be reported under a given contract. The CRQL for each target analyte is defined in the contract Statement of Work and is the same as the Method Reporting Limit.

Gas Chromatography (GC): An analytical technique used for separating compounds of interest.

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Thermal Conductivity Detector (TCD) - An analytical detector commonly used with GC that senses changes in the thermal conductivity of a gas stream containing analytes in relation to a reference gas

Mass Selective Detector (MSD) or Mass Spectrometer (MS): An analytical technique used for quantitative analysis and/or for the qualitative determination of the molecular composition of a sample. An MS consists of ionization of compounds to generate charged molecules, separation of ions based on their mass-to-charge ratio (*m/z*), and detection of the charge induced or current produced when ions interact with the detector's surface.

Method Detection Limit (MDL): The minimum concentration of a target analyte that can be identified, measured, and reported with 99% confidence that the analyte concentration is greater than zero.

Method Reporting Limit (MRL): The lowest concentration of a target analyte, which will be reported under a given contract. The MRL for each target analyte is defined in the contract SOW and is the same as the CRQL.

Millitorr (mtorr): pressure measured in thousandths of a millimeter of Hg, usually measured relative to absolute zero pressure.

Organic Chemistry (OC): The CEMRC Organic Chemistry department.

Percent Difference (%D): A method of comparing two values to one another in which the difference between an experimentally-determined value and a known value is divided by the known value and expressed as a percent.

Ppbv: parts per billion volume.

Psia: pounds per square inch of pressure, measured relative to absolute zero pressure.

Psig: pounds per square inch gauge pressure, measured relative to local barometric pressure.

Revolutions per minute (RPM): A unit of measure used by the high vacuum or molecular drag pump in the Entech Canister Cleaning system.

Standard cubic centimeters per minute (sccm): a measure of gas flow rate.

Sample Train: A passivated air sampling apparatus used in the collection of ambient air samples.

Spiked Sample: An aliquot from a canister spiked with a gas calibration-standard containing target compounds of a known concentration and pressurized with humid

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ultra-high purity nitrogen or zero air so that the final nominal level of each target compound is ~10 ppbv.

Statement of Work (SOW) – A written document describing the agreement of the client and contracted laboratory. The SOW includes, but is not limited to, technical tasks, quality assurance requirements, data reports and deliverables, and other considerations.

Target Compounds: The compounds of interest as determined by the applicable SOW or contract.

Ultra High Purity (UHP): A grade of reagent gas certified as free of impurities.

Volatile Organic Compounds (VOC): An organic compound that has a high enough vapor pressure to be in the gas phase at ambient temperature and pressure.

Waste Isolation Pilot Plant (WIPP): A deep geological repository for transuranic waste outside of Carlsbad, NM.

Volatile Organic Compound Monitoring Program (VOCMP): A WIPP monitoring program designed to determine the concentrations of VOC in air inside and emitted from the underground repository.

1.6 Safety

Proper personal protective equipment shall be worn while performing this procedure. Samples and standards analyzed in this procedure may contain compounds which present a health hazard. Vent pressurized canisters in a snake vent or fume hood to avoid inhalation of the contents. During the cleaning cycle, caution should be used to avoid touching heated canisters. Use caution when handling tanks of compressed gas. Tanks should always be moved with caps in place. Eating or drinking in the laboratory is strictly prohibited.

2.0 APPARATUS, INSTRUMENT, REAGENTS, AND FORMS

2.1 Instrumentation and Apparatus

2.1.1 Canister Cleaning System Description

The Entech Model 3100 Canister Cleaning System comes equipped with a universal device backplane which provides a passive connection between the SmartLab® I/O Card device software installed in the instrument control computer and the Universal SmartLab® Control Card (UNICARD), which controls the valves, and other control devices within the system. Cleaning cycle parameters are input through the SmartLab® Instrument Control Software.

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A diaphragm pump acts as a roughing pump to obtain a rough vacuum. The molecular drag-pump serves as the high-vacuum pump and will engage when the roughing pump brings the system down to a vacuum of 1-2 psia or less. A stainless steel manifold with multiple canister attachment positions allows simultaneous cleaning of several canisters per cleaning batch.

The cleaning system is an oven-based cleaning system. In this system, the cleaning manifold is placed in an oven, which heats the canisters during cleaning. There are two types of ovens. The smaller oven contains manifolds and connections for cleaning up to twenty mini-canisters with 0.4 L capacity. A larger oven with six connections is available for six cleaning canisters with 6 L capacity. The larger oven shall be reserved for cleaning canisters used only for ambient level sampling. The advantage of using the oven-based canister cleaning system is that more even heating of the canisters is possible, which promotes better cleaning of heavier organic compounds. The oven based cleaning system employs separate fill and exhaust lines to effectively eliminate heavy and polar contaminants. For typical cleaning, the oven should be set to 80-100°C. The oven temperature must not exceed the limits of canisters, gauges, or sample trains. For example, if a pressure gauge is attached to the canister, the oven temperature should not exceed 80°C during cleaning.

A diagram of the cleaning system is shown below. Note that the canisters and cleaning manifold are located inside the oven.

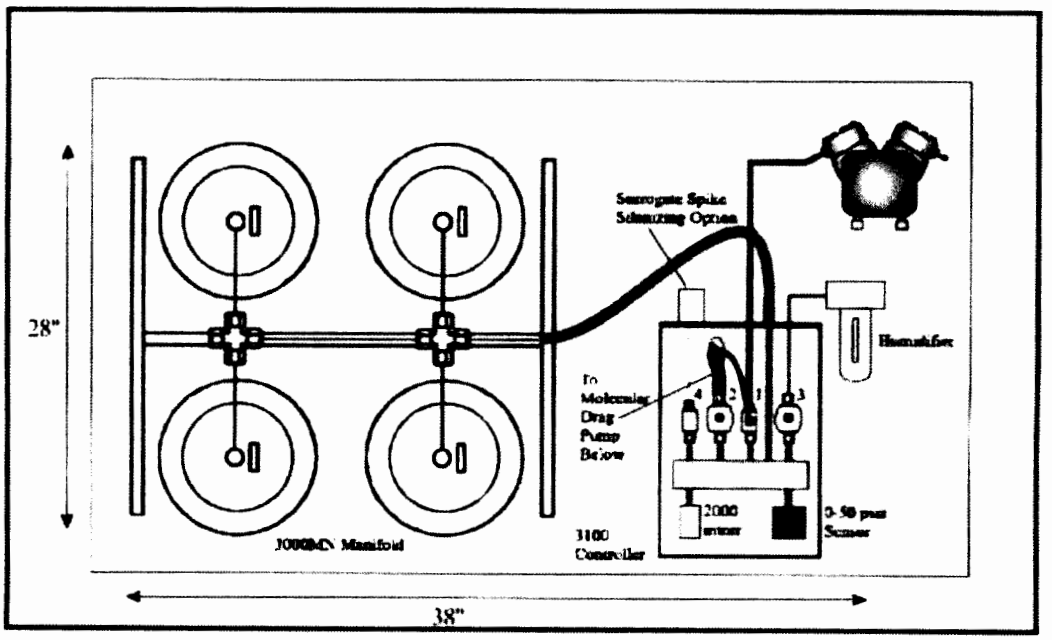

Figure 1 ***Top View 3100***

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2.1.2 Description of GC/MSD Analytical System

An Agilent 6890/5973 or 6890/5975 Gas Chromatograph/Mass Spectrometer may be used to analyze the blank and spiked samples for the certification processes. The systems consist of a sample introduction system, a gas chromatograph, an ion source, and a quadrupole mass spectrometer. For ambient level canisters, an Entech 7100 Preconcentrator is used as the sample introduction system. A detailed description of this system and its operation may be found in OC-PROC-003.

2.1.3 Description of GC/TCD Analytical System

The Agilent 6890N GC/TCD instrument as configured at CEMRC consists of three major components: the gas chromatograph (including the column and oven), the TCD, and the MSD. The MSD portion of the configuration will not be utilized in this procedure. An equivalent GC/TCD may be used to analyze the blank and spiked samples for the certification processes. The Entech 7032A-L MiniCan™ Autosampler can introduce up to 21 samples from 400 mL or 1000 mL MiniCans™ or 3 samples from 6 L canisters into the GC. A detailed description of this system and its operation may be found in OC-PROC-009.

2.1.4 Maintenance of the Cleaning System

The fittings and tubing in the cleaning system will require occasional replacement as leaks develop. Parts of the cleaning system may require occasional cleaning with methanol to remove organic contaminants. The solenoid valves may begin to stick after long use. Perform pump maintenance as described in the pump operator manual. The pump will need to be rebuilt on occasion. Solenoid valve and pump rebuild kits are available from Entech Instruments, Inc.

Check the humidification chamber occasionally during operation of the cleaning system to assure that water levels are maintained according to the specification in the operator's manual. The chamber should be refilled using only ultrapure deionized water.

Maintenance of the canister cleaning system shall be documented in the Canister Cleaning System Maintenance Logbook (See Section 7.1.1).

2.1.5 Sample Train Cleaning System Description

The sample train cleaning system utilizes the Entech 4600 Dynamic Diluter. A pressurized canister of ultra-high purity nitrogen is connected to the Calibration Gas Inlet 1 port at the back of the Dynamic Diluter. The Calibration Gas Out port on the front of the Dynamic Diluter is connected to copper tubing leading into the oven. The gas line is connected to a

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manifold fitted with Swagelok connections that connect to the inlets of the sample trains.

2.2 Reagents and Gases

Ultrapure Water: Deionized water with a specific resistance of 17.8 megaohm/cm or greater, free of ionized impurities, organics, microorganisms and particulate matter larger than 0.2 μm (available from the Barnstead® system located in the Environmental Chemistry Laboratory).

Methanol: Reagent grade, for cleaning parts

Gas Calibration Standards: cylinder(s) containing a known concentration of the compound(s) of interest

Ultra-high purity nitrogen (UHPN): ≥99.999% nitrogen

2.3 Forms and Paperwork

2.3.1 Stability Certification

The stability certification process must be documented using a form similar to the Stability Certification Form (see Appendix A).

2.3.2 Canister Cleaning and Leak Testing

Canister cleaning and leak testing must be documented using a form similar to the Canister Cleaning Batch Form (See Appendix B).

2.3.3 Sample Train Cleaning Batch Cleaning and Certification

Sample train cleaning must be documented using a form similar to the Sample Train Cleaning Batch Form (See Appendix C).

3.0 SAMPLE STABILITY SPIKE STANDARDS

Gas calibration standard mixtures used to prepare canisters for the stability certification shall be purchased ready-made from a reputable supplier and shall come with a certificate of analysis. Standards shall be logged in to the appropriate Standards Logbook and be given a unique standard identification number according to CP-PROC-009, "Standards Log-in and Preparation."

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4.0 PROCEDURE

4.1 Pressurization Leak Test Procedure

All canisters must undergo the pressurization leak test procedure before use for sampling. Follow the canister custody control and storage requirements of OC-PROC-006.

4.1.1 Ensure that all canisters are at room temperature before proceeding.

4.1.2 If the canister cleaning system is not already on, turn it on per section 4.2.

4.1.3 Turn on the tank valve of the diluent/fill gas. Observe the regulator to ensure that there is adequate pressure in the tank. If the high pressure side of the regulator is reading less than 100-200 psi, the tank should be replaced with a full tank prior to pressurizing canisters. The regulator should be set to an exit pressure of around 30-50 psig. Use of a higher pressure may result in inaccurate pressurization of canisters.

4.1.4 Open all canister valves.

4.1.5 Activate the "DILUENT" solenoid valve from the "RUN" screen in the software.

4.1.6 When the pressure reading is around 43 psia (approximately 30 psig), click on the "ALL OFF" button to stop the fill gas and immediately close all canister valves. (Note: Alternatively, individual canisters may be pressurized using the "Pressurize" feature on the Entech Dynamic Diluter.)

4.1.7 Remove the canisters from the cleaning system.

4.1.8 If the canister cleaning system is not needed again, turn it off following steps 4.2.19.1- 4.2.19.3. Close the valve on the fill gas cylinder.

4.1.9 For canisters without an attached pressure gauge, check for leaks in the canister valve by attaching a certified pressure gauge without opening the valve. Note: If a leak exists, the gauge reading will change over time. Canisters with leaking valves shall be tagged "Out of Service." On canisters with an attached gauge, the gauge reading represents the internal canister pressure even with the valve closed.

4.1.10 Document the "Initial Pressure" readings and all other appropriate information required in section 2.3.2. The test pressure should be 30 ± 5 psig.

4.1.11 Close the canister valve tightly. For canisters not having an attached pressure gauge, remove the pressure gauge. Place a "Leak Test in Progress" tag on the canister. Optionally, one canister in the batch may be tagged to show that the batch is being leak tested.

4.1.12 Install a cap on the canister outlet. The valve alone should not be used to preserve sample pressure or vacuum.

4.1.13 Repeat this until the initial pressures have been recorded for all canisters.

4.1.14 After at least 24 hours following the initial pressure reading, attach the pressure gauge to each canister again (if needed) and document the "Final Pressure" readings along with all other appropriate information required in section 2.3.2. If the final pressure indicates a leak (see section 6.1), try to determine the cause and correct it. If the leak cannot be eliminated, the

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canister should be tagged "Out of Service", segregated, and not used for sampling.

4.1.15 Record the leak-test completion date/time and operator performing the leak test on the CEMRC canister tag for each canister. Remove any "Leak Test in Progress" tags. If the canister has not yet been cleaned, proceed with canister cleaning. For previously cleaned canisters, the blank nitrogen within the canister may be used to perform a blank check. Ensure that the CEMRC canister tag also shows the canister serial number.

4.2 Canister Cleaning Procedure

The Entech Canister Cleaning System is set up according to the manufacturer's instructions. The power to the system is usually left off to protect it from power fluctuations unless it is in use. Tubing connectors must be leak tight, but over tightening the fittings may also result in leaks. Ultra-high purity nitrogen should be used as the fill gas. Make certain to install the stainless steel plugs on the ends of the manifold compression connections when not being used to prevent contamination of the system. It is not necessary to cover the ends of the quick-connectors because they incorporate a valve which closes off when not in use.

The system must be turned on following the five steps below in sequential order:

1. Supply power to the roughing pump.
2. Toggle the molecular drag-pump "Start" switch on the front of the controller unit.
3. Wait until the green indicator light comes on under the "=27000 RPM" label near the molecular drag-pump controls.
4. Enter the SmartLab® software on the control computer by clicking on the "3100 CAN CLEANER" icon.
5. Ensure that the "Link Active" indicator lights up on the front of the controller unit.

Prior to cleaning canisters, the SmartLab® Instrument Control Software must be programmed for the correct number and lengths of desired pressure (soak) and evacuation cycles. The Canister Cleaning System may be programmed to allow filling and evacuation based on either a time or pressure basis. Choosing to fill and evacuate based on pressures rather than time can give a more consistent cleaning from batch to batch. In the case of canisters utilized for samples having very high levels of VOC compounds, it may be necessary to use heating of canisters and a greater number of cycles (10-20) to facilitate thorough cleaning. For general cleaning, Entech Instruments, Inc. recommends five cycles of pressurization and evacuation. The general use CEMRC cleaning procedure employs eight such cycles.

4.2.1 In the software, either choose an already existing cleaning method or enter your own parameters for cleaning. For general cleaning of ambient level (ppbv) canisters, use the "CLEAN1" method. A new method can be created depending on the condition of the canisters to be cleaned. In

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general, it is better to clean routine-sampling canisters in a separate batch from those suspected of having higher concentrations of VOC.

4.2.2 Before attaching canisters to the cleaning system, allow them to vent to atmospheric pressure in a fume hood or vent snorkel if the pressure exceeds 20 psig.

4.2.3 Close the canister valves.

4.2.4 Any flammable tags or cable ties on the canister should be removed prior to cleaning.

4.2.5 Attach each canister to be cleaned to the manifold. For canisters having compression connectors, use care not to strip the nuts. If fewer than the maximum number of canisters are being cleaned simultaneously on a manifold, cap off all unused compression connector ports with a stainless steel cap and tighten securely. Quick-connectors contain a valve which is normally closed when not in use, so it is not necessary to cap them off.

4.2.6 From the SmartLab® Instrument Control Software main menu at the control computer, enter the "RUN" screen.

4.2.7 Click on the button next to the roughing pump and wait until the pressure reads 1.0-2.0 psia or less.

4.2.8 Click on the button next to the high vacuum pump (i.e. molecular drag-pump) and wait 2-5 minutes for the vacuum readout to drop below 100 mtorr. If the vacuum readout does not drop below 100 mtorr, a leak exists in the canister connection or through the canister valve. Ensure that the canister valves are closed. Tighten each canister fitting one by one, if necessary, while monitoring the vacuum. Observe which one has the greatest effect on the vacuum reading. If the system uses quick-connect fittings, make certain that each fitting is properly seated. It may be necessary to remove the canister from the fitting, then reinstall it to obtain a good seal. If a quick-connect fitting is found to be leaking, it should be replaced, using a graphite-Vespel ferrule. Do not use stainless steel ferrules on the fused silica lined manifold connections.

4.2.9 Once the system is leak tight, i.e., the pressure is below 100 mtorr, click on the "ALL OFF" button.

4.2.10 Turn on the tank valve of the diluent/fill gas. Observe the regulator to ensure that there is adequate pressure in the tank. If the high pressure side of the regulator is reading less than 100-200 psi, the tank may not have sufficient pressure present to complete the cleaning cycle. The regulator should be set to an exit pressure of around 30 to 50 psig, depending on the number and sizes of canisters being cleaned.

4.2.11 Open the canister valves.

4.2.12 Click on "GO" and automated cleaning will commence.

4.2.13 Make certain that the oven power is on and the temperature is set to 80-100°C. *Note: If a pressure gauge is attached to the canister, the oven temperature should not exceed 80°C during cleaning.*

4.2.14 After the completion of the cleaning program and when the canisters reach a final vacuum below 50 mtorr, close all canister valves, using oven mitts to prevent burns. If canisters read a final vacuum below 50 mtorr, this is indicative that the canisters are leak free. This is a sufficient leak check for CEMRC canisters used in making standards.

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4.2.15 Turn off power to the oven and leave the oven door open to facilitate cooling of the canisters. A fan may be used to speed up the cooling process.

4.2.16 Record the appropriate information required in section 2.3.2.

4.2.17 Document the cleaning date/time and operator performing the cleaning of the canister on the CEMRC canister tag.

4.2.18 Remove the canisters and attach the canister tags. Ensure that the CEMRC canister tag also shows the canister serial number. Store the cleaned canisters until they are delivered to the client, conforming to the canister custody control and storage requirements of OC-PROC-006.

4.2.19 If the canister cleaning system is not needed again, turn it off. Close the valve on the fill gas regulator. The system must be turned off following the four steps below (4.2.19.1 to 4.2.19.3) in sequential order.

4.2.19.1 Turn off the cleaning system by first exiting the software. Use the "EXIT" button in the software, not the "close window" button, to close the software.

4.2.19.2 When the "Link Active" light on the front of the cleaning system goes off, toggle the molecular drag-pump "Stop" switch on the front of the controller unit. The green light described in section 4.2, item #3, should go off after a few minutes.

4.2.19.3 Turn off the power switch on top of the roughing pump.

4.3 Cleaning Certification Procedure

The effectiveness of the cleaning procedure is demonstrated by analyzing a blank sample from a cleaned canister using the GC/MSD for VOCs and GC/TCD for Hydrogen & Methane. The procedure is proven effective when no target VOCs are present at a level equal to or higher than 0.20 ppbv for VOCs certification, while detection of Hydrogen & Methane must be at a level equal to or less than MDL for their certification. For ambient level canisters, the total concentration of identified target and non-target compounds present may not exceed 10ppbv (excluding internal standards and tuning compounds) for the canister to be considered clean. One canister from each batch may be used to perform a batch cleaning certification.

4.3.1 If the canister is not already pressurized, do so by following steps 4.1.1-4.1.8 of Section 4.1, substituting 30 psia for the 43 psia in step 4.1.6.

4.3.2 Attach a calibrated pressure gauge to the canister and document the appropriate information required in section 2.3.2. The preconcentrator pressure gauge reading in the "Smart Report" may also be used to read the pressure in the canister.

4.3.3 Place a tag on the canister to show that it is being used to test cleaning effectiveness. Optionally, one tag may be placed on a single canister from a batch to indicate that the batch is being tested.

4.3.4 Analyze each blank sample with the GC/MSD analysis system by the same procedure used to analyze samples, described in OC-PROC-003.

4.3.5 Analyze each blank sample with the GC/TCD analysis system by the same procedure used to analyze samples, described in OC-PROC-009.

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4.3.6 After analysis results have shown the canister(s) to be sufficiently cleaned, open the canister valve(s) to release the pressure to just above atmospheric conditions.

4.3.7 Remove any tags placed on the canister(s) in step 4.3.3 and evacuate the canister as described in section 4.4.

4.5 Canister Evacuation Procedure

4.5.1 Ensure that all canisters are at room temperature before proceeding.

4.5.2 If the canister cleaning system is not already on, turn it on per section 4.2.

4.5.3 Open all canister valves to release any pressure.

4.5.4 Install the canisters on the cleaning system.

4.5.5 In the software, either choose an already existing evacuation method or enter your own parameters for evacuation. For general evacuation of ambient level (ppbv) canisters, use the "Evacuation1" method. A new method can be created depending on the condition of the canisters to be evacuated. (Alternatively, canisters may be evacuated manually: From the "RUN" screen, turn on the roughing pump. When the system vacuum is below 1-2 psia, turn on the high vacuum pump and allow the pumps to run until the system vacuum stabilizes (must be below 50 mtorr). Record the final system vacuum reading in the appropriate space on the canister cleaning form).

4.5.6 Close all canister valves while the high vacuum pump is still running. Make sure the high vacuum pump is then turned off before removing canisters from the manifold, to avoid damage to the equipment.

4.5.7 Document the evacuation date/time, and the operator performing the evacuation on the CEMRC canister tag for each canister.

4.5.8 Conform to the canister custody control and storage requirements of OC-PROC-006.

4.5.9 If the canister cleaning system is not needed again, turn it off per steps 4.2.19.1 to 4.2.19.3.

4.6 Stability Certification Procedure

Certification shall be performed on each new ambient level canister received by CEMRC, unless manufacturer documentation is provided to show that the canister is certified. The spiked sample should be prepared so that each target compound is at a concentration level of approximately 10ppbv. It is not necessary that the spiked sample is prepared exactly the same for each canister. What is important is that the final results compare to the initial results after the sample is held within the container for seven days, allowing for a 30% deviation.

4.6.1 Evacuate a clean canister as described in Section 4.4.

4.6.2 Place a tag on the canister to show that it is being used to test sample stability.

4.6.3 Prepare a spiked sample at the 10 ppbv level according to OC-PROC-004. The gas calibration-standard must meet the requirements of Section 3.0.

4.6.4 Record the appropriate information required in section 2.3.1.

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4.6.5 Repeat steps 4.5.1 to 4.5.4 for each canister requiring stability certification.

4.6.6 Analyze each spiked sample with the GC/MSD analysis system by the same procedure used to analyze samples.

4.6.7 Document the results and other appropriate information required in section 2.3.1.

4.6.8 Securely close the valve on each canister.

4.6.9 Install the brass caps on all canister outlets.

4.6.10 Allow the canisters to sit for seven days while conforming to the canister custody control and storage requirements of OC-PROC-006.

4.6.11 At the end of seven days, reanalyze the spiked samples and document the results and other appropriate information required in section 2.3.1.

4.6.12 Calculate the relative percent difference between the initial and final results. If it is less than ± 30% for all compounds of interest, the canister is certified as inert to target compounds. If not, see section 6.3.

4.6.13 In a fume hood or snake vent, open each canister valve to release the spiked sample.

4.6.14 Remove the tag placed on the canister in step 4.5.2 and place a tag on the canister to show that it needs cleaning. Cleaning certification should be done after cleaning to show that no traces of the spiked sample remain.

4.6.15 Document the stability certification date on the canister.

4.6.16 Complete and file the documentation required in section 2.3.1.

4.7 Sample Train Cleaning Procedure

4.7.1 Configure the sample train cleaning system as described in section 2.1.4.

4.7.2 Enter the SmartLab® software on the control computer by activating the NT4600 icon.

4.7.3 Set the UHP nitrogen cylinder regulator to 5-8 psi, by decreasing regulator pressure to zero and slowly increasing to desired level while flushing the lines.

4.7.4 Humidify the sample train by adding 100 µL of ultra pure water.

4.7.5 Connect sample trains to flushing ports and tighten fittings with wrenches to ensure a good seal.

4.7.6 Flush sample trains with UHP nitrogen for minimum of 2 hours at approximately 75°C. The general use CEMRC cleaning procedure employs 3 hours at approximately 75°C. Monitor temperature using digital thermometer at top of oven and ensure that a temperature of 80°C is not exceeded.

4.7.7 After completion of flush, allow equipment inside of the oven to cool to room temperature prior to handling. The cooling process may be sped up by using a box fan to blow room air into the oven.

4.8 Sample Train Cleaning Certification Procedure

4.8.1 Set the UHP nitrogen cylinder regulator to 5-10 psi as described in section 4.6.3.

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4.8.2 Connect the UHP nitrogen source to the inlet of the sample train and an evacuated and cleaned 0.4 L canister to the outlet.

4.8.3 Flush air through the sample train into the canister (generally 0.4 L), until the canister pressure is approximately 5-10psig (approximately 20 minutes). Close the valve on the canister. Disconnect the canister from the sample train and manifold.

4.8.4 Place a tag on the canister to show that it is being used to test cleaning effectiveness.

4.8.5 Analyze blank sample for VOCs certification with the GC/MSD analysis system by the same procedure used to analyze samples, OC-PROC-003. The injected sample volume should be 100 mL, which is appropriate for the canister size (0.4L) and suggested pressure.

4.8.6 After demonstrating that a batch of sample trains has been sufficiently cleaned, they may be returned to the client.

4.8.7 Tags may be removed from the canister used in blank testing and the canister must undergo canister cleaning as described in section 4.2 before reusing for certification.

5.0 QUANTIFICATION

5.1 Analyte Quantification by GC/MSD

For ambient level canisters, the GC/MSD must be calibrated according to the calibration procedure in OC-PROC-003 prior to analysis of blank or spiked samples associated with the canister cleaning or certification processes. Analyte concentrations are calculated by the instrument software using response factors normalized to internal standards.

5.2 Analyte Quantification by GC/TCD (as applicable)

For ambient level canisters, the GC/TCD must be calibrated according to the calibration procedure in OC-PROC-009 prior to analysis of blank or spiked samples associated with the canister cleaning or certification processes. Analyte concentrations are calculated by the instrument software using the appropriate calibration method file. Detailed description can be found in OC-PROC-009.

5.3 Calculations

The relative percent difference calculation for the stability determination is performed by the analyst using analyte concentration results.

5.3.1 Sample Stability Calculation

For the spiked sample analysis, the relative percent difference is calculated between peak response of the target analyte on the initial day of preparation and that on the analysis following the seven-day holding time, using the following equation:

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$$\text{Percent Difference (\%D)} = \left[\frac{(C_0 - C_7)}{C_0}\right] \times 100$$

Where:
C_0 = concentration of analyte at initial analysis
C_7 = concentration of analyte after 7 day holding time

6.0 QUALITY CONTROL

To minimize interferences that may influence quantitation of analytes-of-interest, all canisters must be certified to be leak free and may be tested for the stability/recovery of target compounds. Also, canisters must be cleaned using a cleaning process certified as effective. Any canister incapable of passing the certification process will be evaluated to determine the cause of the failure, the problem corrected (if possible), and passed through the certification process again. Any canister failing certification for a second time will be tagged "Out of Service" and will not be used for sampling or standard preparation. If the canister is new, it shall be returned to the supplier for replacement. If the canister is not a new one and the cause of certification failure cannot be determined and corrected, the canister must be discarded. Canisters may require periodic re-certification as determined by the Laboratory Manager or designee if any unusual sample or analysis conditions exist which may indicate the need for re-certification.

Table 1
Canister Testing Requirements Summary

Test	Ambient Level Sampling Canisters
Leak Test	Required prior to use and after each cleaning.
Cleanliness Test (Blank Check)	Required after cleaning for one canister in a cleaning batch or as specified in the client's contract.
Stability Test (Spiked Blank Stability)	Required on all new CEMRC canisters. Required in case of canister malfunction or repair.

6.1 Leak Test

During the pressurized leak test, the difference between the initial and final pressures recorded for the canister should not vary by more than ±2.0 psig. Any significant changes over a 24-hour period of pressurization may indicate that compounds are off-gassing from the canister walls or that the valve and weld seams are not leak tight. Either situation must be corrected and the canister shown to be leak tight prior to use. Any canisters that repeatedly fail the leak test must be set aside and tagged "Out of Service."

6.2 Cleanliness Certification

No target compound shall be present above its MDL for hydrogen and methane and ≥ 0.20 ppbv for all target VOC compounds for any canister. For ambient level canisters, the total concentration of identified target and non-target compounds

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present, excluding internal standards and tuning compounds shall not exceed 10 ppbv for a cleaned canister to be considered "clean". If analysis results of a blank sample show that these limits are exceeded, the canister must be put through another cleaning cycle before analyzing another blank sample. If it fails again, it must be set aside and tagged "Out of Service."

6.3 Stability Certification

The stability certification is intended only for canisters used in ambient level testing. It is not required by the WIPP permit, but should be performed on CEMRC canisters purchased for use in preparation of calibration standards. At ppbv concentration levels, it is important to show that canisters do not possess active sites which may adsorb significant amounts of compounds of interest. Adsorption on active sites can lead to lower recovery of certain compounds over time.

For the stability certification, the acceptable relative percent difference for any target compound at a nominal 10-ppbv concentration in humidified air or ultrapure nitrogen is ± 30%. A greater percent difference indicates a problem with the canister (e.g. leaky seams or active sites on the canister interior). Any problem with sample recovery or stability must be resolved, after which the canister shall be cleaned again and undergo a second stability certification. If, after a second analysis, the percent difference is still unacceptable, the canister must be set aside and tagged "Out of Service".

The stability certification should be done before use of the canister for the first time and is not repeated unless unusual sample or analysis conditions exist which indicate the need for re-certification.

7.0 QUALITY ASSURANCE

7.1 Documentation

Stability certification, leak testing, and cleaning activities shall be properly documented and maintained. See appendices for examples of forms that may be used to document these activities.

7.1.1 Cleaning System Maintenance Logbook

A Cleaning System Maintenance Logbook shall be maintained which shall include records of all cleaning system maintenance, such as parts cleaning or replacements, malfunctions and their resolutions, and any routine maintenance performed. The maintenance log shall include the operator's initials or signature, date maintenance was performed, and a detailed description of what was done.

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7.1.2 Stability Certification Forms

Forms documenting the stability certification process are completed and filed for all canisters used by the CEMRC for either sampling or preparation of standards at ppbv concentration levels. This form documents the preparation of spiked samples for determining if samples were prepared accurately and consistently. It also documents the analysis results of the spiked samples. No new canister should be used prior to performance of the stability certification procedure unless the canister is documented as certified from the vendor or provider. If vendor documentation is used (such as Entech canister certification tags) in place of CEMRC certification documents, the vendor documentation must be kept.

7.1.3 Canister Cleaning Batch Forms

A canister may not be used for preparation of standards or provided to the client for sampling without proper documentation of cleaning and leak testing activities. Copies of Canister Cleaning Batch forms shall be kept with their respective canister cleaning data packages.

7.1.4 Client Canister Cleaning Data Package

For samples cleaned under the WIPP VOCMP, a data package will be provided to the client along with the cleaned canisters. The canister cleaning data package will include the canister cleaning batch forms and a summary of cleaning results, and may include the instrument results of the cleaning blank runs, sample receiving and tracking form, and removal from service forms, if applicable. Spiked blank recovery data may be included with data packages where canister certification is requested. Canisters will be packaged in boxes or containers for return to the client. Tamper-resistant labels may be used to seal shipping containers, where applicable. Cleaned canisters, a chain-of-custody form, and cleaning documentation may be picked up by client representative at the time of delivery of new samples. Alternately, they may be delivered by a CEMRC employee at the client's request. Recent copies of chain-of-custody forms for cleaned canisters returned to the client shall be maintained in the VOC Chain-of-Custody Logbook until archived.

7.1.5 Sample Train Cleaning Batch Forms

A sample train may not be returned to the client for use in sampling without proper documentation of cleaning activities. Copies of Sample Train Cleaning Batch forms shall be kept with their respective sample train cleaning data packages.

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7.1.6 Client Sample Train Cleaning Data Package

For sample trains cleaned under the WIPP VOCMP, a data package will be provided to the client along with the cleaned canisters. The sample train cleaning data package will include the sample train cleaning batch forms, a summary of the cleaning results, and the instrument results of the cleaning blank runs. Sample trains will be returned to the client in the packaging they were delivered in, if applicable, with the cleaning documentation.

8.0 REFERENCES

CP-PROC-009, Standards Log-in and Preparation

EPA Method TO-14, "Determination of Volatile Organic Compounds (VOC) in Ambient Air Using Specially Prepared Canisters with Subsequent Analysis by Gas Chromatography", Compendium of Methods for the Determination of Toxic Organic Compounds in Air, Second Edition, January 1999.

EPA Method TO-15 "Determination of Volatile Organic Compounds (VOC) in Air Collected in Specially–Prepared Canisters and Analyzed By Gas Chromatography/Mass Spectrometry (GC/MS)", Compendium of Methods for the Determination of Toxic Organic Compounds in Ambient Air, Second Edition, January 1999.

OC-PROC-003, Gas Chromatography-Mass Spectrometry Analysis of Volatile Organic Compounds (VOC) in Ambient Air from Canisters at PPBV Concentration Levels

OC-PROC-004, Preparation of Calibration Standards in Specially Prepared Canisters for Analysis by Gas Chromatography/Mass Spectrometry.

OC-PROC-006, Receipt, Control, and Storage of Gas Samples in Passivated Canisters

Operator's Manual for the Entech 3100 Canister Cleaning System, Rev. 001, Entech Instruments, Inc., Simi Valley, CA.

US EPA Contract Laboratory Program Volatile Organics Analysis of Ambient Air in Canisters, VCAAO 1.0, 1991.

9.0 RECORDS

Stability Certification Forms
Canister Cleaning Batch Forms
Sample Train Cleaning Batch Forms
Canister Cleaning System Maintenance Logbook
Organic Chemistry Standards Logbook

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10.0 APPENDICES

Appendix A: Stability Certification Form
Appendix B: Canister Cleaning Batch Form
Appendix C: Sample Train Cleaning Batch Form

11.0 REVISION HISTORY

Revision #	Date	Description
0	12/19/03	Initial document, issued as EC-PROC-021.
1	05/25/04	Procedure updated to accommodate headspace gas canister information.
2	02/1/506	Review and revision; transfer responsibility to Organic Chemistry and issued as OC-PROC-002.
3	04/18/08	Review and revision to incorporate contents of temporary deviation and new SOW requirements.
4	05/07/10	Biennial review and revision to incorporate temporary deviations, including sample train cleaning and certification.
5	09/08/11	Revision to incorporate temporary deviations, minor changes.

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Appendix A: STABILITY CERTIFICATION FORM

Waste Isolation Pilot Plant Volatile Organic Compound Monitoring

Client: _____

Canister Number: _____ **Canister Volume:** _____

Date/Time Prepared: _____ **Prepared By:** _____

Standard (for Spiked Blank)

CEMRC Standard ID#:	
Standard concentration (ppbv):	
Standard expiration date:	

Spiked Sample Prep (Dynamic Diluter used)

Standard Volume Added:	
Diluent Volume Added:	
Fill gas used: ultra-high purity nitrogen	Lot#:
Approximate Concentration (ppbv)	

List of target compounds required (* additional requested analytes)

COMPOUND NAME	Conc. Required (ppbv)	First Analysis Results (ppbv)	Second Analysis Results (ppbv)	Relative % Difference
Carbon tetrachloride	~10			
Chlorobenzene	~10			
Chloroform	~10			
1,1 Dichloroethylene	~10			
1,2-Dichloroethane	~10			
Methylene chloride	~10			
1,1,2,2-Tetrachloroethane	~10			
Toluene	~10			
1,1,1-Trichloroethane	~10			
Benzene *	~10			
Trichloroethylene *	~10			
Tetrachloroethylene *	~10			
Chloromethane*	~10			
trans-1,2-Dichloroethylene*	~10			
1,2,4-Trimethylbenzene*	~10			
p,m-Xylene*	~20			
Trichloromonofluoromethane*	~10			

Comments:

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Appendix B: CANISTER CLEANING BATCH FORM

CLEANING, LEAK TEST, CERTIFICATION, AND EVACUATION FORM

Client:_____

Cleaning/Leak Test:

Cleaning Date & Time:_____ **Performed By:** _____

System Used: **Entech 3100** Method: **Clean 1**

No. Of Cycles: 8 **Cycles Heat Used:** 1-8

Gauge Used : _____ **Fill Gas Lot #:** _____

Performed By: _____ **Performed By:** _____

Canister Number	Initial Pressure Reading (psig)	Initial Pressure Date & Time	Final Pressure Reading (psig)	Final Pressure Date & Time

Cleaning Batch Certification:

Pre-Certification Pressure @ ~15 psig? (Initials): _____

Batch-Certification Canister Number: _____

Batch-Certification Injection ID

VOC injection ID:_____ HM injection ID:_____

Canister Pressure (psia): _____ (from Preconcentrator QC Report)

Canister Evacuation:

Evacuation Date & Time:_____ **Performed By:** _____

Evacuating System : Entech 3100 **Final Vacuum Reading (mtorr):** _____

Canisters from Data Package: _____

Comments:_____

_____

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Appendix C: SAMPLE TRAIN CLEANING BATCH FORM

Client: _____

Cleaning:

Cleaning Date & Time: _____ **Performed By:** _____

System Used: Entech 3100

Flush Gas Lot #: _____

Sample Train S/N	High-purity water added (100 µL)

Cleaning Batch Certification:

Batch-Certification Sample Train S/N: _____

Batch-Certification 0.4L Canister S/N: _____

Pre-Certification Pressure > 5 psig? (Initials): _____

Batch-Certification Injection ID: _____

Canister Pressure (psia): _____ **(from Preconcentrator QC Report)**

Comments:

_____

_____

_____

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TEMPORARY PROCEDURAL DEVIATION FORM

Carlsbad Environmental Monitoring & Research Center

Title: Gas Chromatography-Mass Spectrometry Analysis of Volatile Organic Compounds (VOC) in Ambient Air from Canisters at ppbv concentration levels.	
Document No: OC-PROC-003-003-021706-001	Implementing Program: Organic Chemistry

Deviation:
The information included in the deviation here is to verify mainly that all target analytes in lab blanks should be < 0.5 ppbv (contract –specified limit) as described in the Table 2 of SOW on page 19.

Section 1.3: 6th line of the first paragraph on page 3 of 21 read 'An additional blank which contains less than the contract required quantitation limit (CRQL) of any compound of interest will suffice to demonstrate that there is no carryover.....'

It should read as follows: An additional blank which contains less than 0.5 ppbv, of any compound of interest will suffice to demonstrate that there is no carryover.....'

Section 2.1.5.2: Line 7th of the section read 'If the MDL as determined by this method is lower than 0.2 ppbv for any compound, the default value of 0.2 ppbv shall be used as the compound MDL for reporting purposes'.

It should read as follows: Actual MDL will be reported as calculated. MDL's of ≤ 0.5ppbv for target analytes shall be obtained for the analytical system as described in the SOW page 13.

Section 5.4: Third line of the paragraph on page 13 of 21 read. 'The analysis results of the blank must be <CRQL for all target compounds and < 10 ppbv concentration of total VOC for the system to pass the certification.'

It should read as follows: The analysis results of the blank must <0.5 ppbv, for all target compounds and < 10 ppbv concentration of total VOC for the system to pass the certification.'

Section 6.3: Second line of the 6th paragraph on page 17 of 21 read 'The concentration of all compounds of interest must be below CRQL and the total concentration of VOC less than 10 ppbv in order to proceed.'

It should read as follows: 'The concentration of all compounds of interest must be below 0.5 ppbv and the total concentration of VOC less than 10 ppbv in order to proceed.'

Justification: In accordance to the revised statement of work (SOW prepared on January 11, 2011).

Valid for: _Until Next Revision_____

Approved by: _____ Responsible Manager 02/08/2011 Date

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STAFF TRAINING

I have reviewed and I understand the procedure deviation described above.

Print Name	Signature	Date

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TEMPORARY PROCEDURAL DEVIATION FORM

Carlsbad Environmental Monitoring & Research Center

Title: GAS CHROMATOGRAPHY-MASS SPECTROMETRY ANALYSIS OF VOLATILE ORGANIC COMPOUNDS (VOC) IN AMBIENT AIR FROM CANISTERS AT PPBV CONCENTRATION LEVELS	
Document No: OC-PROC-003-003	**Implementing Program: Organic Chemistry**

Deviation:

Add to the end of Section 4.2 the following text: Chemstation will calculate the concentration of analytes in the sample introduced for analysis. Any prior dilutions resulting from off-line static pressure dilutions or injection volume are recorded on the "Sample Preparation Form", where a total dilution factor is determined. The total dilution factor for a sample is entered in the "Results Summary" section of the verified spreadsheet (VOCsDP.xls) and is used to calculate the concentration of analytes in the undiluted sample.

Justification:

In accordance with an NRE (101409 OC 09), this change is made to clarify the calculation and application of dilution factors.

Valid for: Until next revision

Approved by: EKStone — **Responsible Manager** — 5/18/10 — **Date**

STAFF TRAINING

I have reviewed and I understand the procedure deviation described above.

Print Name	Signature	Date

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CARLSBAD ENVIRONMENTAL MONITORING & RESEARCH CENTER (CEMRC)

GAS CHROMATOGRAPHY-MASS SPECTROMETRY ANALYSIS OF VOLATILE ORGANIC COMPOUNDS (VOCS) IN AMBIENT AIR FROM CANISTERS AT PPBV CONCENTRATION LEVELS

OC-PROC-003-003-021706-_ _ _

Effective Date: May 3, 2010

CARLSBAD ENVIRONMENTAL MONITORING & RESEARCH CENTER

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GAS CHROMATOGRAPHY-MASS SPECTROMETRY ANALYSIS OF VOLATILE ORGANIC COMPOUNDS (VOC) IN AMBIENT AIR FROM CANISTERS AT PPBV CONCENTRATION LEVELS

1.0 GENERAL DISCUSSION

1.1 Purpose

The purpose of this procedure is to describe the analysis of VOC in ambient air as outlined in EPA Methods TO-14A and TO-15. The VOC are separated by gas chromatography and identified and quantified by a mass spectrometer. This method is applicable to specific VOC that have been tested and determined to be stable when stored in pressurized and sub-atmospheric pressure canisters. Compounds of interest are defined by the statement of work for a given project. The instrument commonly used for these analyses is a GC/MSD operating in the scan mode. This procedure is intended for use by trained analysts; it is not intended to provide step-by step-instructions on the operation of the instrument. The user is referred to the instrument manuals for detailed operating instructions.

1.2 Measurement Principle

An aliquot of air sample from the sample canister is introduced into the Entech 7100 preconcentrator. Compounds of interest are concentrated and prefocused, while most water vapor and carbon dioxide are removed from the sample. The sample then passes into GC.

The analytes are introduced into a GC column within an oven, via the helium carrier gas. Oven temperature may be either held steady or increased at a specified ramp rate during the run according to the components to be separated in the samples. The GC column is coated with a polysiloxane film. Compounds will interact with the surface of the column based on their chemical and physical properties, namely vapor pressure. The strength of these interactions relates to the time that the compound is retained on the column. With appropriate oven temperature and carrier gas flow settings, controlled by the instrument software, compounds of interest can be separated from each other based on their different retention times.

Upon exiting the GC column, analytes pass through a heated interface and enter an ion source. The ionization technique used by the source is electron ionization, in which 70eV electrons enter the ionization chamber, guided by a magnetic field. The electrons interact with the sample by ionizing molecules and inducing fragmentation. The positive voltage on the repeller pushes the positive ions into the lens stack, where they pass through several electrostatic lenses. These lenses concentrate the ions into a tight beam, which is directed into the mass filter.

The mass analyzer, or mass spectrometer, separates the ions according to their mass-to-charge ratio. The types of mass analyzers in use at CEMRC are quadrupole instruments. The quadrupole mass analyzer is a mass filter constructed of four, parallel conductive, rods of gold-metalized fused-silica (quartz). The four hyperbolic surfaces create the complex electrical fields necessary for mass selection. Only ions of a particular mass-to-charge ratio can pass through the detector at any instant. A combined direct current (dc) and radio

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frequency (RF) signal is applied to the two pairs of rods. The magnitude of the RF voltage determines the mass-to-charge ratio of the ions that pass through the mass filter and reach the detector. The ratio of dc-to-RF voltage determines the resolution (widths of the mass peaks).

During data analysis and quantitation, the detector response to a specified mass-to-charge ratio at a specified retention time is integrated and the area response is used in the concentration calculation. The detector response is normalized to the response of an internal standard, introduced at a known concentration. Normalization of sample responses to the internal standard responses corrects for instrument drift. The normalized analyte response is called a response ratio. The response ratio is used to calculate the concentration ratio of the analyte relative to the internal standard, by interpolation from a multi-point calibration curve developed from calibration standards. Finally, the concentration ratio is converted to a concentration by multiplying by the internal standard concentration.

Compounds are identified based on the spectrum of ions produced during ionization of the component. Relative peak heights of different ions are compared to known spectra in the current National Institute of Standards & Technology (NIST) Mass Spectral Library to determine the probable identity of all tentatively identified compounds. Results of library search values are manually reviewed by the analyst to confirm compound identity.

1.3 Interferences

Contamination of the preconcentrator apparatus or internal standard with compounds of interest can cause erroneous quantitation results. Internal standard mixes shall be tested upon receipt to confirm a lack of contaminants in order to avoid this problem. It also necessary to run a blank or low-level sample for clean-up following any sample or standard showing concentrations greater than 500 ppbv for any compound of interest. The results of clean-up samples shall not be reported. An additional blank which contains less than the contract required quantitation limit (CRQL) of any compound of interest will suffice to demonstrate that there is no carryover following analysis of a higher concentration sample.

In the analysis of compounds by GC/MS, interferences can include compounds having the same GC retention time as a compound of interest coupled with an ion in the mass spectrum in common with the quantitation ion of a compound of interest. In most cases, this interference may be resolved by changing the GC method conditions to chromatographically resolve the compounds of interest from other possible components. In cases where resolution is impractical due to time or other constraints, the interference can be avoided by assuring that the chosen quantitation ion for any compound of interest is an ion not found in the spectra of any co-eluting compounds. Qualifier ions may also be defined in the quantitation database to further identify the integration area associated with the defined analyte. A qualifier ion is not used for quantitation, but is a significant ion from the mass spectrum of the analyte whose abundance relative to the quantitation ion may be used to confirm the presence of an analyte. The use of qualifier ions in setting up the quantitation database can minimize the impacts of interferences.

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1.4 Quantitation Limits, Precision and Accuracy

The lower limit of quantitation for the analysis system in use at the CEMRC is approximately 0.2 ppbv for most compounds of interest analyzed under EPA Method TO-15. The upper limit of calibration is 100 ppbv. For samples with concentrations of analytes exceeding 100 ppbv, the sample will have to be serially diluted, as described in section 6.5.

The accuracy of analysis will be defined and evaluated through the use of continuing calibration checks and laboratory control samples (LCS). Periodic analysis of method performance standards will also indicate accuracy. Based on analysis of continuing calibration checks and method performance samples, a ± 30% accuracy level, based on percent recovery, can be expected. Based on analysis of secondary source standards (LCS), a ± 40% accuracy level can be expected. Instrument precision is to be defined and evaluated by the relative percent difference (RPD) values calculated between duplicate analysis of samples and LCS. The target precision is ≤ 25% RPD, per data quality objectives.

Current MDL will be archived as the Organic Chemistry Data Quality Objectives in the "H:\OC DQOs\" directory. MDLs will be updated yearly, or more frequently if major changes to the analytical system occur.

1.5 Personnel

Laboratory personnel should verify that this procedure is the latest available revision by checking the Organic Chemistry Laboratory Procedure Manual against the version in "H:\document control\oc" directory. The latest revision of this document shall be in the Organic Chemistry laboratory at all times. If this procedure is found to be unsatisfactory in any way and a procedure revision is required, notify the Project Manager or designee.

1.6 Definitions

Bromofluorobenzene (BFB)- Also called p-bromofluorobenzene or 4-bromofluorobenzene, this is a gaseous organic compound commonly used as a tuning standard in GC/MS.

Calibration Blank- A standard composed of ultra high purity nitrogen or zero air used to establish a baseline for the analytical system.

Cryogen- A refrigerant used to obtain very low temperatures in the trap of the preconcentrator system. Liquid nitrogen is used as the cryogen at CEMRC; its boiling point is -195.8°C.

Calibration Standard (CAL)- A specially prepared canister containing a known concentration of each VOC of interest.

Continuing Calibration Standard (CCAL)- A daily calibration check standard at the midpoint of the calibration curve.

Contract Required Quantitation Limit (CRQL)- The lowest concentration of a compound of interest which will be reported under a given contract. The CRQL for each compound of

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interest is defined in the contract Statement of Work.

Dynamic Calibration - Calibration of an analytical system using calibration gas standard concentrations in a form identical or very similar to the samples to be analyzed and by introducing such standards into the inlet of the analytical system in a manner very similar to the normal analytical process.

EPA TO-14(A) and TO-15 - The United States Environmental Protection Agency (EPA) standardized method for analysis of toxic organic compounds in ambient air. EPA TO-14 is the original method, EPA TO-14a is a revised version, and EPA TO-15 is the newest revision with an expanded analyte list.

Gas Chromatography (GC) - An analytical technique used for separating compounds of interest.

Internal Standard (IS) - Analyte(s) of known concentration added to all samples, blanks, and calibration standards in known amounts; these are used to correct for fluctuations in instrument response. The relative responses of the IS signal in the calibration standards relative to the IS signal in the samples indicates the presence of signal enhancement or suppression, or instrument drift.

Laboratory Control Sample (LCS) - A standard containing compounds of interest which has been prepared from a different source than that used to prepare the calibration standard.

Linear Dynamic Range (LDR) - The concentration range where the instrument response is linear.

Mass Spectrometer (MS) or Mass Selective Detector (MSD) - An analytical detector used for quantitative analysis and/or for the qualitative determination of the molecular composition of a sample. A MS consists of ionization of compounds to generate charged molecules, separation of ions based on their mass-to-charge ratio (*m/z*), and detection of the charge induced or current produced when ions interact with the detector's surface.

Method Detection Limit (MDL) - The minimum concentration of a compound that can be identified, measured, and reported with 99% confidence that the compound concentration is greater than zero.

National Institute of Standards and Technology (NIST) - A federal agency that develops and promotes measurement, standards, and technology.

Organic Chemistry (OC) - The CEMRC Organic Chemistry department.

Percent Area Response Change (%ARC) - Percent area response change of the internal standard peak in the sample relative to the mean area response of the same internal standards over the most recent initial calibration.

Percent Relative Standard Deviation (%RSD) - For an initial calibration curve, the standard deviation of the relative response factors at each calibration level over the mean of the

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relative response factors times one hundred. This serves as a measure of the linearity of instrument response over the calibration curve, and should be ≤30% for a five point calibration curve.

Quality Assurance (QA) - The systematic monitoring and evaluation of a project to ensure that standards of quality are being met.

Quality Control (QC) - The process of ensuring a certain level of quality in analytical measurements.

Ppbv - parts per billion by volume, the concentration calculated as the volume of the compound of interest divided by the volume of the dilution gas.

Psia - Pounds per square inch absolute pressure, measured relative to vacuum. The psia unit of measure is not affected by local barometric pressure and is used by the Dynamic Diluter.

Psig - Pounds per square inch gauge pressure, measured relative to local barometric pressure.

Relative Percent Difference (RPD) - A method of comparing two duplicate measurements and a measure of precision. In this calculation, the difference of two measurements is divided by their average and expressed as a percent.

Relative Response Factor (RRF) - The response (peak area or height) of a target analyte to the known concentration of the target analyte. For the purposes of calibration, the RRF is averaged over five calibration points.

Retention Time (RT) - The time from the start of the GC analysis run until the compound of interest reaches the detector.

Tentatively Identified Compound (TIC) - An unknown compound that is tentatively identified by Chemstation Software based on its mass spectrum's similarity to a known compound's mass spectrum. Known compounds' mass spectra are stored in an electronic library stored on the instrument and data analysis computers.

Tuning standard - A canister containing a known concentration of a tuning compound, normally 4-bromofluorobenzene (BFB), that is used to verify the mass calibration and sensitivity of the instrument are within acceptable limits prior to performing an analysis.

Ultra High Purity (UHP) - A grade of reagent gas certified as free of impurities

Volatile Organic Compounds (VOC) - An organic compound that has a high enough vapor pressure to be in the gas phase at ambient temperature and pressure.

1.8 Safety

Safety standards and precautions listed in this section are for environmental safety and

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health awareness, and are not quality affecting requirements. Standards and possibly samples analyzed by this method contain known carcinogens. Exposure to these compounds will be minimal as long as the operating procedure is followed.

The standards and the helium carrier gas used for analysis are stored at high pressure. Care must be taken to ensure the integrity of the cylinders is maintained. Large cylinders must be secured at all times. Protective caps must be in place when moving large gas cylinders.

The liquid nitrogen used as a cryogen for this method poses a frostbite hazard. When properly configured, the system will not expose the operator to liquid nitrogen. Do not touch bare portions of the liquid nitrogen transport line or fittings while liquid nitrogen is flowing in the system. Before installation of the liquid nitrogen on the preconcentrator, ensure that the dewar is at the proper pressure.

The GC injection port, the GC oven, the GC/MSD interface, the ion source, the quadropole mass filter, the high vacuum pump and the foreline pump operate at temperatures high enough to cause serious burns. These parts must never be touched when the MSD is on. The GC/MSD interface is heated by the GC and will remain at high temperature when the MSD is off. It is also well insulated and cools off very slowly.

When replacing pump fluid, use appropriate chemical-resistant gloves and safety glasses.

Eating or drinking in the laboratory is strictly prohibited.

2.0 INSTRUMENT, APPARATUS, REAGENTS, AND FORMS

2.1 Instrumentation and Apparatus

2.1.1 Preconcentrator System and Optional Autosampler

The Entech 7100 Preconcentrator System consists of a control module, a transfer line for internal standard, a transfer line for calibration gas standard, four heated sample transfer lines which carry sample from the sample canisters to the preconcentrator, and a heated transfer line which carries the prefocused sample from the preconcentrator to the GC column. The control module contains various valves and mass flow controllers that control the volume of sample introduced into the preconcentrator and the movement of the carrier gas and sample between the two concentrator traps and the prefocusing trap. Traps are cooled by the introduction of liquid nitrogen and heated by trap and bulkhead heaters. The control of all valves, flow controllers, and heaters is accomplished though the use of three SmartLab controller cards within the preconcentrator. The controller cards are controlled remotely by a desktop computer via a USB cable and the 7100A Software.

An aliquot of the sample canister is introduced into the first trap containing glass beads, held at subambient temperature. A predetermined amount of an internal standard is also added to all standards, blanks, and samples during the first trapping phase. Then the trap temperature is raised to slightly below ambient

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temperature and held while helium is slowly passed through it, quantitatively transferring the compounds of interest to the second trap and leaving behind most of the water vapor. The second trap, which is packed with Tenax, is held at below 0°C during the trapping phase. This temperature allows for trapping of the compounds of interest, while carbon dioxide from the air sample is reduced. The trap temperature is then raised to above 150°C and the compounds are backflushed via helium flow onto the prefocusing trap. This trap consists of an empty silonite-coated tube which is held at below -120°C during the trapping phase. Finally, the prefocusing trap is rapidly heated to quickly inject the prefocused sample onto the GC capillary column.

The Preconcentrator may be used alone to analyze up to four canisters at one time using the four sample inlets. Alternatively, the Entech 7016CA Autosampler may be installed on one of the Preconcentrator inlets. Up to sixteen canisters at a time may be loaded onto the autosampler and run in sequence.

2.1.2 Gas Chromatograph Mass Spectrometer

The Hewlett Packard (Agilent) GC/MSD instrument consists of four major components: the GC (including the column and oven), the GC/MSD interface, the ion source, and the quadrupole MS. Each component can be further divided into several modules.

The gas chromatograph consists of a heated inlet and a chromatography column housed in an oven. The column is a fused silica capillary column coated with a cross-linked polysiloxane film coating.

The GC/MSD interface is a heated conduit into the MSD for the capillary column. One end of the GC/MSD interface passes through the side of the gas chromatograph and extends into the GC oven. The other end of the GC/MSD interface fits into the ion source.

Within the ion source, ions are electronically produced in an ionization chamber by filaments. Then a positively charged repeller forces the positive ions through a series of focusing lenses to produce a condensed beam of ions entering the mass spectrometer detector.

The quadrupole mass spectrometer consists of four parallel conductive rods (quadrupoles) through which current passes to produce complex electrical fields necessary for mass selection. Following separation by mass-to-charge ratio, the ions enter an electron multiplier, which amplifies the signal allowing for maximum sensitivity.

2.1.3 Entech 4600A Dynamic Diluter

The Entech 4600A Dynamic Diluter is capable of diluting samples by static pressure dilution, as described in Section 6.5

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2.1.4 Control Systems

Mass flow controllers within the preconcentrator and GC control all gas flows. Trap temperatures and gas flows within the preconcentrator are controlled by the Entech 7100A Software. GC column temperature and gas flow rates, mass spectrometer operation, and sample data analysis functions are performed by the Chemstation® computer software. Some offline data analysis may occur on personal computers with software dedicated to offline data analysis.

2.1.5 Characterization

2.1.5.1 Initial Establishment of Operating Conditions

The instruments are initially set-up according to the instrument manufacturer's specifications. Additional optimization is performed by OC laboratory staff as deemed necessary. Optimization will consist of analysis of standards for the determination of acceptable peak shape, retention time, interferences, sensitivity, and linearity.

Instrument method files are archived along with raw sample data in electronic files stored in the "envchem" directory on the CEMRC network: "J:\VOC Data Packages\" in the appropriate subfolder according to year and quarter.

2.1.5.2 Method Detection Limits

MDLs are determined as follows. Seven replicate analyses of a low level standard containing compounds of interest at or near the expected detection limit are analyzed and the standard deviation determined for each analyte of interest. The MDL is the value obtained when this standard deviation is multiplied by 3.14 (the "Student's t value" appropriate for a 99% confidence level for six degrees of freedom). This MDL determination method is based on that described in 40 CFR Part 136, Appendix B. If the MDL as determined by this method is lower than 0.2 ppbv for any compound, the default value of 0.2 ppbv shall be used as the compound MDL for reporting purposes. MDLs are determined prior to sample analysis for laboratory commissioning and at least yearly thereafter, or following any major repairs to the instrument or significant changes to the analysis method.

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2.1.6 Maintenance

At a minimum, the routine maintenance schedule provided by the manufacturer will be used. All maintenance will be recorded in the appropriate maintenance logbook for the instrument.

2.2 Gases

- UHP nitrogen gas
- UHP helium gas
- Liquid nitrogen

2.3 Reagents

- VOC Calibration Mixture
- VOC Calibration Verification Mixture (LCS)
- Internal Standards
- Tuning Mixture

2.4 Forms and Paperwork

Analysis logs will be maintained. They will record the samples analyzed, date analyzed, standards used for calibration and instrument verification, instrument methods used, and data file storage information. Any problems encountered during analysis or reruns required will also be noted.

3.0 STANDARDS

An initial multipoint dynamic calibration using five levels of standards is performed on the GC/MS system. For routine operations, a blank and a midpoint calibration standard are used to verify the calibration. For the initial calibration, three calibration standards, generally at 2, 25, and 100 ppbv, are prepared according to OC-PROC-004, Preparation of Calibration Standards in Specially Prepared Canisters for Analysis by Gas Chromatography/Mass Spectrometry. The standards are attached to the calibration gas or sample inlet of the preconcentrator. Different volumes of the standard gases are injected to obtain the five calibration levels.

For sample analyses in the range of 0.2-100 ppbv, the volumes and corresponding standard levels are given in the table below (assuming that the standard sample injection volume is 300 mL):

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Table 1: Calibration Standard Levels

Standard Level (ppbv)	Standard Concentration	Injection Volume (mL)
1	2	150
10	25	120
25	25	300
50	100	150
100	100	300

The IS and tuning standard mid will contain 20-100 ppbv each of bromochloromethane, 1,4-difluorobenzene, chlorobenzene-d5, and 4-bromofluorobenzene. The mix may either be prepared according to OC-PROC-004 or purchased from a certified standard gas supplier. A volume of IS and tuning standard sufficient to result in an effective concentration of 10 ppbv is introduced into the preconcentrator for every sample injection.

4.0 QUANTITATION

4.1 Calibration

The calibration is performed by analysis of five calibration standards levels (as described in section 3.0). The method uses the response factors (ppbv/area count) of the calibration standards to create a calibration curve. As prescribed in EPA Method TO14A and TO-15, the instrument is calibrated initially and whenever the daily response factors change by more than 30% from the initial calibration. The %RSD (i.e. the standard deviation divided by the mean expressed as a percentage) of the response factors for the five calibration standards must be $\leq$ 30% for the curve to be acceptable. If the %RSD is > 30%, recalibration is required. The sample concentrations are calculated using the mean of the relative response factors from the most recent acceptable initial calibration.

The retention time shift for each of the internal standards at each calibration level must be within ± 20 seconds compared to the mean retention time over the initial calibration range for each internal standard, unless otherwise specified.

The %ARC must be calculated at each calibration level for the internal standard used. The %ARC is calculated as the difference between area response of the internal standard at a concentration level and the mean area response of the internal standard over the initial calibration divided by the mean area response of the internal standard over the entire initial calibration. The %ARC at each calibration level must be within ± 40% of the mean area response over the initial calibration range unless otherwise specified.

4.2 Calculations

For ambient air samples the equation below is used to calculate sample concentrations:

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$$X_a = \frac{(A_x \times C_{is} \times DF)}{(A_{is} \times RRF)}$$

Where:

X_a = compound concentration, ppbv;

A_x = area response of the characteristic ion for the compound to be measured;

C_{is}= concentration of the internal standard spiking mixture, ppbv;

DF= sample dilution factor;

A_{is}= area of the characteristic ion response for the internal standard;

RRF= the mean relative response factor from five point initial calibration.

Note that the above equation is valid only if the concentration of internal standard added and the volume of sample or QC standard analyzed is the same for each analysis. For qualitative analysis (e.g., TIC analysis) the above equation is used, however, the RRF used in the calculation will be 1. All qualitative results will be considered estimated values and will be indicated as such on the final report.

4.3 Electronic Data Transfer

Upon completion of the analysis, perform a QC (calibration verification and blanks verification) overview using the data files from the current run. If the data appears acceptable, transfer the entire contents of the file created by the sequence to the "envchem" directory: "J:\VOC Data Packages\". Store data in the appropriate subfolder by year and quarter. Periodically, check to see that all data files from the local hard drive have been transferred, and then purge them from the local drive to make space for new files.

5.0 QUALITY CONTROL

5.1 Performance Samples

When available, independent quality assurance samples will be obtained and analyzed to verify the proficiency of instrumentation and analysts. Both blind (obtained from an outside source, true values not known at the time of analysis) and reference samples (samples obtained from an outside source with known true values or internally prepared standards) can be used to perform this function. The percent recovery for the audit standards must fall within ± 30%. If the recovery results for the performance sample analyses are not acceptable, an investigation must be performed to determine the source of the conflicting result(s) and corrective action initiated. Daily tuning and calibration checks will also provide reference sample performance data.

5.2 Tuning

At the beginning of each day of operation or prior to calibration, the GC/MS system must be tuned to verify that acceptable performance criteria are achieved. The internal standard also contains the tuning standard, usually BFB. The continuing calibration standard run at the start of each day may be evaluated as a tune standard as well as a continuing calibration.

The key ions and ion abundance criteria that must be met are illustrated in the table below. Analysis must not begin until these criteria are met. The tuning verification must be

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repeated for each 24 hours of operation. This is easily accomplished, since the BFB is added to every injection along with the internal standards.

Table 2. BFB Tune Criteria

Mass	Ion Abundance Criteria
50	8 to 40% of mass 95
75	30 to 66% of mass 95
95	Base Peak, 100% Relative Abundance
96	5 to 9% of mass 95
173	<2% of mass 174
174	50 to 120 % of mass 95
175	4 to 9% of mass 174
176	93 to 101% of mass 174
177	5 to 9% of mass 176

5.3 Continuing Calibration Verification Analyses

The instrument calibration is verified daily before using the instrument for sample analysis, and at least once every 24 hours using a standard (CCAL) at or near the mid-point of the calibration curve. The RPD of each response factor from the mean response factor of the initial calibration curve must be ≤ 30% for continued use of the mean response factors. If the RPD is > 30%, recalibration is probably required.

The %ARC for the internal standard must be within ± 40% of the mean area response in the most recent calibration. The retention time shift for the internal standard must be within ± 20 seconds compared to the mean retention time in the most recent calibration. Any problems encountered must be corrected before analysis can continue.

5.4 Blank Verification

Prior to sample analysis and following the continuing calibration evaluation, a humid UHP nitrogen or zero air blank is analyzed. The gas stream is allowed to run through the analytical system in the same way as a sample. The analysis results of the blank must be <CRQL for all target compounds and <10 ppbv concentration of total VOC for the system to pass the certification. Any problems encountered must be corrected before analysis can continue. The blank verification must be repeated for each 24 hours of operation. When an unusually concentrated sample is encountered (see section 1.3) a blank sample should be analyzed prior to continuing analysis.

5.5 Sample Analysis Technical Acceptance Criteria

The %ARC for the internal standard must be within ± 40% of the mean area response in the most recent calibration. The retention time shift for the internal standard must be within ± 20 seconds compared to the mean retention time in the most recent calibration. If the internal standard does not meet these criteria the system must be inspected for malfunction and the sample reanalyzed. If reanalysis also fails, both sets of data will be reported with an explanation in the narrative of the problems encountered and the corrective actions taken.

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Sample results must fall within the concentration range of the calibration standards. Samples with compounds of interest falling outside the calibration range must be diluted and rerun. Some samples may require serial dilutions to obtain results within the calibration range.

5.6 Laboratory Control Sample

A spiked blank canister, called LCS, must be prepared using a different source for the compounds of interest than the calibration standards. This spiked canister must be analyzed with each batch of ten or fewer samples. Recovery of the compounds of interest must be within ± 40% of the calculated true value of the canister. If the LCS recovery is outside the acceptance windows, reanalysis of all associated samples must be performed.

5.7 Duplicate Analyses

For each batch of ten samples or less a duplicate analysis of a sample and a duplicate analysis of the LCS must be analyzed. The RPD must be ≤ 25% for all target compounds present at concentrations greater than the lowest calibration level from the initial calibration. Recovery outside the acceptance limits indicates that reanalysis is required.

5.8 Data Validation and Feedback

All data shall undergo an exhaustive review for QC acceptance and for sample anomalies by the primary analyst. When QC acceptance criteria indicate a failure in instrumental analysis, the analyst will schedule reanalysis. If reanalysis is not possible or re-analysis also fails, a "Non-Routine Event" form will be completed. The Enviroquant® software provided with the instrument will be used to perform QC evaluation and to produce the final report with the exception of the internal standard percent recovery, LCS recovery, and LCS and sample duplicate precision, which are calculated in Microsoft Excel®. Data validation will be conducted according to OC-PROC-005, "Data Validation and Reporting of Volatile Organic Compounds from Gas Chromatography/ Mass Spectrometry Analysis of Ambient Air in Canisters."

6.0 PROCEDURE

6.1 Analytical Configuration

The analytical configuration used will follow the concepts outlined in EPA Procedure TO-14A and TO-15. The canister to be analyzed will be connected by way of a heated transfer line to the Entech 7100 Preconcentrator. Up to four samples at a time may be connected to the preconcentrator without an autosampler. A pressurized internal standard/tuning standard mixture is attached to the internal standard inlet. A pressurized calibration standard is attached to either the calibration standard inlet or a sample inlet.

6.2 Instrument Start-up

Perform the routine maintenance procedures outlined by the manufacturer.

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If it is not already loaded, load instrument software, Chemstation®. Allow the system to perform diagnostic checks. If an error message occurs, follow instructions supplied by the error message.

Check the helium carrier gas cylinder pressure prior to the start of an analysis sequence to be certain that there is sufficient pressure to maintain proper flow rates within the system. The high side of the helium pressure gauge should read greater than 200 psig and the low side should read 80-100 psig. If there is insufficient pressure in the tank, the tank will need to be changed prior to the start of analysis. Prior changing the gas cylinder, be sure to turn off heated zones and stop gas flow to prevent damage to the instrument. Note the pressure in the internal standard canister by looking at the gauge on the canister regulator.

Open the most recent preconcentrator QA report. These files are located in the "C:\Smart" directory and are named by their run date according to the following convention: "R_YYMMDD." Note the blank canister pressure by looking at the system pressure reading for event 9 on a blank sample run. Note the calibration standard pressure by looking at the system pressure for event 9 on a calibration standard run. The pressure in the canister will change approximately 0.5 psia for each 100 mL of sample or standard injected. Using this estimated drop rate, estimate the pressure needed for the calibration standard and blank sample for the entire day's run. Subtract these amounts from the actual pressure in the canisters. If the resultant residual pressure in any canister will likely fall below 17 psia during the run, change the calibration standard canister and/or pressurize the blank canister prior to starting the run. This will ensure sufficient pressure in each required canister to assure reproducible injection volumes.

Check the liquid nitrogen level in the large tank which provides cryogen to the preconcentrator. If it is low, change out the tank prior to beginning a sequence, if possible. When the tank feels fairly empty, as checked by observing the weight when it is tipped slightly to one side, it should be changed.

When initially setting up the instrument or following significant repairs, perform a leak check of each connection using an identifiable gas (argon or BFB). Set the mass spectrometer collection parameters to monitor correct masses for the gas used. Start run and monitor for peaks corresponding to the gas. Releases near the connection between the column and the interface area, the terminal end of the column, the inlet end of the column and the sample introduction area must be monitored. Correct any leaking connections. Helium leaks may be found using a portable gas leak detector.

An initial five level calibration should be performed prior to initiation of quantitative analysis. Calibration must be repeated whenever the continuing calibration check does not meet the specified performance criteria. Refer to Section 3.0 for detailed instructions on performing an initial calibration.

Check the instrument tuning and calibration every 24 hours of operation using the continuing calibration standard. The tuning results must meet the criteria outlined in section 5.2 and CCAL must meet the criteria in section 5.3 prior to initiation or continuation of sample analysis. A method blank must also be run every 24 hours following the CCAL and the blank must meet the criteria in Section 5.4 prior to initiation or continuation of sample

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analysis.

Prior to beginning the sequence of QC samples, it may be necessary to run a pre-blank, or blank sample to clear out the preconcentrator traps and connections.

6.3 Routine Operation

From the Instrument Control screen of the ChemStation software, load the method "VOCTO15". From the toolbar, select "View", then "Top", then "Sequence". Create a sequence that includes a continuing calibration standard followed by a method blank. Be certain to enter the correct data file storage names in the sequence table. The naming convention used for data files is MMDDYYXX, where MMDDYY is the run date and XX indicates the run number within the sequence. For example, 04271003 is the third GC/MS analysis on April 27, 2010. For sample runs, enter sample names in the sequence with no spaces and no identical entries. Create the same sequence for the preconcentrator using the Entech 7100A Software, using the sample naming convention used for the ChemStation software. Ensure that the internal standard volume is set to the desired injection volume (usually, 150cc for a 20ppbv standard).

After canisters are attached to the autosampler or preconcentrator inlet, a manual leak test is to be performed to ensure tight connections between the canisters and manifold. Ensure that the canister valves are closed prior to initiating the leak test. In the preconcentrator software, click the screwdriver icon labeled "MANL." Set the stream and location to an inlet and port in use and click "update." (On the GC/MSD 5973, the first line is "5" for autosampler 1 and the second line corresponds to the autosampler inlets, labeled 1-16.) Pull a vacuum on the port by clicking "vacuum" until the pressure is less than 1psi and then click "isolate." The pressure should maintain below 1.5psi for 30 seconds. If the vacuum does not hold or changes quickly, adjust the connection and repeat the leak test until it passes. After each utilized inlet has passed the leak check, canister valves should be opened and analysis can proceed. It is highly recommended that a blank is run after performing a leak test to eliminate contamination in the preconcentrator introduced from the leak test.

To start a sequence, always start the ChemStation sequence first, then the Entech 7100A sequence. The preconcentrator will wait for a "ready" signal from the GC. When the GC sequence starts, a prompt will come up asking where the data files will be stored. Ensure that the correct location is listed: "C\MSDChem\1\DATA\MMDDYY", where "MMDDYY" is the run date. Check to be certain the valves are open on the internal standard and calibration standard canisters.

Following the end of a CCAL run, enter the Data Analysis screen and open the data file from the run. Select the peak nearest the known retention time of BFB and select the average of the mass spectrum of the peak. From the toolbar, select "Tuner", then "Evaluate BFB to Screen." This will compare the selected spectrum to the preset BFB tuning criteria which match those in Section 5.2, Table 2. Provided that the tuning criteria are met, the flag to the right of each ion will read "PASS". If the tuning criteria are not acceptable (indicated by a "FAIL" message on the appropriate criteria), re-tune the MSD and try running the standard again. If the criteria are still not met, the source should be cleaned and the MSD re-tuned. Consult the operator's manual and/or the Agilent service center for other possible

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corrections.

If the tune evaluation is acceptable, proceed to the CCAL evaluation as follows. From the toolbar, select "ConCal", then "Evaluate File as Continuing Cal to Screen". Make certain that the results of this standard meet the criteria in Section 5.3. If not, a second continuing calibration sample may be run. If the second sample also fails to meet the criteria, perform another five point instrument calibration prior to proceeding with analysis, unless an obvious reason is observed for the failure of the standards to pass criteria.

Next, to ensure that the system is clean, analyze a blank sample consisting of a volume of ultrapure nitrogen or zero air equivalent to the sample injection volume. The concentration of all compounds of interest must be below CRQL and the total concentration of VOC less than 10 ppbv in order to proceed. If they are not, contamination of the system or instrument problems may exist. If necessary, consult with the laboratory manager or an experienced analyst for help in identifying the problem.

When acceptable results are achieved for the blank, analyze a standard (LCS) of known concentration prepared from a different stock standard than the calibration standard. Check to be certain the quantitation results of this sample meet the requirements in Section 5.6. If not, attempt to determine the cause and repeat the sample run until an acceptable result is obtained prior to continuation of the analysis. Evaluate instrument precision by injecting a duplicate sample of the LCS. Check that quantitation results of the LCS and LCS duplicate meet the requirements in Section 5.7.

When all QC criteria have been met, install up to sixteen sample canisters on the preconcentrator and/or autosampler and add them to the sequences on both the ChemStation software and the preconcentrator software. Start the analytical run. Samples may be run for up to 24 hours following a successful continuing calibration check, BFB tune, and blank check. At that time, the CCAL, BFB tune, and blank must be rerun prior to continuing with sample analysis.

6.4 Instrument Shut Down

The Preconcentrator and GC/MS automatically return to standby mode when analysis is complete. The vacuum, carrier gas and GC oven systems should remain on, when possible. If complete shutdown is necessary, refer to the operator's manual for all shut down requirements. *NOTE: The MSD source is heated during operation of the instrument. Do not vent the MSD for any reason without first completing a "Vent" cycle from the ChemStation software to avoid damaging the mass analyzer. Venting a hot source can cause oxidation of delicate components within the ion source.*

6.5 Sample Dilution

Samples analyte concentrations greater than 100 ppbv exceed the calibration limit and may be diluted either by serial actual dilution with UHP nitrogen or by injection of a proportionately smaller volume of sample (for example, injection of a 150 mL sample would result in an effective 1 to 2 dilution). Samples and standards may be injected in

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volumes ranging from 40 to 800 mL. Higher and lower injection volumes may result in inaccurate preconcentration and may invalidate the quantitation.

Sample dilutions will be recorded on a sample preparation form. If it is necessary to release pressure in a canister before performing a dilution, release excess sample into the exhaust ventilation duct in order to minimize personnel exposure to potentially hazardous substances. Sample dilutions are performed as follows:

6.5.1 Turn on the Entech 4600A Dynamic Diluter if not already on.

6.5.2 Open the UHP nitrogen cylinder valve. Adjust the regulator pressure accordingly based on sample canister size. For a 6L canister, 30-50psig is an appropriate regulator pressure.

6.5.3 Open the 4600A software and press the "Flush" button to enter the canister pressurization controls screen. Set the flush time to 1 minute and press "Go". After the initial 1 minute flush, the flush time should be set to 0.1 minutes for flushes between dilutions.

6.5.4 Press the "Flush" button on the diluter to purge it with UHP nitrogen.

6.5.5 Attach the canister to the fill line.

6.5.6 Set the Entech software to perform the appropriate relative dilution of "2x" or "3x". Open the canister and allow the initial pressure to equilibrate. Then press the pressurize button on the Dynamic Diluter.

6.5.7 After the software shows the canister is pressurized, close the canister valve and remove it from the fill line.

6.5.8 Record appropriate information for entry onto a sample preparation form.

6.5.9 Close the UHP nitrogen valve.

6.5.10 If the dilution process must be repeated to achieve analyte concentrations in the calibration range, a two hour equilibration-period must be allowed prior to subsequent dilution.

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7.0 QUALITY ASSURANCE

7.1 Standards

7.1.1 Primary Standards

All primary stock standards must be recorded in the VOC Primary Standards Logbook upon receipt and given a unique identification number. Each standard is an independent entry. Each entry contains the standard name or description, the vendor or manufacturer, the matrix in which the standard is made, if applicable, the manufacturer's lot number, the date received, the expiration date, the laboratory assigned standard identifier, and the receiving person's initials. If a standard does not have an expiration date from the manufacturer, the default expiration date one year from receipt will be assigned. Each entry also contains a column for date removed from service. This column will be completed when the standard is consumed, expired, or found to be compromised in any way.

The independent laboratory identifiers for VOC primary standards are assigned as follows. The VOC Primary Standards Logbook shall be identified as VOC Primary Standards-XXX, where the XXX values will begin with 001 and are incremented every time a new logbook is started. Each page is numbered in the logbook prior to initial use of the book. Each line on the left-hand page of the logbook will be numbered. A single standard will be entered per line. The primary standard's identifier will be the combination of "VOC", the logbook number, the page number and the line number. For example standard "VOC 001-018-23" would be the standard entered in login notebook number one on page eighteen on line twenty-three. This identifier is written on the label of the standard and on its certificate. Completing the "date out of service" column in the logbook facilitates completion of the chemical inventory when required. This identifier will be used in analysis logbooks, on preparation bench sheets, and in intermediate standard logbooks to unambiguously identify the standards used in preparation and analysis.

7.1.2 Intermediate Standards

All intermediate standards prepared for use in calibration must be documented in the "VOC Intermediate Standards Preparation" logbook. Standards will be prepared according to OC-PROC-004 and CP-PROC-009.

7.2 Maintenance Logs

Maintenance logs will be kept up-to-date. All routine maintenance operations will be documented as well as instrument service. All service entries will include malfunction symptoms, a description of repairs made, and a description of how a return to acceptable operation was determined. Routine maintenance procedures, along with repairs and troubleshooting procedures applied will be documented in the appropriate instrument maintenance log. The instrument manufacturer will provide documentation of repairs performed by service engineers.

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7.3 Instrument Analysis Logs

Analysis logs will be maintained and they will record a list of samples analyzed, sample injection volumes, internal standard injection volumes, standards used, inlet number and/or autosampler location used, the date analyzed and the data file storage information. Analysis log headers will contain enough information to identify all standards used in analysis and their expiration dates, along with the model numbers and serial numbers used to perform the analysis run. Any problems encountered during analysis should also be noted. Analysis logs should be updated daily or as needed.

7.4 Calibration Records

Electronic copies of calibration files will be maintained in archive in the envchem directory, "J:\envchem\VOC Data Packages".

7.5 Control Charts

Control charts may be used to perform trend analyses on the laboratory data, but will not be used to establish acceptance windows.

8.0 REFERENCES

40 CFR Part 136, Appendix B, Definition and Procedure for the Determination of the Method Detection Limit, Rev.1.11

CP-PROC-009, Standards Log-in and Preparation

Entech 7100 Preconcentrator Operators Manual. Entech Instruments, Simi Valley, CA.

EPA Method TO-14A "Determination of Volatile Organic Compounds (VOC) In Ambient Air Using Specially Prepared Canisters With Subsequent Analysis By Gas Chromatography", Compendium of Methods for the Determination of Toxic Organic Compounds in Air, Second Edition, January 1999.

EPA Method TO-15 "Determination of Volatile Organic Compounds (VOC) in Air Collected in Specially–Prepared Canisters and Analyzed By Gas Chromatography/Mass Spectrometry (GC/MS)", Compendium of Methods for the Determination of Toxic Organic Compounds in Ambient Air, Second Edition, January 1999.

HP 5973 MSD Hardware Manual. Hewlett Packard Company, Wilmington, Delaware, First Edition, May 1996.

HP 6890 Series Gas Chromatograph Hardware Manual. Hewlett Packard Company, Wilmington, Delaware, Second Edition, December 1997.

OC-PROC-004, Preparation of Calibration Standards in Specially Prepared Canisters for Analysis by Gas Chromatography/Mass Spectrometry

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OC-PROC-005, Data Validation and Reporting of Volatile Organic Compounds from Gas Chromatography/Mass Spectrometry Analysis of Ambient Air in Canisters for the WIPP Volatile Organic Compound Monitoring Plan

SM-03, Carlsbad Environmental Monitoring and Research Center Chemical Hygiene Plan

"Volatile Organics Analysis of Ambient Air in Canisters", USEPA Contract Laboratory Program, VCAAO 1.0, Publication 9240.1-18, EPA/540/r/085, PB95-963524, December 1994.

9.0 RECORDS

9.1 GC/MS Analysis Logbook
9.2 GC/MS Maintenance Logbook
9.3 VOC Intermediate Standards Preparation Logbook
9.4 VOC Primary Standards Logbook

10.0 APPENDICES

NA

11.0 REVISION HISTORY

Revision #	Date	Description
0	12/19/03	Initial Document, issued as EC-PROC-020.
1	02/17/06	Review and revision; reflects transfer of responsibility to the organic chemistry group; released as OC-PROC-003
2	04/03/08	Incorporate the temporary deviation and make minor changes.
3	05/03/10	Incorporate temporary deviation and minor modifications for clarification

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TEMPORARY PROCEDURAL DEVIATION FORM

Carlsbad Environmental Monitoring & Research Center

Title: Preparation of Calibration Standards in Specially Prepared Canisters for Analysis by Gas Chromatography Mass Spectrometry	
Document No: QC-PROC-004-004-021606	**Implementing Program: Organic Chemistry**

Deviation:
On page 4, section 2.3.1 reads 'The standard shall be humidified by the addition of approximately 50 µl of ultrapure water to the canister'.

It should be read as follows: "The standard shall be humidified by the addition of approximately 50 µl of ultrapure water to the canister if applicable.' Fused silica lined canisters (e.g. Silonite Canisters) need no additional water to keep VOCs in the gas phase".

On page 6, section 3.1.7 reads "Prepare a syringe for addition of ultrapure water to canister by rinsing the syringe with the ultrapure water three times. Keeping the canister valve closed, add 50 µl of ultrapure water to the inlet of the canister".

It should be read as follows: "Add 50 µl of ultrapure water to the inlet of the canister if applicable. (50 µl of ultrapure water should be added to the top of a SUMMA canister, while Fused silica lined canisters (e.g. Silonite Canisters) need no additional water)
In case of ultra pure water addition to canister, Prepare a syringe by rinsing with the ultrapure water three times. Keeping the canister valve closed, add 50 µl of ultrapure water to the inlet of the canister".

Justification: This deviation is in accordance of the Entech 4600 Dynamic Diluter's operation manual. Linde Electronics and Specialty Gases also suggest avoiding water addition to the canister for use with a TO-15 standard because of the polar compounds. Analyst also experienced the higher deviation in the responses of some compounds with water addition to the Silonite canisters.

Valid for: __Until next revision_____

Approved by:__ _____ 07/21/11
Responsible Manager **Date**

STAFF TRAINING

I have reviewed and I understand the procedure deviation described above.

Print Name	Signature	Date

DC-PROG-001-007 070297

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TEMPORARY PROCEDURAL DEVIATION FORM

Carlsbad Environmental Monitoring & Research Center

Title: Preparation of Calibration Standards in Specially Prepared Canisters for Analysis by Gas Chromatography Mass Spectrometry	
Document No: OC-PROC-004-004-021606	**Implementing Program: Organic Chemistry**

Deviation: On page 11, third paragraph reads "Preparation of Canisters for Ambient Air Sampling", OC-PROC-002.

It should read as follows: "Preparation of Canisters and Sample Trains for Ambient Air Sampling", OC-PROC-002.

Justification: Title of OC-PROC-002 was changed after OC-PROC-004-004-021606 was issued.

Valid for: until next revision

Approved by: EAStone **Responsible Manager** 5/14/10 **Date**

STAFF TRAINING

I have reviewed and I understand the procedure deviation described above.

Print Name	Signature	Date

DC-PROG-001-007-070297

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CARLSBAD ENVIRONMENTAL MONITORING & RESEARCH CENTER (CEMRC)

PREPARATION OF CALIBRATION STANDARDS IN SPECIALLY PREPARED CANISTERS FOR ANALYSIS BY GAS CHROMATOGRAPHY MASS SPECTROMETRY

OC-PROC-004-004-021606-_ _ _

Effective Date: April 19, 2010

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PREPARATION OF CALIBRATION STANDARDS IN SPECIALLY PREPARED CANISTERS FOR ANALYSIS BY GAS CHROMATOGRAPHY MASS SPECTROMETRY

1.0 PURPOSE

The purpose of this procedure is to describe the preparation of standards and laboratory blanks for use in the analysis of VOC in ambient air as outlined in EPA Method TO-15. VOC are separated by gas chromatography and identified and quantitated by mass spectrometry. This method is applicable to specific VOC that have been tested and determined to be stable when stored in pressurized and sub-atmospheric pressure canisters. Compounds of interest and levels of calibration standards prepared will depend upon individual project requirements.

1.1 Scope and Limitations

Included in this procedure are instructions for the following activities:

- Preparation of calibration and quality control standards.
- Preparation of laboratory blanks

1.2 Process Overview

Calibration standards are prepared in cleaned canisters (canister preparation is described in OC-PROC-002) by dilution of a certified primary standard using the Entech 4600 Dynamic Diluter as described below. Laboratory blanks are prepared in cleaned canisters by filling with UHP nitrogen.

1.3 Definitions

Calibration Standard (CAL) - A gas standard of known concentration for each compound of interest prepared in a specially prepared canister. A primary standard gas mixture is diluted with a diluent gas (UHP nitrogen), to appropriate concentration(s).

Gas Chromatography (GC) - An analytical technique used for separating compounds of interest.

Identification Number (ID) - A unique number given to primary and intermediate standards.

Laboratory Control Sample (LCS) - A standard containing compounds of interest which has been prepared by dilution of a standard mixture from a source (or batch of raw materials) different than that used to prepare the calibration standard.

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Mass Spectrometer (MS) - An analytical technique used for quantitative analysis and/or for the qualitative determination of the molecular composition of a sample. A MS consists of ionizing compounds to generate charged molecules, separation of ions based on their mass-to-charge ratio (*m/z*), and detection of the charge induced or current produced when ions interact with the detector's surface.

National Institute of Standards and Technology (NIST) - A federal agency that develops and promotes measurement, standards, and technology.

Organic Chemistry (OC) - The CEMRC Organic Chemistry department.

ppbv - Parts per billion by volume, a unit of concentration in which the volume of a gaseous analyte is normalized to 10^6 volumes of ambient air.

psia - Pounds per square inch absolute pressure, measured relative to vacuum. The psia unit of measure is not affected by local barometric pressure and is used by the Dynamic Diluter.

psig - Pounds per square inch gauge pressure, measured relative to local barometric pressure.

sccm - Standard cubic centimeters per minute, a flow rate measurement for gases.

Ultra High Purity (UHP) - A grade of reagent gas certified as free of impurities.

Volatile Organic Compounds (VOC) - An organic compound that has a high enough vapor pressure to be in the gas phase at ambient temperature and pressure.

2.0 REQUIREMENTS

2.1 Training Requirements

Laboratory personnel should verify that this procedure is the latest available revision by checking the Organic Chemistry Laboratory Procedure Manual against the version in H:\document control\oc. The latest revision of this document shall be in the Organic Chemistry laboratory at all times. If this procedure is found to be unsatisfactory in any way and a procedure revision is required, notify the project manager or designee.

2.2 Safety

Some standards prepared in this method contain known carcinogens. Exposure to these compounds will be minimal as long as the operating procedure is followed.

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Before use, the user must ensure that the laboratory exhaust system is functioning properly.

The standards and the diluent gas used for standards preparation are stored at high pressure. Care must be taken to ensure that the integrity of the cylinders is maintained. Handling of standard gases and diluent gases will be done in accordance with the CEMRC Chemical Hygiene Plan, SM-03.

Eating or drinking in the laboratory is strictly prohibited.

2.3 Equipment and Maintenance

2.3.1 Entech 4600 Dynamic Diluter

The Entech 4600 Dynamic Diluter can be used to create intermediate standards by diluting a primary standard to a target concentration. This technique utilizes the Dynamic Diluter's pressurization feature, provided that the standard dilution factor is no greater than 25x of the primary standard. For larger dilution factors, it is not possible to accurately measure the pressure of primary standard added. Static dilutions can be performed on a prepared standard to achieve lower concentrations. The standard shall be humidified by the addition of approximately 50µl of ultrapure water to the canister.

The Entech 4600 Dynamic Diluter will be set up according to the manufacturer's instructions. The linearity of the Dynamic Diluter's pressure transducer readings should be periodically checked against a calibrated pressure gauge, when pressure dilutions are being regularly performed.

2.3.2 Stainless steel passivated SUMMA® or equivalent canisters (e.g. Silonite®, TO-Can™)

Standard canisters are prepared for use as specified in OC-PROC-002. Cleaned and certified stainless steel canisters, typically 400-6000mL capacity, are required for standard, LCS, and blank preparation.

2.3.3 Calibrated Pressure Gauge

A pressure gauge is used to confirm above-ambient pressures in canisters and to check the linearity of the Dynamic Diluter pressure transducer readings.

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2.4 Primary Standards and Reagents

NIST traceable primary standard gases are used to create the humid static working standards. All primary standards shall be purchased from a qualified vendor in accordance with CP-PROC-017, "Procurement of Items and Services". Primary standards shall be received and documented according to CP-PROC-009, "Standards Log-in and Preparation". Purchased standards are assigned a unique standard ID number upon receipt that is used to identify the standard on all standard preparation documentation in which that standard is used to prepare an intermediate or working standard. All purchased primary standards must be accompanied by a certificate of analysis, which are kept in the "OC Primary Standards Certificate of Analysis Logbook." All purchased reagent gases must be accompanied by a certificate of analysis, which are kept in the "Ultra-High Purity Gas Certificate of Analysis Logbook".

The following is a list of primary standards and reagents used in this procedure:

- Certified TO-15 VOC mixture standard, balanced with nitrogen. This standard is used for preparation of CAL standards.
- Certified TO-15 VOC mixture standard, balanced with nitrogen from an independent source (i.e. different lot of raw materials). This primary standard is used for the LCS.
- UHP nitrogen
- Ultrapure deionized water (for humidification of standards; specific resistance of 17.8 megaohm-cm or greater, free of measurable ionic impurities, organics, microorganisms and particulate matter larger than 0.2μm).

2.5 Prerequisite Actions

2.5.1 Make certain that personal protective equipment is available and used when appropriate.

2.5.2 Check the expiration dates on all standards to be used to ensure that they are not expired.

2.5.3 Check that cleaned canisters are available for all standards that must be prepared.

2.5.4 Check the maintenance log to be certain that no routine maintenance is due prior to standard preparation.

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3.0 PROCEDURE

3.1 Standard Preparation

3.1.1 Before beginning to prepare a standard, make certain that the appropriate diluent gas (UHP nitrogen) is attached to the diluent gas inlet channel of the dynamic diluter. Check the connections for leaks prior to starting the standard preparation. Make certain the humidification chamber on the diluent inlet has sufficient water.

3.1.2 Open the UHP nitrogen valve.

3.1.3 If it is not already on, turn on the Dynamic Diluter. Click the 4600A software using the icon on the desktop of the attached computer. In this program click the "Flush" icon to enter the canister pressurization controls screen. Set the flush time to 1 minute and press "Go".

3.1.4 Press the "Flush" button on the Dynamic Diluter to purge the line with UHP nitrogen.

3.1.5 Open the "Intermediate Standard Preparation Form" Microsoft Excel file in the envchem/VOCs/ directory. Save the spreadsheet as its appropriate intermediate standard ID in the most recent "OC Intermediate Standards Preparation Logbook" folder. The intermediate standard ID begins with an "I" for intermediate, followed by the current logbook number (e.g. "005), followed by the next sequential page number in the logbook, and finally the number of the intermediate standard documented on the page. Fill in the canister serial number, type of standard (CAL or LCS), designated concentration, diluent lot number, primary standard number, and expiration date. Copy the primary standard concentrations to the spreadsheet from the primary standard certificate of analysis. Save the spreadsheet.

3.1.6 Begin with a cleaned and evacuated canister.

3.1.7 Prepare a syringe for addition of ultrapure water to canister by rinsing the syringe with the ultrapure water three times. Keeping the canister valve closed, add 50µL of ultra pure water to the inlet of the canister.

3.1.8 Attach the canister to the Dynamic Diluter. Set the Entech software to dilute to a target pressure of 13psia with UHP nitrogen and set the "allowed offset" to zero.

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3.1.9 Open the canister valve and wait for 5 seconds for the pressure reading to stabilize. Confirm that the canister is at a vacuum and that there are no leaks. Press the pressurize button on the Dynamic Diluter.

3.1.10 After the software shows the canister is pressurized, close the canister valve and remove it from the fill line.

3.1.11 Log the actual pressure on the Intermediate Standards Preparation Form as the "reading after initial diluent added."

3.1.12 Set the regulator on the primary standard to approximately 2.5psig and open valve to gently flush its lines. A box fan set to medium speed should be used to blow the primary standard away from the operator. Such ventilation will help to minimize operator exposure to VOC. *(Note: the primary standard remains detached from the Dynamic Diluter the standards preparation.)*

3.1.13 Connect the canister to the primary standard while it is flushing. An extension may be necessary to extend the primary standard output to the canister. Open the canister valve and let the primary standard enter the canister. The longer the valves are open, the more standard will enter the canister.

3.1.14 Close the valves on the canister and primary standard and release the primary standard regulator. Disconnect the canister from the primary standard.

3.1.15 Check the pressure on the receiving canister. Log this pressure on the Intermediate Standard Preparation form as the "reading after standard added."

3.1.16 Use the spreadsheet to determine the final canister reading needed to achieve the desired concentration. The required calculation is:

Final pressure (psia) = (Initial standard concentration(ppbv)/ Desired final standard concentration (ppbv))* Pressure of standard added (psia)

3.1.17 Pressurize the canister to the desired pressure using the Dynamic Diluter and log the final pressure on the Intermediate Standard Preparation form as the "final canister reading."

3.1.18 Close the UHP nitrogen canister valve.

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3.1.19 Complete the Intermediate Standard Preparation form by filling in the date/time prepared and expiration date, which is one month from its preparation, and the preparer of the standard.

3.1.20 Print a copy of the spreadsheet and archive it into the VOCs Intermediate Standards Preparation Logbook and sign.

3.1.21 Fill out a standard preparation label containing the standard concentration, standard type (e.g., "CAL", "LCS", or "MDL"), preparation analyst's initials, date prepared, standard ID number (see CP-PROC-009 for details), and the standard's expiration date and place it on the back of the CEMRC canister tag.

3.2 Laboratory Blank Preparation

3.2.1 Turn on the Entech 4600A Dynamic Diluter if not already on.

3.2.2 Open the UHP nitrogen cylinder valve. Adjust the regulator pressure accordingly based on laboratory blank canister size. For a 6L canister, the regulator pressure should be approximately 30-50psig.

3.2.3 Open the 4600A software and press the "Flush" button to enter the canister pressurization controls screen. Set the flush time to 1 minute and press "Go".

3.2.4 Press the "Flush" button on the diluter to purge it with UHP nitrogen.

3.2.5 Take a cleaned and evacuated canister and attach the canister to the fill line.

3.2.6 Set the Entech software to dilute to a target pressure of approximately 30psia. Open the canister and press the pressurize button on the Dynamic Diluter.

3.2.7 After the software shows the canister is pressurized, close the canister valve and remove it from the fill line.

3.2.8 Close the UHP nitrogen valve.

3.2.9 Label the canister as a laboratory blank (LB), along with the date prepared, preparer's initials, and UHP nitrogen lot number.

3.3 Static Dilution

3.3.1 Turn on the Entech 4600A Dynamic Diluter if not already on.

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3.3.2 Open the UHP nitrogen cylinder valve. Adjust the regulator pressure accordingly based on laboratory blank canister size. For a 6L canister, the regulator pressure should be approximately 30-50psig.

3.3.3 Open the 4600A software and press the "Flush" button to enter the canister pressurization controls screen. Set the flush time to 1 minute and press "Go".

3.3.4 Press the "Flush" button on the diluter to purge it with UHP nitrogen.

3.3.5 Take the canister to be diluted and position it underneath a snake vent. Open the valve to release the pressure until only a low hissing sound remains. A hissing sound is desirable and indicates that the pressure inside the canister is greater than ambient pressure.

3.3.6 Set the Entech software to do a dilution by choosing a preset dilution factor (1.5, 2.0, or 3.0) or choosing an appropriate target pressure. Ensure that the final pressure of the canister will not exceed 45psia. Open the canister and press the pressurize button on the Dynamic Diluter.

3.3.7 After the software shows the canister is pressurized, close the canister valve and remove it from the fill line.

3.3.8 Close the UHP nitrogen valve.

3.3.9 Record the initial and final pressure readings, dilution factor, and the date and time the dilution was performed. If diluting an intermediate standard, this information must be recorded on the Intermediate Standard Preparation Form.

4.0 QUALITY ASSURANCE

4.1 Documentation

4.1.1 Standards

All primary standards must be recorded in the OC Primary Standards Certificate of Analysis Logbook upon receipt and must be given a unique identification number. All intermediate standards prepared for use in calibration and QC checks must be documented in the OC Intermediate Standards Preparation Logbook. Documentation of standards is outlined in CP-PROC-009.

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4.1.2 Maintenance Logs

Maintenance logs will be kept up to date. All routine maintenance operations will be documented as well as instrument service. All service entries will include malfunction symptoms, a description of repairs made, and a description of how return to acceptable operation was determined. The routine maintenance schedule provided by the manufacturer may be used to document performance of routine maintenance procedures. The instrument manufacturer will provide documentation of repairs performed by service engineers. This will be incorporated into the maintenance log.

4.2 Reagent and Standard Verification

Nitrogen used to dilute primary standards will be analyzed by GCMS prior to use in standard preparation to assure that it contains no significant contamination.

Analysis of LCS and performance check samples will be used to verify calibration standards. The accuracy of the calibration may be verified by obtaining an LCS recovery of +/- 40% and / or recovery of a performance sample within +/- 30% of the true values. Linearity of the calibration is demonstrated by obtaining a calibration curve in which the relative standard deviation is less than 30%. Obtaining results which meet these performance criteria indicates the proper preparation and analysis of calibration standards.

4.3 Holding Time

Intermediate standards in SUMMA or equivalent canisters are valid for one month from the date of preparation.

4.4 Equilibration Time

Standards prepared into SiloniteTM coated canisters by any of the methods listed herein must be allowed to equilibrate for a minimum of 2 hours between preparation and analysis. Similarly, standards must be allowed to equilibrate for a minimum of 2 hours between preparation and dilution. Equilibration assures that consistent results will be obtained by repeated analysis.

5.0 REFERENCES

EPA Method TO-15 "Determination of Volatile Organic Compounds (VOCs) In Air Collected In Specially-Prepared Canisters And Analyzed By Gas Chromatography/Mass Spectrometry (GC/MS)", Compendium of Methods for the Determination of Toxic Organic Compounds in Ambient Air, Second Edition, January 1999.

Entech 4600 Dynamic Diluter Operator's Manual, Entech Instruments, Simi Valley, CA.

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"Volatile Organics Analysis of Ambient Air in Canisters", USEPA Contract Laboratory Program, VCAAO 1.0, Publication 9240.1-18, EPA/540/r/085, PB95-963524, December 1994.

Carlsbad Environmental Monitoring and Research Center Chemical Hygiene Plan, SM-03.

"Preparation of Canisters for Ambient Air Sampling", OC-PROC-002.

"Gas Chromatography – Mass Spectrometry Analysis of Volatile Organic Compounds (VOCs) in Ambient Air from Canisters at PPBV Concentration Levels", OC-PROC-003.

"Standards Log-in and Preparation", CP-PROC-009.

6.0 REVISION HISTORY

Revision #	Date	Description
0	03/19/03	Initial document issued as EC-PROC-023.
1	12/19/03	Rewritten to include use of the dynamic diluter
2	02/16/06	Review and major rewrite to incorporate standards preparation by the pressure dilution technique. Also, transfer of responsibility to OC Lab and reissue as OC-PROC-004.
3	03/20/08	To incorporate the temporary deviation and make minor changes.
4	04/19/10	Major rewrite to include additional details of sample preparation using the Dynamic Diluter (formerly called the pressure dilution technique) and to remove manual standards preparation. Validation and Verification of the "Intermediate Standard Preparation Form" Microsoft Excel file. Shading was not necessary due to major rewrite.

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CARLSBAD ENVIRONMENTAL MONITORING & RESEARCH CENTER (CEMRC)

DATA VALIDATION AND REPORTING OF VOLATILE ORGANIC COMPOUNDS FROM GAS CHROMATOGRAPHY/MASS SPECTROMETRY ANALYSIS OF AMBIENT AIR IN CANISTERS FOR THE WIPP VOLATILE ORGANIC COMPOUND MONITORING PLAN

OC-PROC-005-005-022106-_ _ _

Effective Date: May 17, 2011

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DATA VALIDATION AND REPORTING OF VOLATILE ORGANIC COMPOUNDS FROM GAS CHROMATOGRAPHY/MASS SPECTROMETRY ANALYSIS OF AMBIENT AIR IN CANISTERS FOR THE WIPP VOLATILE ORGANIC COMPOUND MONITORING PLAN

1.0 GENERAL DISCUSSION

1.1 Purpose

The purpose of this procedure is to describe the steps taken to perform validation and verification of GCMS results and to prepare final reports for the analysis of VOC in ambient air as outlined in EPA Methods TO-14A and TO-15, and the EPA Contract Laboratory Program-Statement of Work for Volatile Organic Compounds in Ambient Air (EPA CLP-SOW VOC in Air). This procedure is written to comply with the WIPP "Confirmatory Volatile Organic Compound Monitoring Plan", WP12-VC.01, and "Volatile Organic Compounds Monitoring Quality Assurance Project Plan", WP12-VC.02.

1.2 Personnel

Laboratory personnel should verify that this procedure is the latest available revision by checking the Organic Chemistry Laboratory Procedure Manual against the version in H:\document control\oc. The latest revision of this document shall be in the VOC laboratory at all times. If this procedure is found to be unsatisfactory in any way and a procedure revision is required, notify the laboratory manager or designee.

1.3 Definitions

Bromofluorobenzene (BFB) - Also called p-bromofluorobenzene or 4-bromofluorobenzene, this is a gaseous organic compound commonly used as a tuning standard GC/MS.

Calibration Standard (CAL) - A specially prepared canister containing a known concentration of each VOC compound of interest.

Continuing Calibration Standard (CCAL) - A daily calibration check standard near the midpoint of the calibration curve.

Contract Required Quantitation Limit (CRQL) - The lowest concentration of a compound of interest which will be reported under a given contract. The CRQL for each compound of interest is defined in the contract Statement of Work. This is the same as the Method Reporting Limit (MRL) for the WIPP VOC Monitoring Plan.

Dilution Factor - Multiplier which adjusts for the difference in the volume of sample injected for analysis and the volume of standard injected for

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calibration. For example, if 200 ml of standard was injected for calibration, but only 100 ml of sample was injected for analysis, the dilution factor would be 2. Dilution factors may also apply to physical dilutions of samples with ultrahigh purity nitrogen.

Dynamic Calibration - Calibration of an analytical system using calibration gas standard concentrations in a form identical or very similar to the samples to be analyzed and by introducing such standards into the inlet of the sampling or analytical system in a manner very similar to the normal sampling or analytical process.

Electronic Data Deliverable (EDD) - An electronic file that summarizes the results of sample and QC standards for a batch of samples.

EPA TO-14(A) and TO-15 - The United States Environmental Protection Agency (EPA) standardized method for analysis of toxic organic compounds in ambient air. EPA TO-14 is the original method, EPA TO-14a is a revised version, and EPA TO-15 is the newest revision with an expanded analyte list.

Gas Chromatography (GC) - An analytical technique used for separating compounds of interest.

Internal Standard Area Response - The integrated area of the quantitation ion of the internal standard.

Internal Standard (IS) - Analyte(s) of known concentration added to a sample, blank, or calibration standard in known amounts; these are used to correct for fluctuations in instrument response. The relative responses of the IS signal in the calibration standards relative to the IS signal in the samples indicates the presence of signal enhancement or suppression, or instrument drift or stability.

Laboratory Control Sample (LCS) - A standard containing compounds of interest which has been prepared from a different source than that used to prepare the calibration standard. An LCS is the same as a spiked blank or blank spike.

Laboratory Control Standard Duplicate (LCSD) - A second analysis of the LCS. Used to verify the instrument calibration, quantitative analytical accuracy, and laboratory precision.

Laboratory Method Blank- A canister pressurized with zero air or ultra-high purity nitrogen. Used to monitor for potential contamination.

Mass Spectrometer (MS) or Mass Selective Detector (MSD) - An analytical detector used for quantitative analysis and/or for the qualitative determination of the molecular composition of a sample. A MS consists of ionization of

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compounds to generate charged molecules, separation of ions based on their mass-to-charge ratio (m/z), and detection of the charge induced or current produced when ions interact with the detector's surface.

National Institute of Standards and Technology (NIST) - A federal agency that develops and promotes measurement, standards, and technology.

Organic Chemistry (OC) - The CEMRC Organic Chemistry department.

Percent Area Response Change (%ARC) – Percent change of the internal standard peaks at each level of the initial calibration, as compared to the mean internal standard response over the entire calibration range.

Percent Relative Standard Deviation (%RSD) - For an initial calibration curve, the standard deviation of the relative response factors at each calibration level divided by the mean of the relative response factors times one hundred. This serves as a measure of the linearity of instrument response over the calibration curve, and it should be less than or equal to 30% for a five point calibration curve.

ppbv - parts per billion by volume, the concentration calculated as the volume of the compound of interest divided by the volume of the dilution gas.

psia - pounds per square inch of pressure, measured relative to absolute zero pressure.

psig - pounds per square inch gauge pressure, measured relative to local barometric pressure.

QA – Quality Assurance

Relative Response Factor (RRF) - The relative response (peak area or height) of an internal standard of known concentration to the response of the analyte of interest.

Retention Time (RT) - The time from the start of the GC analysis run until the center of the peak of the compound of interest reaches the detector.

Retention Time Shift - Retention time of the internal standard peaks in the sample relative to the retention time of the same peaks in the current continuing calibration.

Statement of Work (SOW) – A written document describing the agreement of the client and contracted laboratory. The SOW includes, but is not limited to, technical tasks, quality assurance requirements, data reports and deliverables, and other considerations.

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Tentatively Identified Compound (TIC) - An unknown compound that is tentatively identified by Chemstation Software based on its mass spectrum's similarity to a known compounds mass spectrum. Known compounds' mass spectra are stored in an electronic library stored on the instrument and data analysis computers.

Tuning Standard - A canister containing a known concentration of a tuning compound, normally 4-bromofluorobenzene (BFB), that is used to verify the mass calibration and sensitivity of the instrument are within acceptable limits prior to performing an analysis. The tuning standard results are used to evaluate qualitative accuracy.

Volatile Organic Compounds (VOC) - An organic compound that has a high enough vapor pressure to be in the gas phase at ambient temperature and pressure.

Waste Isolation Pilot Plant (WIPP) – A deep geological repository for the permanent disposal of transuranic waste.

2.0 APPARATUS, INSTRUMENT, REAGENTS, AND FORMS

2.1 Instrument Output

The hardcopy printout for each GC/MS sample and blank reported must include the chromatogram, integration results, library search results, the ion spectra for each peak integrated and the NIST ion spectra for the tentatively identified compounds. For each 24 hours of analysis, hardcopy printouts for the CCAL evaluation, tuning report, chromatogram, and integration results are needed.

2.2 Reporting Forms

The final reports will be in the format approved by the client. Blank forms for the reports have been created in Microsoft Excel®; these will be completed by the analyst for final reporting. For analyses performed under the WIPP VOC Monitoring Program, the blank forms to be used are in the file "I:\envchem\VOC Data Packages\VOC Forms\VOCDP.xls". Copies of the blank forms from this file are contained in Appendices A-D. Some forms are created by functions within the ChemStation software using the EnviroForms package.

Note: Appendices A-D forms are provided in this procedure as informational copies only. Actual report forms are used in the file "I:\envchem\VOC Data Packages\VOC Forms\VOCDP.xls" as approved by the clients.

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3.0 STANDARDS

For all data reported under this procedure, make certain that appropriate standards were used to calibrate the instrument prior to analysis. Check the expiration dates of all standards prior to use.

Gas calibration-standards used to produce standards for calibration of the instrument shall be purchased from a reputable source and shall be certified and traceable to a NIST standard. Certificates of Analysis supplied with standards shall be kept in the "C/MS Standards Logbook. Standards shall be logged in to the Organic Chemistry Primary Standards Logbook and be given a unique standard identification number according to CP-PROC-009, Standards Log-in and Preparation.

4.0 PROCEDURE

4.1 Daily Tune Evaluation

Once every 24 hours when the GC/MS is in use, a BFB tuning standard must be analyzed. The ion abundances must meet the criteria in Table 1.

The analyst must verify that the abundance criteria have been met prior to further analysis. The results of the tuning test are calculated by the ChemStation software and reported using the "BFB Tune" report function. Hardcopies of the tuning reports generated by the software will be maintained as a part of the data package.

Table 1: BFB Tune Criteria

Mass	Ion Abundance Criteria
50	8-40% of mass 95
75	30-66% of mass 95
95	Base Peak, 100% Relative Abundance
96	5-9% of mass 95
173	<2% of mass 174
174	50 - 120% of mass 95
175	4-9% of mass 174
176	93-101% of mass 174
177	5-9% of mass 176

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4.2 Initial Calibration

Nine VOC are of particular interest for the WIPP Confirmatory Volatile Organic Compound Monitoring Program because these are known or likely components of the mixed waste being entombed in the repository. The compounds of interest are:

- Carbon tetrachloride (tetrachloromethane)
- Chlorobenzene (monochlorobenzene, benzene chloride)
- Chloroform (trichloromethane)
- 1,1-Dichloroethylene (vinylidene chloride, 1,1-DCE)
- 1,2-Dichloroethane (1,2-DCA, DCA, ethylene dichloride, EDC, sym-dichloroethane)
- Methylene chloride (dichloromethane)
- 1,1,2,2-Tetrachloroethane (sym-tetrachloroethane)
- Toluene (methylbenzene)
- 1,1,1-Trichloroethane (methyl chloroform)

Eight additional requested analytes are added to the list of compounds of interest. These compounds will be reported as Additional Requested Analyte. These additional requested analytes are:

- Benzene
- Trichloroethylene
- Tetrachloroethylene
- Chloromethane (Methyl Chloride)
- trans-1,2-Dichloroethylene
- 1,2,4-Trimethylbenzene
- p,m-Xylene
- Trichloromonofluoromethane (Freon-11)

The instrument is calibrated with five calibration standards. The nominal standard concentrations are 1, 10, 25, 50, and 100ppbv for each of these compounds. The average RRF for target compounds over all calibration levels is compared to the target compound responses in the sample for determining analyte concentration.

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4.2.1 Relative Response Factors

The RRF is calculated for each VOC compound normalized to the appropriate IS using the following equation:

$$RRF = \frac{(A_x C_{is})}{(A_{is} C_x)}$$

Where:
RRF = relative response factor;
A_x = area of the peak for the primary ion of the compound to be measured;
A_{is} = area of the peak for the primary ion of the IS;
C_x = concentration of the compound in the calibration standard in ppbv; and
C_{is} = concentration of the IS spiking mixture in ppbv.

The mean RRF is calculated by averaging the values obtained at the five calibration standard concentrations. The standard deviation of the RRFs is calculated using the equation shown below.

$$SD_{ref} = \sum_{i=1}^{N} \sqrt{\frac{(RRF_i - RRF_{mean})^2}{(N-1)}}$$

Where:
SD_{ref} = standard deviation of the initial response factors (per compound);
RRF_i = relative response factor at a concentration level;
RRF_{mean} = mean of the relative response factor (per compound); and
N = the number of calibration levels.

The %RSD is calculated using the following equation, and must be equal to or less than 30% for all compounds:

$$\%RSD = \left(\frac{SD_{ref}}{RRF_{mean}}\right) X100$$

The RRF for each calibration level, the mean RRF for the calibration, and the %RSD for the calibration for each compound are calculated by the ChemStation software and reported using the "Response Factor Report" form.

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4.2.2 Internal Standard Area Response

The mean area response for each IS compound over the initial calibration range is calculated. The %ARC is calculated for each IS at each level of the calibration using the following equation:

$$\%ARC = \left[\frac{(A_x - Y)}{Y}\right] \times 100$$

Where:
$\%ARC$ = percent area response change;
A_x = area response, in counts, of the primary ion for the IS at calibration level; and
Y = mean area response, in counts, of the primary ion for the IS over the entire calibration range.

The %ARC at each calibration level must be ± 40% for each IS.

The %ARC is calculated and reported using the "Initial Calibration Internal Standard %ARC" form in the "envchem\VOCDataPackages\VOCForms\VOCDP.xls" spreadsheet.

4.3 Continuing Calibration

The instrument calibration is verified daily or every 24 hours using a calibration standard at or near the mid-point of the calibration curve.

4.3.1 Percent Difference in Relative Response Factors

The RRF for the calibration (RRF_c) is calculated for each compound to the appropriate IS using the equation outlined in 4.2.1. The percent difference with reference to the daily RRF_c is calculated for each compound as follows:

$$\%D = \left[\frac{(RRF_c - RRF_{meani})}{RRF_{meani}}\right] \times 100$$

Where:
$\%D$ = percent difference;
RRF_c = RRF of the compound in the continuing calibration standard; and
RRF_{meani} = mean RRF of the compound in the most recent initial calibration.

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The %D for each compound must be within ± 30% in order to proceed with the analysis of blanks and samples. The initial calibration mean RRF, the continuing calibration RRF and the %D (labeled "%Dev" in the software report) are calculated by the ChemStation software and reported using the "Evaluate Continuing Calibration" report form.

4.4 Blank Analysis

To monitor for laboratory contamination, a blank is analyzed with each sample analysis batch or at least once per 24 hours. The blank is analyzed after the continuing calibration and before any samples are analyzed. The blank is analyzed in the same manner as field samples and the same calculations apply. The calculations are outlined in Section 4.6, "Sample Analysis".

The area response of each IS added to the blank sample must be within +/- 40% of the area response of the current continuing calibration. The RT for each IS must be within ± 20 seconds (or 0.33 minutes) of the IS RT of the current continuing calibration. The blank must not contain any target compound/additional requested analyte at a concentration above 0.5 ppbv. It also must not contain any compounds with elution characteristics and mass spectral features that would interfere with identification of target compounds. The total level of VOC in the blank must not exceed 10 ppbv for Confirmatory Monitoring Samples.

Blank results are reported using the ChemStation "Quantitation Report" form. A summary of the blank results is reported on a form similar to the Blank Summary Form in Appendix A. A template of this form may be found in the "envchem\VOCDataPackages\VOCForms\VOCDP.xls" spreadsheet. The limits for IS response and RT are calculated on the "Internal Standard Area and Retention Time Summary" form in the same spreadsheet.

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Table 2: Contract Required Quantitation Limits for Compounds of Interest

Compound	CRQL (or MRL) (for Confirmatory Monitoring Samples) (ppbv)	CRQL (or MRL) (for Closed Room Samples) (ppbv)
Carbon Tetrachloride	2	500
Chlorobenzene	2	500
Chloroform	2	500
1,1- Dichloroethene	5	500
1,2- Dichloroethane	2	500
Methylene Chloride	5	500
1,1,2,2-Tetrachloroethane	2	500
Toluene	5	500
1,1,1-Trichloroethane	5	500
Benzene*	2	500
Trichloroethylene*	2	500
Tetrachloroethylene*	5	500
Chloromethane*	2	500
trans-1,2-Dichloroethylene*	2	500
1,2,4-Trimethylbenzene*	2	500
p,m-Xylene*	5	500
Trichloromonofluoromethane*	2	500

Note: * indicates Additional Requested Analytes

4.5 Laboratory Control Samples

The LCS and LCSD are analyzed at a rate of once per batch, or once each ten samples, whichever is more frequent. The LCS is a blank spiked with the compounds of interest. It is used to verify the instrument calibration and method accuracy and precision. The percent recovery of the LCS is used in evaluating method accuracy. A comparison of LCS/LCSD results is used in evaluating method precision. The LCS and LCSD are analyzed in the same manner as field samples and the same calculations apply. The calculations are outlined in section 4.6 Sample Analysis. In addition, the LCS percent recovery is calculated as follows:

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$$LCS\%recovery = \left(\frac{Conc_R}{Conc_S}\right) \times 100$$

Where:
$Conc_R$ = Concentration reported by analysis
$Conc_S$ = Concentration spiked

The duplicate precision is calculated as follows:

$$RPD = \left[\frac{(|LCDS_{conc} - LCSD_{conc}|)}{\frac{(LCS_{conc} + LCSD_{conc})}{2}}\right] \times 100$$

Where:
RPD = Relative Percent Difference
LCS_{conc} = LCS concentration
$LCSD_{conc}$ = LCSD concentration

The area response of each IS must be within ± 40% of the area response of the current continuing calibration. The RT for each of the IS must be within ± 20 seconds (or 0.33 minutes) of the IS RT of the current continuing calibration. The LCS % Recovery must be within ± 40% for all target compounds. The RPD must be 25% or less for all target compounds. In the case of a failure to meet these criteria, the cause of the failure shall be determined and corrected prior to continuation of analysis.

LCS results are calculated and reported using the "Laboratory Control Sample Data Sheet" in the Excel spreadsheet "J:\envchem\VOCDataPackages\VOCForms\VOCDP.xls". The IS area and RT limits are calculated and reported using the "Internal Standard Summary" report form in the same spreadsheet. The LCD/LCS duplicate precision is calculated and reported on the form "LCS Duplicate Precision" in the same spreadsheet. A copy of the LCS Summary Form and the LCS Duplicate Precision Form are in Appendices C and B.

4.6 Sample Analysis

Samples are analyzed as described in OC-PROC-003, "Gas Chromatography-Mass Spectrometry Analysis of Volatile Organic Compounds (VOC) in Ambient Air from Canisters at PPBV Concentration Levels".

Each analytical run must be checked for saturation. When a high concentration sample is analyzed such that the detector response on the quantitation report indicates saturation of the detector with a given ion or

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ions, this analysis must be followed by a blank analysis. If the blank analysis indicates a concentration of any compound of interest ≥ 0.5 ppbv or greater than 10 ppbv total VOC, the system must be decontaminated. Analysis cannot resume until a blank analysis verifies the system is free from contamination. Secondary ion quantitation is only allowed when interferences prevent use of the primary ion. The reasons for secondary ion quantitation must be documented in the final report.

4.6.1 Concentration

The following equation is used for calculating concentrations:

$$X_a = \frac{(A_x C_{is} DF)}{(A_{is} RRF)}$$

Where:
X_a = compound concentration, ppbv;
A_x = area of the characteristic ion for the compound to be measured;
C_{is} = concentration of the IS spiking mixture;
A_{is} = area of the characteristic ion for the specific IS;
RRF = average relative response factor from the initial five point calibration; and
DF = dilution factor.

The raw sample analysis data is calculated and reported using the "Quantitation Report" form in the ChemStation software, without inclusion of dilution factors. Dilution factors are included in the final reported sample concentrations in the Results Summary spreadsheet.

A summary table of all sample quantitation results shall be included in the client data package. Sample quantitation results shall be reported in the summary table on the basis of CRQL. Any results reported between the laboratory MDL and the CRQL shall be qualified as "J" values. Results below the MDL shall be reported as "U", not as a numerical value.

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4.6.2 Internal Standard Retention Time

The RT for each of the IS for each sample must be within +/- 20 seconds (or 0.33 minutes) of the RT of the corresponding IS in the current continuing calibration. The IS RT limits are calculated and reported using the "Internal Standard Summary" report form in the Excel spreadsheet "J:\envchem\VOC DataPackages\VOCForms\ VOCDP.xls".

4.6.3 Internal Standard Area

The areas for each of the IS for each sample must be within 60-140% of the area of the corresponding IS in the CCAL. The IS area limits are calculated and reported using the "Internal Standard Summary" report form in the Excel spreadsheet "J:\envchem\VOCDataPackages\VOCForms\VOCDP.xls".

4.6.4 Qualitative Analysis

The target compounds reported shall be identified based on two criteria: (1) elution of the sample component at the same GC RT as the standard component, and (2) comparison of the relative intensities of quantitation ions to qualifying ions as compared to the reference spectra.

4.6.5 Tentatively Identified Compounds

A library search shall be executed for non-target sample components with area responses greater than 10% of the area response of the nearest IS for the purpose of tentative identification. For this purpose the most recent release of the NIST library shall be used. The compounds reported shall be identified based on their visual comparison to the nearest library search compound. All significant ions present in the reference spectra must be present in the sample spectrum. Relative intensities of ions should agree between sample and reference spectra by visual comparison. Ions present in the sample spectra, but not in the reference spectra must be accounted for by the analyst. If in the technical judgment of the mass spectral interpretation specialist, no valid tentative identification can be made, the compound must be reported as unknown. If additional classification of the compound can be made, it should be included.

TICs are reported using the "Library Search Compound Report" form in the ChemStation software. The search criteria shall be set up so that a forward search of the NIST library of mass spectra is performed for all Chromatographic peaks greater than ten percent of

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the nearest IS peak. Quantitation estimates for TICs should be calculated relative to the nearest IS, if there are no interferences, and assuming an RRF of 1.0.

4.6.6 Duplicate Analysis

A sample duplicate is analyzed with each sample analysis batch or every ten samples, whichever is more frequent.

The duplicate precision is calculated as follows:

$$RPD = \left(\frac{(|Sample_{conc} - SampleDup_{conc}|)}{\frac{(Sample_{conc} + SampleDup_{conc})}{2}} \right) \times 100$$

Where:
RPD = Relative Percent Difference
$Sample_{conc}$ = Reported sample concentration
$SampleDup_{conc}$ = Reported sample duplicate concentration

The RPD must be 25% or less for all target compounds having concentrations greater than or equal to the lowest calibration level from the five-point initial calibration. The sample duplicate precision is calculated and reported in the "Sample Duplicate Precision Data Sheet" form in the Excel spreadsheet "J:\envchem\VOCDataPackages\VOCForms\VOCDP.xls". A copy of this form is in Appendix D.

4.7 Completeness

As required by the client Statement of Work, a Sample Completeness Log will be maintained to assure that the completeness of analysis of samples submitted is greater than 95%. Non-reporting of a sample may occasionally occur due to lost samples, handling errors, or rejection of analytical data due to incomplete or unresolved QA/QC issues. Non-reporting of sample data should be minimized whenever possible.

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5.0 QUALITY ASSURANCE

Quality assurance practices will be in accordance with the CEMRC Quality Assurance Plan (QAP) CP-QAP-004 and requirements specified in the client's SOW. In the case of a failure to meet all performance criteria, the cause of the failure shall be determined and corrected if possible and the batch rerun up to three times or until criteria are met. If the problem cannot be resolved, the sample data may not be reported.

5.1 Documentation

5.1.1 Sample Receiving and Tracking

Sample receiving and tracking will be documented on the "VOC Sample Receiving and Tracking Log" form (see Appendix E). The original form will be maintained in a Sample Receiving and Tracking Logbook. A copy of the form will be included in the client data package. Custody seals may be used at the discretion of the client.

5.1.2 Standards

All stock standards used to prepare calibration standards must be recorded in the "Organic Chemistry Standards Log" and must be assigned a unique identification number. The preparation of intermediate standards, used for calibration and calibration verification must be recorded in the "Organic Chemistry VOC Intermediate Standards Preparation Log". Documentation of standards is outlined in CP-PROC-009 "Standards Log-in and Preparation". The instrument log must contain enough information to identify the exact standard, stock or intermediate, used for calibration, calibration verification, internal/tuning, and laboratory control samples.

5.1.3 Canister Cleaning

Sample canisters are cleaned following successful analysis. A copy of the canister cleaning and/or certification documentation for each canister cleaned shall be maintained with the hardcopy of the canister cleaning data package. The originals shall be provided to the client along with cleaned canisters and chain of custody form. Details of the canister cleaning and certification process and documentation thereof are found in OC-PROC-002, "Preparation of Canisters and Sample Trains for Ambient Air Sampling".

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5.1.4 Client Data Package Contents

The client data package contents are specified by the Statement of Work. For the WIPP Volatile Organic Compound Monitoring Program, the data package shall include the following elements:

- Analysis Request Form (both client and CEMRC forms)
- Laboratory Narrative
- Sample Results Summary
- Sample Chromatograms and Quantitation Reports (including target compound scans)
- Sample Library Search Results
- BFB Tune Summary Report and Raw Data (for both initial calibration and sample runs)
- Initial Calibration Summary and Raw Data and IS %ARC report
- LCS and LCSD Summary Reports, Quantitation Reports and Raw Data (including target compound scans)
- Blank Summary Report, Quantitation Report and Raw Data (including target compound scans and library search reports)
- Continuing Calibration Verification Summary Report and Raw Data (including target compound scans)
- Internal Standards Area and Retention Time Summary
- Duplicate Precision Datasheets (Sample and LCS Duplicates)
- Standards Preparation Logs and Stock Standard Certificates
- Instrument Run Logs (for initial calibration and sample runs)
- Sample Preparation Logs
- Sample Receiving and Tracking Log
- Chain-of-Custody Forms
- Sample Receipt Discrepancy Form (if applicable)
- Non-Routine Events or Non-Conformance Form (if applicable)
- Data Review Checklist

All handwritten entries, such as standards preparation logs, run logs, etc. must include the signature of the analyst performing the task and the date it was performed.

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5.2 Holding Time

Humid air canister standards and samples are valid for 30 days from the date of preparation or collection. Results of analyses performed after the holding time shall not be reported. Analytical Data Reports for routine samples shall be submitted to the client within 30 calendar days of sample receipt. Data reports for rush samples shall be submitted within seven calendar days of sample receipt.

5.3 Data Review

Logbooks, sample custody, and analysis records will be reviewed routinely by the QA officer or designee. At a minimum, they will be checked for completeness and correctness.

A Volatile Organic Compounds Data Package Inventory Sheet shall be filled out to assure completeness of the data package. A copy of this form is included in Appendix F.

Data packages shall be reviewed for completeness and correctness prior to delivery to the client. Data package review shall be conducted by the laboratory supervisor or designee. Each data package shall include a completed "VOC Data Review Checklist" (see Appendix G). A minimum of ten percent of data packages will receive a QA review performed by a QA specialist or designee.

6.0 QUALITY CONTROL

6.1 Control Charts

The RPD for LCS/LCSD will be entered into a control chart to track laboratory precision. The %Recovery for LCS and LCSD will be entered into a control chart to track laboratory accuracy. Control charts are titled "LCS_ControlChart_VOCs" and are stored in the following folder: J:\VOCs_Data_Packages.

7.0 ELECTRONIC DATA DELIVERABLE

A summary electronic file must be submitted for each analysis batch. The contents and format of the EDD shall be specified by the client and shall not be changed except at the client's request. EDD shall be delivered to the client by e-mail as soon as they are validated.

8.0 RECORDS

All instrument printouts and reporting forms described in Section 2.0 must be stored in accordance with the client's Statement of Work and with CP-PROC-013,

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"Records Maintenance, Storage, and Establishment of a Filing System". All documents generated to support the WIPP Volatile Organic Compound Monitoring Plan shall be stored in a 1-hour fire rated container until an electronic archive file is prepared on CD. The container shall bear the UL label (or equivalent) certifying 1-hour fire protection. After preparation of the electronic archive file, the CD may be kept in the 1-hour fire rated container and the hardcopies may be filed in a designated records storage area. All raw or processed GC/MS data must be archived on the CEMRC network, stored on magnetic tape, or CD and kept for the duration of this program.

9.0 REFERENCES

CP-PROC-009 "Standards Log-In and Preparation"

EPA Method TO-14 "Determination of Volatile Organic Compounds (VOC) In Ambient Air Using Specially Prepared Canisters With Subsequent Analysis By Gas Chromatography", Compendium of Methods for the Determination of Toxic Organic Compounds in Air, Second Edition, January 1999.

EPA Method TO-15 "Determination of Volatile Organic Compounds (VOC) in Air Collected in Specially–Prepared Canisters and Analyzed By Gas Chromatography/Mass Spectrometry (GC/MS)", Compendium of Methods for the Determination of Toxic Organic Compounds in Ambient Air, Second Edition, January 1999.

OC-PROC-002 "Preparation of Canisters and Sample Trains for Ambient Air Sampling"

"Quality Assurance Project Plan for Confirmatory Volatile Organic Compound Monitoring", WP 12-VC.02, Current Revision.

"Volatile Organics Analysis of Ambient Air in Canisters", USEPA Contract Laboratory Program, VCAAO 1.0, Publication 9240.1-18, EPA/540/r/085, PB95-963524, December 1994.

"Volatile Organic Monitoring Compound Plan", WP12-VC.01, Current Revision.

"WIPP Hazardous Waste Permit, Attachment N", February 27, 2007.

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10.0 APPENDICES

Appendix A: Volatile Organics in Ambient Air-Canister; Blank Summary
Appendix B: Volatile Organics in Ambient Air-Canister; LCS Duplicate Precision Data Sheet
Appendix C: Laboratory Control Sample Data Sheet
Appendix D: Volatile Organics in Ambient Air-Canister; Sample Duplicate Precision Data Sheet
Appendix E: VOC Sample Receiving and Tracking Log
Appendix F: Volatile Organic Compounds Data Package Inventory Sheet
Appendix G: VOC Data Review Checklist

11.0 REVISION HISTORY

Revision #	Date	Description
0	11/26/02	Initial document, released as EC-PROC-027
1	12/19/03	Changed to conform to client SOW
2	02/21/06	Reviewed, with major re-write, released as OC-PROC-005
3	04/18/08	Reviewed and changed slightly to reflect current SOW requirements
4	05/05/10	Minor revisions and more definitions added.
5	05/17/11	Reviewed and changed as needed to reflect current SOW requirements and request of client via e-mail regarding additional requested analytes. Incorporated temporary deviations.

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Appendix A: VOLATILE ORGANICS IN AMBIENT AIR-CANISTER

BLANK SUMMARY

Client:_____ Contract:_____
Batch No.:_____

Blank ID:_____ Date Analyzed:_____ Instrument ID:_ _
Blank File ID:_____ Time Analyzed:_____ Column ID: RTX-1, 0.32mm id__

THIS BLANK APPLIES TO THE FOLLOWING SAMPLES:

	Sample No.	Laboratory ID			Analysis	
		Sample	File	Canister	Date	Time
1						
2						
3						
4						
5						
6						
7						
8						
9						
10						
11						
12						
13						
14						
15						
16						
17						
18						
19						
20						

Comments:

FORM II

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Appendix B: VOLATILE ORGANICS IN AMBIENT AIR-CANISTER

LCS DUPLICATE PRECISION DATA SHEET

Client:_____ Contract:_____
Batch No.:_____ Canister No.:_____
Lab File ID.:_____
Client Sample No.:_____ Lab Duplicate File ID:_____
Instrument ID:_____ _____ GC Column ID:____RTX-1, 0.32mm I.d.__

COMPOUND NAME	LCS Conc.	Duplicate LCS Conc.	RPD
Carbon tetrachloride			
Chlorobenzene			
Chloroform			
1,2-Dichloroethane			
1,1-Dichloroethene			
Methylene chloride			
1,1,2,2-Tetracholoethane			
Toluene			
1,1,1-Trichloroethane			
Benzene*			
Trichloroethylene*			
Tetrachloroethylene*			
Chloromethane*			
trans-1,2-Dichloroethylene*			
1,2,4-Trimethylbenzene*			
p,m-Xylene*			
Trichloromonofluoromethane*			

FORM X

Note: * indicates Additional Requested Analytes

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Appendix C: LABORATORY CONTROL SAMPLE DATA SHEET

Client:_____ Contract:_____
Batch No.:_____

Sample ID:_____ Date Analyzed:_____ Instrument ID:
Lab File ID:_____ Time Analyzed:_____ Column ID: RTX-1, 0.32mm id
Inj. Vol.(cm^3): _____

		Concentration (ppbv)		%	
CAS RN	COMPOUND NAME	Spiked	Reported	Recovery	Q

%Recovery QC Limits : 60-140%

LCS Recovery:_____outside limits out of _____total.

Comments:

_____
_____
_____
_____

FORM III

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Appendix D: VOLATILE ORGANICS IN AMBIENT AIR-CANISTER

SAMPLE DUPLICATE PRECISION DATA SHEET

Client:_____ Contract: _____
Batch No.:_____ Canister No.: _____
Lab File ID.:_____
Client Sample No.:_____ Lab Duplicate File ID:_____
Instrument ID:_____ _____ GC Column ID:____RTX-1, 0.32mm I.d.____

COMPOUND NAME	Sample Conc.	Duplicate Conc.	RPD
Carbon tetrachloride			
Chlorobenzene			
Chloroform			
1,2-Dichloroethane			
1,1-Dichloroethene			
Methylene chloride			
1,1,2,2-Tetracholoethane			
Toluene			
1,1,1-Trichloroethane			
Benzene*			
Trichloroethylene*			
Tetrachloroethylene*			
Chloromethane*			
trans-1,2-Dichloroethylene*			
1,2,4-Trimethylbenzene*			
p,m-Xylene*			
Trichloromonofluoromethane*			

FORM X

Note: * indicates Additional Requested Analytes

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Appendix E: VOC SAMPLE RECEIVING AND TRACKING LOG

Batch: _____ **Received By:**_____ **Date Received:**_____

___**Chain of Custody Present and Complete**
___ **Request for Analysis Form Present and Complete**
___ **Label/Seal Present and Complete**
___**Canister Valves Closed**
___ **Canisters Properly Labeled/Tagged**
___ **Canisters and Packaging in Good Condition**

Sample Tracking

Client Sample ID	Canister ID	Laboratory Sample ID	Sample Location	Sample Date	Pressure Upon Receipt	Analysis Completed Date	Canister Cleaned Date	Blank Check Date	Pressure Leak Check Date

Comments: _____

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Appendix F: VOLATILE ORGANIC COMPOUNDS DATA PACKAGE INVENTORY SHEET

Analysis of Air Samples Collected in Canisters

(USEPA Method TO-15)

Batch: _____

	Page Numbers: From	To
Title Page	_____	_____
1. Analysis Request Form and Laboratory Narrative	_____	_____
2. Sample Results and Raw Data	_____	_____
a. Sample Results Summary		
b. Chromatograms		
c. Library Search Results		
3. QC Results and Raw Data		
a. BFB Tune (Summary and Raw Data)	_____	_____
b. Initial Calibration (Summary and Raw Data)	_____	_____
c. Laboratory Control Sample (Summary and Raw Data)	_____	_____
d. Method Blank (Summary and Raw Data)	_____	_____
e. Continuing Calibration Verification (Summary and Raw Data)	_____	_____
f. Internal Standard Summary	_____	_____
g. Duplicate Results Summary (Sample and LCS Duplicates)	_____	_____
h. Standards Preparation Logs and Stock Standard Certificates	_____	_____
i. Instrument Run Logs	_____	_____
j. Sample Preparation Logs (Dilution Factors & Internal Std Add.)	_____	_____
4. Shipping and Receiving Documents		
a. Sample Receiving and Tracking Form	_____	_____
b. Chain-of-Custody Forms	_____	_____
c. Sample Receipt Discrepancy Form (if applicable)	_____	_____
5. Other Records		
a. Non-Routine Events or Non-Conformance Form (if applicable)	_____	_____
b. Data Review Check Sheet	_____	_____

Comments:

_____

_____

_____

Completed By: _____ Date Completed: _____

Signature: _____

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Appendix G: VOC DATA REVIEW CHECKLIST FOR MMDDYY_00X

_____Lab Narrative is correct (method description, receiving & analysis notes)
_____Unusual circumstances documented in notes section below
_____Client Sample Ids correct (check COC against sample Log in)
_____Sample Receiving and Tracking Log complete & correct
_____BFB Tune criteria met for each 24 hour analysis period
_____Initial Calibration data meets QC criteria
_____Continuing Calibration meets criteria for each 24 hours of analysis
_____Method blank results meets QC criteria for each 24 hours of analysis
_____LCS/LCS Dup, Sample/Sample Dup results met %RPD criteria
_____Hold time met for all samples
_____Correct amount of sample analyzed
_____Internal Standards RFs and RTs within QC limits
_____Dilution Factors correctly calculated
_____Peak integration correct
_____Retention Times verified
_____Spectral defense included where required
_____TICs resemble reference spectra and are consistently identified in duplicate samples
_____Hand entered data on Sample Results Summary, Blank Summary, Initial Calibration %ARC, LCS /LCSD Summaries, LCS/Sample Duplicate summaries, and Internal Standards Summary checked for accuracy
_____Hand entered data on Intermediate Standard Preparation Forms consistent with primary standard certificate of analysis and LCS Sample Data Sheets
_____VOC Data Package Inventory Sheet is complete

Notes:

_____
_____
_____
_____
_____
_____
_____
_____

Reviewed By: _____

Date Reviewed: _____

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TEMPORARY PROCEDURAL DEVIATION FORM

Carlsbad Environmental Monitoring & Research Center

Title: Receipt, Control, and Storage of Gas Samples in passivated Canisters	
Document No: OC-PROC-006-003-~~022106~~ 022004 2/8/11	**Implementing Program: Organic Chemistry**

Deviation:
Section 7.8: second line of the paragraph on page 5 of 6 read 'The sample tag and original COC will be returned or copied for placement in the project file.

It should read as follows: The sample tag will be discarded when appropriate (before canister cleaning), and original COC will be returned or copied for placement in the project file.

Justification: Based on the client's request via e-mail on Dec 16, 2011.

Valid for: Until Next Revision

Approved by: _____ **Responsible Manager** 02/08/2011 **Date**

STAFF TRAINING

I have reviewed and I understand the procedure deviation described above.

Print Name	Signature	Date

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TEMPORARY PROCEDURAL DEVIATION FORM

Carlsbad Envi

Carlsbad Environmental Monitoring & Research Center

Title: Receipt, Control, and Storage of Gas Samples in Passivated Canisters	
Document No: OC-PROC-006-003-022006	Implementing Program: Organic Chemistry

Deviation: On page 3, fifth paragraph reads OC-PLAN-001, Quality Assurance Project Plan for Analysis of Volatile Organic Compounds in Canister Samples

It should read as follows: OC-PLAN-001, Quality Assurance Project Plan for Analysis of Volatile Organic Compounds and/or Hydrogen and Methane in Canister Samples

On page 3, sixth paragraph reads OC-PROC-002 Preparation of Canisters for Ambient Air Sampling

It should read as follows: OC-PROC-002 Preparation of Canisters and Sample Trains for Ambient Air Sampling

On page 5, last paragraph reads . . . Preparation of Canisters for Ambient Air Sampling . . .

It should read as follows: . . . Preparation of Canisters and Sample Trains for Ambient Air Sampling . . .

On page 6, section 9.0 reads . . . Quality Assurance Project Plan for Analysis of Volatile Organic Compounds in Canister Samples, OC-PLAN-001.

It should read as follows: . . . Quality Assurance Project Plan for Analysis of Volatile Organic Compounds and/or Hydrogen and Methane in Canister Samples, OC-PLAN-001.

Justification: Titles of OC-PLAN-001 and OC-PROC-002 were changed after OC-PROC-006-003-022006 was issued.

Valid for: until next revision

Approved by: Ed Stone 5/14/10

Responsible Manager Date

STAFF TRAINING

I have reviewed and I understand the procedure deviation described above.

Print Name	Signature	Date

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CARLSBAD ENVIRONMENTAL MONITORING & RESEARCH CENTER (CEMRC)

RECEIPT, CONTROL, AND STORAGE OF GAS SAMPLES IN PASSIVATED CANISTERS

OC-PROC-006-003-022006-_ _ _

Effective Date: May 3, 2010

CARLSBAD ENVIRONMENTAL MONITORING & RESEARCH CENTER

NEW MEXICO STATE UNIVERSITY
1400 University Drive, Carlsbad, New Mexico 88220

TELEPHONE (575) 887-2759
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RECEIPT, CONTROL, AND STORAGE OF GAS SAMPLES IN PASSIVATED CANISTERS

1.0 PURPOSE

This procedure is used to ensure the integrity of gas samples from time of receipt at the CEMRC to time at which the sample canisters are cleaned, certified, and relinquished back to the client. It provides instruction for sample receipt, control, and storage, at the CEMRC, of gas samples in passivated (e.g. SUMMA®, Silonite™) canisters to fulfill requirements of the Waste Analysis and the VOC Monitoring Plans of the WIPP Hazardous Waste Facility Permit.

2.0 SCOPE AND LIMITATIONS

This procedure incorporates guidelines from EPA methods TO-14A and TO-15 and the WIPP Hazardous Waste Permit. It is applicable to gas samples in passivated canisters submitted to the CEMRC for VOC analysis and Hydrogen/Methane analysis. Canisters that are cleaned and certified by the CEMRC personnel are also subjected to the custody control and storage requirements that are imposed on actual gas samples.

All gas samples accepted for analysis at the CEMRC must be accompanied by COC documentation from the point of origin. When applicable, the CEMRC personnel will track sample movement through the CEMRC by following the guidelines established in Section 8.0 of Sample Tracking and Chain of Custody procedure, CP-PROC-001.

Approved analytical results must be obtained on a sample such that the time from date of sample receipt until analysis does not exceed thirty days. Shorter holding times may exist depending on contract requirements.

3.0 ACRONYMS

CEMRC - Carlsbad Environmental Monitoring and Research Center
COC - Chain of Custody
EPA - Environmental Protection Agency
NIST - National Institute of Standards and Technology
PPE - Personal Protective Equipment
VOC - Volatile Organic Compounds
WIPP - Waste Isolation Pilot Plant, Carlsbad, NM

4.0 REFERENCES

CP-PROC-001, Sample Tracking and Chain of Custody

CP-PROC-011, Training and Qualification

CP-PROC-012, Nonconformances and Non-Routine Events

CP-QAP-004, CEMRC Quality Assurance Plan

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DC-PROG-001, Document Control Program

EPA Compendium Method TO-14A, "Determination of Volatile Organic Compounds (VOCs) In Ambient Air Using Specially Prepared Canisters With Subsequent Analysis By Gas Chromatography", *Compendium of Methods for the Determination of Toxic Organic Compounds in Ambient Air*, Second Edition, January 1999.

EPA Compendium Method TO-15, "Determination of Volatile Organic Compounds (VOCs) In Air Collected in Specially Prepared Canisters and Analyzed By Gas Chromatography/Mass Spectrometry (GC/MS)", *Compendium of Methods for the Determination of Toxic Organic Compounds in Ambient Air*, Second Edition, January 1999.

OC-PLAN-001, Quality Assurance Project Plan for Analysis of Volatile Organic Compounds in Canister Samples

OC-PROC-002, Preparation of Canisters for Ambient Air Sampling

Quality Assurance Project Plan for Volatile Organic Compound Monitoring, WP 12-VC.02, Current Revision

Statement of Work for Volatile Organic Compound Analysis/Hydrogen Methane Analysis, Current Revision

WIPP Hazardous Waste Permit, Volatile Organic Compound Monitoring Plan, Attachment N

5.0 PERSONNEL RESPONSIBILITIES

Laboratory personnel should verify that this procedure is the latest available revision by checking the Organic Chemistry Laboratory Procedure Manual against the version in H:\document control\oc. If this procedure is found to be unsatisfactory in any way, notify the Laboratory Manager or designee. Personnel shall be trained, as described in CP-PROC-011, before commencing with the implementation of this procedure. The CEMRC Project Manager may designate a qualified individual or subject matter expert to provide and document the training. Personnel shall document any laboratory, equipment, and environmental irregularities, conditions adverse to quality, or other non-routine events on the appropriate nonconformance or non-routine event form as described in CP-PROC-012, when applicable.

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6.0 SAFETY

The minimum PPE required for entry into any room or area designated as a laboratory and/or any other rooms/locations where "laboratory use of hazardous chemicals" occurs is safety glasses with side shields.

7.0 PROCEDURE

Note: The CEMRC Project Manager shall designate individuals as authorized to receive samples.

7.1 Verify that each gas sample received is accompanied by COC documentation from the point of origin. *Note: All samples shipped to the CEMRC must be accompanied by a completed COC. The signature/initials on the shipping container should be consistent with a relinquishing signature on the COC form.*

7.2 Verify that a final relinquishing signature is present on the COC form.

7.3 Inspect the COC documentation for completeness, consistency and continuity.

7.4 Complete the transfer of custody by:

- Signing and recording date/time of physical transfer (receipt).
- Signing and recording date/time shipping container custody seal was broken (if present).

7.5 For each canister:

- Fill out the date and time of receipt and your initials as recipient on the canister identification tag.
- If a signed tamper-proof custody seal is present on the canister, ensure that it is intact and located on the sample canister where it would be apparent if the sample integrity were compromised and verify that the signature/initials on the sample canister custody seal is consistent with a relinquishing signature on the COC form.
- Verify that all canister valves and caps are in the closed position.
- Observe the overall condition of the sample canister and document disposition and any discrepancies, breakage, or tampering on a sample receiving and tracking log.

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- Prior to reading canister receipt pressures, allow the canisters to equilibrate to the laboratory's ambient temperature. Equilibration time improves accuracy of measurement and is particularly important when the canisters arrive at very cold or hot temperatures.

- Check for leaks in the canister valve by attaching a certified pressure/vacuum gauge to the canister without opening the canister valve. *Note: If a leak exists, the gauge readings will change over time. Record any leak comments on the sample receiving and tracking log.*

- Open the valve and determine the current reading of the pressure/vacuum gauge and write the value onto the canister identification tag and on the sample receiving and tracking log. *Note: If the readings are different by more than ±2 psi from the post-sampling readings recorded by the sample collector, the canister may be leaking. If possible, try to determine where the canister is leaking and tighten the appropriate fittings or consult with the CEMRC Project Manager before proceeding.*

7.6 For samples:

- Verify that each canister is tagged/labeled with an identification number

- Verify each canister is accompanied with a copy of the Request-for-Analysis form that identifies the sample canister number(s), sample type and type of analysis requested.

7.7 Maintain custody of the sample by maintaining physical possession until such time as the sample is analyzed or stored in a location meeting the following:

- Access-controlled to individuals authorized by the CEMRC Project Manager

- Free of solvents

- Maintained at an ambient temperature (between 0-40°C). *Note: Temperature will be monitored, as determined by the Project Manager, with a NIST-traceable thermometer.*

7.8 When appropriate (before canister cleaning), remove the sample tag from the canister to release it from storage and custody control requirements. *Note: The sample tag and original COC will be retained or copied for placement in the project files.*

7.9 Place a tag on the canister to show that it needs cleaning or proceed with cleaning and leak checking of the canister as described in Preparation of Canisters for Ambient Air Sampling OC-PROC-002. *Note: It is prudent to clean the canister*

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as soon as reasonably possible. Custody control and storage requirements listed in section 7.7 shall be maintained for cleaned canisters.

8.0 RECORDS

Sample Receiving and Tracking Form
Request for Analysis Form
Canister Identification Tag
Chain of Custody

9.0 QUALITY ASSURANCE

Quality assurance practices will be in compliance with the CEMRC Quality Assurance Plan, CP-QAP-004, and the Quality Assurance Project Plan for Analysis of Volatile Organic Compounds in Canister Samples, OC-PLAN-001.

10.0 REVISION HISTORY

Revision #	Date	Description
0	10/30/03	Initial Document, released as EC-PROC-030.
1	02/20/06	Biennial Review: incorporated temporary procedural deviation; removed the requirements for blank samples in shipping boxes since this would be done at the discretion of the client; added step for checking leaks in canister valve, and other minor changes for clarity. Procedure reissued as OC-PROC-006.
2	04/18/08	Biennial review, minor changes.
3	05/03/10	Biennial review, minor changes.

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CARLSBAD ENVIRONMENTAL MONITORING & RESEARCH CENTER (CEMRC)

ANALYSIS OF HYDROGEN AND METHANE IN PASSIVATED CANISTERS USING GAS CHROMATOGRAPHY WITH THERMAL CONDUCTIVITY DETECTION

OC-PROC-009-002-110907-_ _ _

Effective Date: September 8, 2011

CARLSBAD ENVIRONMENTAL MONITORING & RESEARCH CENTER

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ANALYSIS OF HYDROGEN AND METHANE SAMPLES IN PASSIVATED CANISTERS USING GAS CHROMATOGRAPHY WITH THERMAL CONDUCTIVITY DETECTION

1.0 PURPOSE

The following procedure describes the analysis of hydrogen and methane in passivated canisters by gas chromatography (GC) with thermal conductivity detection (TCD). This procedure is intended to be used by trained operators. It is not intended to provide step-by-step instructions on the operation of the instrument. The user is referred to operator's manuals for detailed operating instructions.

1.1 Scope and Limitations

Included in this procedure are instructions for the following activities:

- Preparation of calibration and quality control standards.
- Calibration and sample analysis.
- Data package generation, including appropriate quality assurance and quality control activities.

1.2 Process Overview

Samples are collected in SUMMA® passivated or equivalent canisters and are shipped or delivered to CEMRC together with appropriate chain of custody and request for analysis documentation. Upon receipt at CEMRC, samples are logged in and stored in an access-controlled area kept between 0-40°C until they are analyzed. Approved analytical results must be obtained on a sample such that the time from date of collection until analysis does not exceed thirty calendar days for normal analyses and ten calendar days for rush analyses. For Shaw samples, the turnaround time is 7 calendar days. Samples that are received at pressures < 4 psig or below ambient pressure must be pressurized to ≥ 4psig before analysis.

Hydrogen and methane are quantified by GC/TCD. Briefly, an aliquot of sample is introduced to an autosampler and travels through a transfer line to a fixed volume sample loop configured before the GC sample inlet. The volume in the loop is injected to the inlet and the analytical column. Ultra-high purity (UHP) nitrogen carries the sample into the column in a heated oven where temperature may be either held steady or increased at a specified ramp rate. The capillary column in which the stationary phase consists of small, porous particles composed of molecular sieve polymers or zeolites. Analytes gas and undergo a gas-solid adsorption/desorption process with the column stationary phase, which results in separation of analytes based on size and shape.

For hydrogen and methane analysis, the reference gas (i.e. UHP nitrogen) and the reference gas plus separated sample analytes (column effluent) pass through a TCD and differences in resistance between the two gas streams are measured. The TCD in use at CEMRC is designed with an electrically heated single filament whose temperature is at equilibrium at a given temperature when only the reference gas passes through the detector. The TCD detector pneumatics switches the column effluent and reference gas across the single filament at a rate of 5 Hz and conductivity measurements are acquired and separated into

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appropriate signals, (signal and reference). Column effluent containing gas analytes other than nitrogen causes the thermal conductivity and the specific heat of the gas surrounding the filament to change. The difference in the conductivity of the two signals is measured and appears as a peak on a chromatogram. Concentrations of analytes are interpolated from a 5-point calibration curve. Calibration standards are prepared from a certified gas standard by filling a clean canister with the hydrogen and methane standard and diluting with UHP nitrogen as necessary.

Data reporting will be compliant with client requirements and changes to data package contents may only be made with the client's permission. Personnel responsible for data package generation shall be familiar with the Agilent ChemStation Environmental Data Analysis software. All data reports shall receive independent technical review Quality control criteria for each target analyte shall be consistent with those listed in Table 1. Appropriate quality control samples shall be run with each sample batch, consistent with the requirements stated in Table 2.

1.3 Definitions

CAS Number - A Chemical Abstract Services (CAS) number is a unique numeric identifier for a chemical substance.

Continuing Calibration Standard (CCAL) - A calibration standard at or near the midpoint of the calibration curve (e.g. ICAL 4).

Contract-Required Quantitation Limit (CRQL) - The lowest concentration of a target analyte, which will be reported under a given contract. The CRQL for each target analyte is defined in the contract Statement of Work (SOW) and is the same as the Method Reporting Limit (MRL).

Dilution Factor - Multiplier which adjusts for the physical dilution of samples with UHP nitrogen.

Dynamic Calibration - Calibration of an analytical system using calibration gas standard concentrations in a form identical or very similar to the samples to be analyzed and by introducing such standards into the inlet of the analytical system in a manner very similar to the normal analytical process.

Electronic Data Deliverable (EDD) - An electronic file that summarizes the results of sample and QC standards for a batch of samples.

Gas Chromatography (GC) - An analytical technique used for separating compounds of interest.

Initial Calibration Standard (ICAL) - A specially prepared canister(s) containing a known concentration(s) of hydrogen and methane.

Laboratory Blank (LB) - A canister pressurized with UHP nitrogen used to monitor for contamination.

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Laboratory Control Sample (LCS) - A standard containing known concentrations of target analytes that has been prepared from a different source (or lot) of materials used to prepare calibration standards.

Laboratory Control Sample Duplicate (LCSD) - A duplicate analysis of the LCS that is used to verify the instrument calibration, quantitative analytical accuracy, and laboratory precision.

Method Detection Limit (MDL) - The minimum concentration of a target analyte that can be identified, measured, and reported with 99% confidence that the analyte concentration is greater than zero. In other words, the minimum concentration reliably detected by an instrument using a specific method.

Porous Layer Open Tubular (PLOT) - A type of gas chromatograph column.

ppmv - parts per million by volume, the concentration calculated as the volume of the target analyte multiplied by 10^6 and divided by the volume of the dilution gas.

psia- pounds per square inch absolute pressure, measured relative to vacuum. The psia unit of measure is not affected by local barometric pressure and is used by the Dynamic Diluter.

psig - pounds per square inch gauge pressure, measured relative to local barometric pressure.

Mass Selective Detector (MSD) or Mass Spectrometer (MS) - An analytical technique used for quantitative analysis and/or for the qualitative determination of the molecular composition of a sample. An MS consists of ionizing compounds to generate charged molecules, separation of ions based on their mass-to-charge ratio (*m/z*), and detection of the charge induced or current produced when ions interact with the detector's surface.

Method Reporting Limit (MRL) - same as CRQL.

Percent Difference (%D) - A method of comparing two values to one another in which the difference between an experimentally-determined value and a known value is divided by the known value and expressed as a percent.

Percent Recovery (%R) – A method of determining analytical accuracy in which the experimentally determined value is divided by the known value and expressed as a percent.

Percent Relative Standard Deviation (%RSD) - For an initial calibration curve, the standard deviation of the response factors (RF) at each calibration level over the mean of the response factors times one hundred. This serves as a measure of the linearity of instrument response over the calibration curve. Generated by Agilent ChemStation Environmental Data Analysis software on the Response Factor Report.

Quality Assurance (QA) - The systematic monitoring and evaluation of a project to ensure that standards of quality are being met.

Quality Control (QC) - The process of ensuring a certain level of quality in analytical measurements.

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Response Factor (RF) - The response (peak area or height) of a target analyte to the known concentration of the target analyte. Generated by Agilent ChemStation Environmental Data Analysis software on the Response Factor Report.

Relative Percent Difference (RPD) - A method of comparing two duplicate measurements and a measure of precision. The equation used for calculating RPD is shown in section 4.13.

Retention Time (RT) - The time from the start of the GC analysis run until the center of the peak of the target analyte reaches the detector. The retention time used for analyte identification is the mean of the retention times of the five calibration levels. This mean RT is manually calculated and manually entered into the quantitation database.

Statement of Work (SOW) – A written document describing the agreement of the client and contracted laboratory. The SOW includes, but is not limited to, technical tasks, quality assurance requirements, data reports and deliverables, and other considerations.

Thermal Conductivity Detector (TCD) - An analytical detector commonly used with GC that senses changes in the thermal conductivity of a gas stream containing analytes in relation to a reference gas.

Ultra High Purity (UHP): A grade of reagent gas certified as free of impurities.

Volatile Organic Compounds (VOC) – An organic compound that has a high enough vapor pressure to be in the gas phase at ambient temperature and pressure.

Table 1: Target Analyte List

Analyte	CAS #	Precision[a] (RPD)	Accuracy[a] (%R)	CRQL	Completeness (%)
Hydrogen	1333-74-0	≤ 25	70-130	As defined in SOW	95
Methane	74-82-8	≤ 25	70-130	As defined in SOW	95

a. Criteria apply to concentrations that are greater than or equal to the CRQL.

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Table 2: Summary of QC Samples and Frequencies for Hydrogen & Methane Analysis

QC Sample	Minimum Frequency	Acceptance Criteria
Continuing Calibration Standard	Every 24 hours of operation	≤30 %D
Laboratory Blank	Every 24 hours of operation	Target analytes ≤ MDL for GC/TCD
Laboratory Control Sample	One (1) per analysis batch	%R of ± 30%
Laboratory Control Sample Duplicate	One (1) per analysis batch	%R of ± 30%, RPD ≤ 25
Sample Duplicate [a]	One (1) per analysis batch [a]	RPD ≤ 25[a]
Performance Samples	Samples and frequency per SOW	Samples and frequency per SOW
Method Detection Limit	Every 12 months	See Table 1

a. Criteria apply to concentrations that are greater than or equal to the CRQL.

2.0 REQUIREMENTS

2.1 References

40CFR, Part 136, Appendix B, Definition and Procedure for the Determination of the Method Detection Limit, Rev.1.11

HP 6890 Series Gas Chromatograph Hardware Manual. Hewlett Packard Company, Wilmington, Delaware, Second Edition, December 1997.

SM-03, Chemical Hygiene Plan

CP-PROC-009, Standards Log-in and Preparation

OC-PROC-002, Preparation of Canisters and Sample Trains for Ambient Air Sampling

OC-PROC-006, Receipt, Control, and Storage of Gas Samples in Passivated Canisters

CP-PROC-017, Procurement of Items and Services

2.2 Training Requirements

This procedure is intended for use by personnel trained in the use of GC/TCD. It is assumed that instrument operators are familiar with the software used to control the GC/TCD and autosamplers. Personnel responsible for data package generation shall be familiar with the Agilent ChemStation Environmental Data Analysis software.

2.3 Equipment List

2.3.1 Gas Chromatograph/Thermal Conductivity Detector

The Agilent 6890N GC/TCD instrument as configured at CEMRC consists of three major components: the gas chromatograph (including the column and oven), the TCD, and the MSD. The MSD portion of the configuration will not be utilized in

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this procedure. An equivalent GC/TCD may be used for analysis provided that the system is capable of meeting all performance and QC criteria specified within this procedure.

2.3.2 Entech 7032A-L MiniCan™ Autosampler

The Entech 7032A-L MiniCan™ Autosampler can introduce up to 21 samples from 400 mL or 1000 mL MiniCans™ or 3 samples from 6 L canisters into the GC. Autosamplers may be plumbed in series so that up to three autosamplers may be attached to one GC/ TCD. Autosamplers are also configured to prepare calibration standards as described in Section 4.2. Autosamplers are controlled by the Entech control software. Equivalent autosamplers may be used, provided that they are capable of meeting all performance and QC criteria specified within this procedure, when properly installed.

2.3.3 Entech 4600A Dynamic Diluter

The Entech 4600A Dynamic Diluter is capable of diluting gas standards either by dynamic dilution or by static pressure dilution. The samples and standards covered by this procedure will be diluted by static pressure dilution.

2.3.4 Calibrated Thermometer(s)

A thermometer with a valid, traceable calibration shall be used to monitor the temperature in areas used for sample storage. Thermometers used in sample storage areas shall have a function which records maximum and minimum temperatures.

2.3.5 Stainless steel passivated SUMMA® or equivalent canisters (e.g. Silonite®, TO-Can™)

Standard and sample canisters are prepared for use as specified in OC-PROC-002. Cleaned and certified stainless steel canisters, typically 400-6000 mL capacity, are required for standard, blank, and laboratory control sample preparation. Canisters are fitted with stainless steel quick connect fittings for attachment to the autosampler. At least 7 canisters are required for standard preparation.

2.3.6 Calibrated Pressure Gauge and Vacuum Gauge

A pressure gauge is used to confirm above-ambient pressures in canisters. Vacuum gauge is used to confirm sub-ambient pressures in canisters.

2.4 Primary Standards

All primary standards shall be purchased from a qualified vendor in accordance with CP-PROC-017, "Procurement of Items and Services".

Primary standards shall be received and documented according to CP-PROC-009, "Standards Log-in and Preparation". Purchased standards are assigned a unique standard identification number upon receipt. This number is used to identify the standard on all

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standard preparation documentation in which that standard is used to prepare an intermediate or working standard. All purchased standards must be accompanied by a certificate of analysis.

The following is a list of primary standards used in this procedure:

- Certified hydrogen and methane gas calibration standards, balanced with nitrogen.
- Certified hydrogen and methane gas calibration standard, balanced with nitrogen from an independent source (i.e. different cylinder#). This standard is used for the laboratory control sample (LCS).

2.5 Precautions and Limitations

2.5.1 Interferences

Since TCD exhibits universal response and detects all gas analytes in the carrier gas, any analyte having the same retention time as a target analyte may produce a false positive. One example is helium. Helium has a retention time near hydrogen. It is unlikely that large amounts of helium will be present in samples, but the current instrument configuration at CEMRC utilizes helium as the carrier gas for the attached MSD. As a result, retention time windows will be set at ± 0.075 minutes. Careful consideration should be made before identifying an analyte as a target analyte when it has a slightly different retention time than the mean retention time of the five calibration levels for that analyte. Choosing an appropriate GC column and shifting the retention times by changing the GC method parameters may also help to eliminate false positives.

The TCD can become contaminated with deposits via column bleed or dirty samples. Signs of contamination include a wandering baseline, increased noise levels, or changes in response on a checkout chromatogram (Agilent 6890 TCD checkout procedure). Thermal cleaning or baking out the detector will often clear out the contamination. The tungsten-rhenium TCD filament has been passivated to protect against oxygen damage, but chemically active compounds such as acids and halogenated compounds may alter filament resistance and permanent changes to detector sensitivity. Avoidance of acidic or halogenated compounds or frequent replacement of the filament can prevent such interferences.

2.5.2 Safety

Samples analyzed by this method may contain known carcinogens in the form of VOCs and other non-target compounds. Exposure to these compounds will be minimal as long as the operating procedure is followed. Each individual working in the laboratory is expected to develop safe chemical hygiene habits aimed at the reduction of chemical exposures to themselves and coworkers.

The standards and nitrogen carrier gas used for analysis are stored at high pressure. Large cylinders must be secured to prevent tipping. Use protective caps when moving large gas cylinders.

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The current instrument configuration has many heated zones and care must be taken in order to prevent serious burns.

2.6 Prerequisite Actions

2.6.1 Check pressure in all compressed gas cylinders, including GC carrier gases, TCD reference gas, diluent gases, and standard gases to assure that no cylinders will need replacement while performing the procedure. When changing out the cylinders for the carrier gas and reference gas, cool down the appropriate heated zones. Make certain carrier gas regulators are set at approximately 80 psig to assure proper carrier gas flow.

2.6.2 Observe the oil level in the view window on the foreline pump to assure that it is at a proper working level. If the oil level is low or the oil appears discolored, perform a proper shutdown of the instrumentation and change the oil before operating the instrument.

2.6.3 Check the exhaust ventilation for proper air flow.

2.6.4 Make certain that personal protective equipment is available and used when appropriate.

2.6.5 Ensure that all thermometers to be used are in proper calibration.

2.6.6 Check the expiration dates on all standards to be used to ensure that they are not expired.

2.6.7 Check that cleaned canisters are available for all standards that must be prepared.

2.6.8 Check the pressure in working standards to assure that they are within operational requirements prior to beginning any analysis.

2.6.9 Check the maintenance log to be certain that no routine maintenance is due prior to analyzing samples.

3.0 RESPONSIBILITIES

CEMRC Laboratory personnel should verify use of the most recent procedure revision by checking the Organic Chemistry Laboratory Procedure Manual against the version in H:\document control\oc. The latest revision of this document shall be in the laboratory at all times. If this procedure is found to be unsatisfactory in any way and a procedure revision is required, notify the Project or Laboratory Manager or designee. This procedure is to be performed only by personnel trained and qualified in its use.

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4.0 PROCEDURE

4.1 Sample Receipt, Control, and Storage

As specified in OC-PROC-006: Receipt, Control, and Storage of Gas Samples in Passivated Canisters. The client shall provide appropriate Request for Analysis and Chain of Custody documentation for samples submitted.

4.2 Calibration Standard Preparation

4.2.1 Attach copper tubing between the primary standard cylinder regulator and an Entech autosampler. Open the cylinder valve. Adjust the regulator pressure accordingly based on intermediate standard canister size.

4.2.2 Open the appropriate Entech autosampler software and press the "Fill" button to enter the canister pressurization controls screen. Set the initial flush time to 1 minute and press "Go". After the initial 1 minutes flush, the flush time should be set to 0.1 minutes for flushes between dilutions.

4.2.3 Attach a quick connect fitting to open the pressurization port to atmospheric conditions. Press the "Flush" button on the autosampler to purge with the hydrogen and methane standard.

4.2.4 Take a cleaned and evacuated canister and attach the canister to the pressurization port.

4.2.5 Set the Entech software to dilute to a target pressure. Open the canister and press the pressurize button.

4.2.6 After the software shows the canister is pressurized, close the canister valve and remove it from the pressurization port.

4.2.7 If necessary, dilute the standard as described in section 4.6 using either the Dynamic Diluter or the fill port on the autosampler and allow an equilibration time of at least 10 minutes between serial dilutions.

4.2.8 Document the calibration standard preparation. The five standards need to be assigned intermediate standard identifications in accordance with CP-PROC-009 and added to the "Standards Logbook".

4.2.9 Label the canisters as calibration standards, along with the intermediate ID, date prepared, preparer's initials, expiration date and level of concentration. The calibration standards are valid until the expiration date of the primary standard.

4.2.10 Repeat steps 4.2.2 to 4.2.9 for the remaining four standards to produce a total of five standards at different levels of concentration. Close all primary standard cylinder valves when not in use. Examples of standard levels are listed in Table 3. Note: The lowest standard level should be ≥50 ppmv. The TCD is not sensitive enough to accurately detect at lower than 50 ppmv.

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Table 3: Calibration Standard Levels (Example)

Standard Level	Prepared from Primary Standard	Dilution
(ICAL1) ~56 ppmv	1000 ppmv (0.1%)	18x
(ICAL2) ~125 ppmv	1000 ppmv (0.1%)	8x
(ICAL3) ~250 ppmv	1000 ppmv (0.1%)	4x
(ICAL4) ~500 ppmv	1000 ppmv (0.1%)	2x
(ICAL5) ~1000 ppmv	1000 ppmv (0.1%)	1x

4.3 Laboratory Control Standard Preparation

The LCS requires preparation of a gas standard from a secondary source in the same manner described in section 4.2.

4.4 Laboratory Blank Preparation

4.4.1 Turn on the Entech 4600A Dynamic Diluter if not already on.

4.4.2 Open the UHP nitrogen cylinder valve. Adjust the regulator pressure accordingly based on laboratory blank canister size.

4.4.3 Open the 4600A software and press the "Flush" button to enter the canister pressurization controls screen. Set the flush time to 1 minute and press "Go".

4.4.4 Press the "Flush" button on the diluter to purge it with UHP nitrogen.

4.4.5 Take a cleaned and evacuated canister and attach the canister to the fill line.

4.4.6 Set the Entech software to dilute to target pressure. Open the canister and press the pressurize button on the Dynamic Diluter.

4.4.7 After the software shows the canister is pressurized, close the canister valve and remove it from the fill line.

4.4.8 Close the UHP nitrogen valve.

4.4.9 Label the canister as a laboratory blank or LB, along with the date prepared, preparer's initials, and UHP nitrogen lot#.

4.5 Preparation of MDL Samples

The ICAL1 standard can be used for the MDL sample or it can be prepared separately in the same manner as the ICAL1 calibration standard.

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4.6 Sample Preparation

Prior to analysis, all samples must be at least 4 psig pressure. Samples that are received at pressures < 4 psig or below ambient pressure must be pressurized to ≥ 4psig before analysis. Samples with analytes whose concentrations exceed the calibration curve will have to be diluted and reanalyzed. Sample dilutions will be recorded on a sample preparation form. If it is necessary to release pressure in a canister before performing a dilution, release excess sample into the exhaust ventilation duct in order to minimize personnel exposure to potentially hazardous substances. Sample dilutions are performed as follows:

4.6.1 Turn on the Entech 4600A Dynamic Diluter if not already on.

4.6.2 Open the UHP nitrogen cylinder valve. Adjust the regulator pressure accordingly based on sample canister size. For a 6L canister, 55-65psig is an appropriate regulator pressure.

4.6.3 Open the 4600A software and press the "Flush" button to enter the canister pressurization controls screen. Set the flush time to 1 minute and press "Go". After the initial 1 minute flush, the flush time should be set to 0.1 minutes for flushes between dilutions.

4.6.4 Press the "Flush" button on the diluter to purge it with UHP nitrogen.

4.6.5 Attach the canister to the fill line.

4.6.6 Set the Entech software to perform the appropriate relative dilution of "2x" or "3x". Open the canister and allow the initial pressure to equilibrate. Then press the pressurize button on the Dynamic Diluter.

4.6.7 After the software shows the canister is pressurized, close the canister valve and remove it from the fill line.

4.6.8 Record appropriate information for entry onto a sample preparation form.

4.6.9 Close the UHP nitrogen valve.

4.7 Initial Calibration

The calibration is performed by analysis of five calibration standards. The method uses the response factors (area count/concentration) of the calibration standards to create a new calibration. The instrument is calibrated initially and whenever the operational QC checks are not within acceptable limits.

4.7.1 Initial Calibration (ICAL) Data Acquisition

4.7.1.1 Make certain that the pressure in each ICAL standard canister is at least 4 psig and that the standards have not expired. Load the five ICAL canisters onto the autosampler(s).

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4.7.1.2 Make certain that the calibration standards are properly recorded in both the GC and autosampler sequence tables and that the specified autosampler positions are correct.

4.7.1.3 Start the sequence in both the GC and autosampler software when temperature zones reach their designated settings.

4.7.2 Initial Calibration Data Analysis and Response Update

4.7.2.1 After the analysis of all initial calibration standards is complete, open the "Environmental Data Analysis" window. Load the data analysis method which includes the most recent calibration responses (filename "MMDDYYgas"). Clear all the calibration responses.

4.7.2.2 In the "Execute" line, type *"enhanced 2"* and hit *"Execute"*. On the toolbar of the enhanced data analysis screen, go to *"Calibration"* and select *"Edit Compounds"*. When *"Edit Compounds"* screen comes up, select the first analyte and click on *"View"*. This will bring up the Page 1 of the chosen analyte. For hydrogen and methane, make sure that the Quant Signal in the box *"Signals to Be Used for Quantitation"* is *"GC Signal 1"*.

4.7.2.3 Click on "Page 3". This will bring up the Calibration level ID and Concentration screen. For hydrogen and methane, in the "Conc." column, fill out the appropriate concentrations for "Calibration Level ID". Save the changes to each analyte.

4.7.2.4 Save the method as "MMDDYYgas.M". For example, "012810gas.M" is the method saved on January 28, 2010.

4.7.2.5 Exit the enhanced data analysis screen by typing "enhanced" in the Execute line and select "Execute".

4.7.2.6 Load the method just created.

4.7.2.7 Open the first calibration data file (ICAL 1). Calculate and generate the quantitation report. Then enter "Qedit" mode and check to be certain that each peak was correctly integrated. Exit Qedit, saving changes to the quantitation.

4.7.2.8 Update the initial calibration for the first level.

4.7.2.9 Repeat steps 4.7.2.7 and 4.7.2.8 for each calibration level.

4.7.2.10 Calculate the mean retention time for each target analyte from the five calibration levels and manually enter the times into the quantitation database.

4.7.2.11 Repeat step 4.7.2.2 to make sure "Signals to Be Used for Quantitation" is "GC Signal 1".

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4.7.2.12 After updating the responses for all the calibration levels, display the response factors for the calibration. Print out the response factor report.

4.7.2.13 Print out the hydrogen and methane calibration curves, then save the method.

4.7.3 Initial Calibration Acceptance

The relative standard deviation %RSD is the standard deviation divided by the mean of the response factors for the five calibration standards and must be ≤ 30%. The %RSD is calculated by the ChemStation Software and reported in the response factor report.

For hydrogen and methane, the five initial calibration standards results will be fitted using the least square method. The correlation coefficient (r^2) value must be greater than 0.990 for the calibration to be acceptable. Qualitatively, hydrogen and methane are identified if there are peaks within the RT window.

Provided all acceptance criteria listed in Table 4 below are met, the initial calibration data may be accepted and analysis may proceed. If the criteria are not met, the cause must be determined and corrected.

Table 4. Calibration Acceptance Criteria

Procedure	Frequency of Procedure	Acceptance Criteria
5-pt initial calibration for hydrogen and methane	Initially, and as needed	• %RSD ≤ 30 • Retention times of each ICAL level must be within ± 0.075 minutes from the initial calibration mean RT • Linear response correlation coefficient (r^2) must be greater than 0.990

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Calculations Associated with Initial Calibration

The RF for each initial calibration level is calculated with the equation:

$$RF = (A_s)/(C_s)$$

Where:

A_s = Peak area of the target analyte counts (Response)
C_s = Concentration of the analyte (ppmv)

The percent relative standard deviation (%RSD) for each analyte is calculated as shown in equation below:

$$\%RSD = (s/x)*100$$

Where:

s = standard deviation of response factors for the analyte (five initial RFs from the calibration)
x = mean RF of the analyte (average of the five RFs from the initial calibration)

Initial calibration curves are constructed in the ChemStation software by using a least squares linear regression to plot area responses versus concentration of analyte for all calibration standards analyzed. The curve fit is *Linear Regr, Force (0,0). Inverse of conc* will be used for the data point weight for linear regressions.

A simplified calibration equation that is forced at zero, but not weighted for the inverse of concentration is listed below:

$$A = mC + b$$

Where:

A	=	Total area response for analyte
m	=	Slope of line (proportionality constant)
C	=	Amount (ppmv) of analyte
b	=	Intercept (regression constant, forced at zero)

4.8 Continuing Calibration Standard

The continuing calibration verification is performed every 24 hours of operation and the acceptance criteria must pass the requirements described in Table 5 below. The CCAL can be the midpoint calibration standard (ICAL 4) or can be prepared as a separate standard at or near the same level of the ICAL 4 standard.

For the CCAL, the following QC criteria must be met:

For hydrogen and methane, the RT must be within ± 0.075 minutes of the initial calibration mean RT. In addition, concentrations must agree within ± 30% of the known standard concentration. If the CCAL does not pass acceptance criteria, a second continuing

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calibration standard may be run. If the second sample also fails to meet the criteria, perform another five-point instrument calibration prior to proceeding with analysis. The CCAL may be retried a third time only in if an obvious reason for the failure of the standards to pass criteria is observed.

Calculations associated with evaluation of the CCAL are detailed below:

For hydrogen and methane, %D is calculated as follows:

$$\%D = \left[\frac{(RF_c - RF_i)}{RF_i}\right] \times 100$$

Where:

RF_c = RF of the analyte in the CCAL standard

RF_i = mean relative response factor of the analyte in the most recent initial calibration.

The %D for each analyte must be within ± 30% in order to proceed with the analysis of blanks and samples. The initial calibration mean RF, the continuing calibration RF and the %D (labeled "%Dev" in the software report) are calculated by the ChemStation Environmental Data Analysis software and reported using the "Evaluate Continuing Calibration" report.

Table 5: Summary of Continuing Calibration Acceptance Criteria

Procedure	Frequency of Procedure	Acceptance Criteria
Continuing Calibration for hydrogen and methane	Every 24 hours of operation	• %D or %Dev must be ≤ 30%. • Retention times must be within ± 0.075 minutes from the initial calibration mean RT.

4.9 Laboratory Blank

Prior to sample analysis and following the continuing calibration evaluation, a laboratory blank is analyzed. The analysis results of the laboratory blank must be less than or equal to the MDLs for hydrogen and methane in order for the system to pass the certification.

4.10 Determination of Method Detection Limits

MDLs are determined as follows. Seven replicate analyses of a low level standard containing target analytes at or near the expected detection limit are analyzed and the standard deviation determined for each target analyte. The MDL is the value obtained when this standard deviation is multiplied by 3.143 (the "student's t value" appropriate for a 99% confidence level for six degrees of freedom). This MDL determination method is based on that described in 40 CFR Part 136, Appendix B. Actual MDL will be reported as calculated. Samples will be flagged "U" for target analytes that have concentrations below MDL before

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applying any multipliers. MDLs are determined prior to hydrogen and methane analysis for laboratory commissioning and at least once per year thereafter, or following any major repairs to the instrument or significant changes to the analysis method. Electronic copies of current MDL data and calculations will be maintained on CEMRC's network (H:\OC DQOs).

4.11 Laboratory Control Sample

An LCS and LCSD are run for each analysis batch. The LCS is prepared using a different source for the target analytes than the calibration standards. Recovery of the target analytes (%R) for the LCS must be within ± 30% of the known spiked concentration of the canister. The LCS/LCSD results are acceptable provided that the RPD is ≤25. If the recovery or RPD are outside the acceptance windows, determine the cause and rerun the LCS/LCSD prior to continuing with the sample batch analysis. For hydrogen and methane, the RT must be within ± 0.075 minutes of the initial calibration mean RT.

Calculations associated with evaluation of the LCS and LCSD are detailed below:

Percent Recovery:

$$\%R = \left(\frac{Conc_R}{Conc_S}\right) \times 100$$

Where:

$Conc_R$ = Concentration reported by analysis
$Conc_S$ = Concentration spiked

Relative Percent Difference:

$$RPD = \left[\frac{|LCS - LCSD|}{\left(\frac{(LCS + LCSD)}{2}\right)}\right] \times 100\%$$

4.12 Sample Batch Procedures

Sample batch analysis may not proceed until all QA criteria are met. The Continuing CCAL and LB must be run prior to any sample analysis and repeated every 24 hours of operation. A LCS and a LCSD must be run once for each sample batch.

4.12.1 Sample Data Quantitation

For all samples in the batch, detected target analytes must meet the following criteria:

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For hydrogen and methane, the RT must be within ± 0.075 minutes of the initial calibration mean RT.

Calculations for Sample Quantitation

Concentrations of the introduced sample are calculated by the ChemStation software. The curve fit is *Linear Regr, Force (0,0)*. Inverse of concentration is used for the data point weight for linear regressions. Any prior dilutions resulting from off-line static pressure dilutions are recorded on the "Sample Preparation Form", where a total dilution factor is determined. The total dilution factor for a sample is entered in the "Result Summary" section of the verified spreadsheet (HMDP.xls) and is used to calculate the concentration of analytes in the diluted samples.

A simplified sample concentration equation that is forced at zero, but not weighted for the inverse of concentration is listed below (results will vary slightly from what is calculated in ChemStation):

$$C = ((A - b)/m) \times DF$$

Where:

Cx	=	Concentration (ppmv) of analyte
A	=	Total area response for analyte
b	=	Intercept (regression constant, forced at zero)
m	=	Slope of line (proportionality constant)
DF	=	dilution factor

4.13 Duplicate Sample Analyses

For each sample batch, a sample duplicate must be analyzed. If none of the samples in a sample batch contain a target analyte at a concentration ≥CRQL, the LCSD will be counted as the sample duplicate. The duplicate precision, defined by the RPD, must be less than or equal to 25%.

Relative Percent Difference (RPD) Calculations

$$RPD = \left[\frac{|Sample - SampleDup|}{\left(\frac{(Sample + SampleDup)}{2} \right)} \right] \times 100\%$$

Where:

Sample = Sample concentration result
SampleDup = Sample duplicate concentration result

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4.14 Performance Samples

When available, independent quality assurance samples will be obtained and analyzed to verify the accuracy of both the instrumentation and the analyst. Both blind (obtained from an outside source, true values not known at the time of analysis) and reference samples (samples obtained from an outside source with known true values, or alternatively, internally prepared standards) can be used to perform this function. The percent recovery for the audit standards must fall within ± 30%. If the recovery results for the performance sample analyses are not acceptable, an investigation must be performed to determine the source of the conflicting result(s) and corrective action initiated.

4.15 Instrument Shut Down

Ensure that the GC and autosamplers return to standby mode when analysis is complete. The vacuum, carrier gas, and GC oven systems should remain on, when possible. If complete shutdown is necessary, refer to the operator's manual for all shut down requirements. NOTE: The MSD is configured as a component of the instrument used for hydrogen and methane analysis. The MSD source is heated during operation of the instrument. Do not vent the MSD for any reason without first completing a "Vent" cycle from the ChemStation software to avoid damaging the mass selective detector. Venting a hot source can cause oxidation of delicate components within the ion source.

5.0 QUALITY ASSURANCE

5.1 Documentation

5.1.1 Standards

All stock standards must be recorded in the Organic Chemistry Primary Standards Logbook upon receipt and must be given a unique identification number. All intermediate standards prepared for use in calibration and QC checks must be documented in the VOCs Intermediate Standards Preparation logbook. Documentation of standards is outlined in CP-PROC-009. Instrument raw data or run logs must contain enough information to identify the exact standard, stock or intermediate, used.

5.1.2 Maintenance Logs

Maintenance logs will be kept up to date. All routine maintenance operations will be documented as well as instrument service. All service entries will include malfunction symptoms, a description of repairs made, and a description of how return to acceptable operation was determined. The routine maintenance schedule provided by the manufacturer may be used to document performance of routine maintenance procedures. The instrument manufacturer will provide documentation of repairs performed by service engineers. This will be incorporated into the maintenance log.

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5.1.3 Instrument Analysis Logs

Analysis logs will be maintained and will contain at a minimum a list of samples analyzed, sample injection ID, autosampler inlet, calibration and LCS standards identification numbers, the date analyzed and the data file storage information. Any problems encountered during analysis or reruns required will also be noted. Analysis logs will be updated when using the instrument for calibration, MDLs, and sample analysis.

5.1.4 Sample Receiving

Samples will be received and documented in accordance with OC-PROC-006.

5.1.5 Canister Cleaning

Sample canisters are cleaned following successful analysis and reporting of results. The effectiveness of the cleaning procedure is demonstrated by analyzing a blank sample from one cleaned canister of the cleaning batch. A copy of the canister cleaning documentation for each canister cleaned shall be maintained with the hardcopy of the canister cleaning data package. The originals shall be provided to the client along with cleaned canisters and chain of custody form. Details of the canister cleaning and certification process and documentation thereof are found in OC-PROC-002, "Preparation of Canisters and Sample Trains for Ambient Air Sampling."

5.1.6 Data Reporting

The contents and format of hydrogen and methane analytical data packages will be in accordance with the current Statement of Work for Volatile Organic Compound/Hydrogen and Methane Analysis and/or specified by the client.

Instrument output: The RF for each calibration level, the mean or average RF for the calibration, and the %RSD for the calibration for each compound are calculated by the ChemStation software and reported using the "Response Factor Report" form. The continuing calibration RF and %D (labeled "%Dev" in the software report) are calculated by the ChemStation software and reported using the "Evaluate Continuing Calibration" report form. CCAL, LB, LCS, and LCSD concentrations are reported using the ChemStation "Quantitation Report" form. Sample concentrations are reported using the ChemStation "Quantitation Report" form without correcting for dilution factors. *Note: Dilution factors are factored in within the "Results Summary" worksheet in the most current version of the file "envchem\VOCs_Data_Packages\VOCsForms\ HMDP.xls".*

Custom Reporting Forms: The format of reporting forms is flexible, but must be acceptable to the client. The reporting forms acceptable for use are in the current version of the file "envchem\VOCs_Data_Packages\VOCsForms\HMDP.xls". This workbook includes worksheets for calculating and/or reporting the CCAL %Recovery, Blank Summary information, LCS %Recovery, LCSD %Recovery, the LCS/LCSD duplicate precision, and the sample duplicate precision. The

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workbook also includes a worksheet that includes a summary table on the basis of the MRLs (specified by the client) and MDLs.

5.1.7 Hydrogen and Methane Completeness Logbook

A Hydrogen and Methane Completeness Logbook will be maintained to assure that the completeness of analysis of samples submitted is greater than 95%. Non-reporting of a sample may occasionally occur due to lost samples, handling errors, or rejection of analytical data due to incomplete or unresolved QA/QC issues. Non-reporting of sample data should be minimized whenever possible.

6.0 DATA REVIEW

Logbooks, sample custody, and analysis records will be reviewed routinely by the QA officer and the laboratory supervisor. At a minimum, they will be checked for completeness and correctness. A "Data Package Hydrogen & Methane Analysis Inventory Sheet" shall be filled out to assure completeness of the data package. The format of the form is flexible, but must be acceptable to the client.

Data packages shall be reviewed for completeness and correctness prior to delivery to the client. Data package review shall be conducted by trained personnel that did not prepare the original data package. Each data package shall include a completed "Hydrogen and Methane Data Review Checklist". The format of the form is flexible, but must be acceptable to the client. A minimum of ten percent of the data packages will receive a QA review performed by a QA specialist.

7.0 ELECTRONIC DATA DELIVERABLES

For each batch of hydrogen and methane sample analysis, a summary electronic file must be prepared and submitted to the client. The contents and format of the electronic file shall be specified by the client and shall not be changed except at the client's request. EDD shall be delivered to the client by e-mail as soon as the EDD and corresponding data package are validated. Verified spreadsheet (HMDP.xls) used to process the EDD may also be sent to the client if requested.

8.0 RECORDS

Analysis Logs
Maintenance Log
Intermediate Standards Logbook
Organic Chemistry Primary Standards Logbook
Hydrogen and Methane Completeness Logbook

9.0 REVISION HISTORY

Revision #	Date	Description
0	11/09/07	Initial Document
1	02/24/10	Biennial Review
2	09/08/11	Revision to incorporate temporary deviations, minor changes

HEAL WIPP SOP List	
Test	SOP Revision
Specific Conductance	S-120.1-6
Total Recoverable Metals, Dissolved Metals	S-6010-8
Anions	S-300.0-10
Mercury	S-245.1_7470-3
Semi-Volatiles	S-8270C-10
Volatiles	S-8260-10
Alkalinity	S-SM2320-6
Total Dissolved Solids	S-SM2540C-6
Total Suspended Solids	S-SM2540D-3
Total Organic Carbon	S-TOC-5
pH	S-SM4500H+B-6
Specific Gravity	S-SM2710F-0
Anatek WIPP SOP List	
Test	SOP Revision
Trace Metal Analysis	ALI-A-315.06

Hall Environmental Analysis Laboratory

SOP Acknowledgement Sheet
SOP Name-6010
SOP Number- S-6010-7

I state that I have read the above referenced SOP. I understand all portions of the document as they apply to me and my responsibilities at Hall Environmental Analysis Laboratory. I further recognize this as the most current application of the SOP and that it supersedes any previous documents that I may have utilized.

Effective ~~9/1/11~~ SPE 9/1/11 12/1/10-9/1/11.

Name	Signature	Date	Supervisor Initials	QA Officer Initials
Ian Cameron		12/2/10		
Tiffany Shaw		11/22/10		
Shelley Vogler		12/1/10		
Rose Galbraith-Saunders		11/24/10		
Eric Suina		12/2/10		

Hall Environmental Analysis Laboratory

Common Name: EPA Method 6010B	S.O.P. Number: S-6010-7
Method: Inductively Coupled Plasma-Optical Emission Spectrometry	
Effective Date: December 1st 2010	Supersedes: S-6010-6
Laboratory Manager:	Quality Assurance/Quality Control Officer:

1.0 Scope and Application

1.1 Inductively coupled plasma-optical emission spectrometry (ICP-OES) is used to determine trace elements in solution. This S.O.P. is applicable to dissolved aqueous samples and aqueous digestates of aqueous, leachate, soil, and wipe samples.

1.2 Method 6010 is an atomic emission spectrometric technique. This method is restricted to use by, or under the supervision of analysts experienced in the use of atomic emission spectrometers for metals analysis, and skilled in the interpretation of atomic emission spectra and their use as a quantitative tool. Each analyst must demonstrate the ability to generate acceptable results with this method. Each analyst must perform a Demonstration of Capability (DOC) in accordance with the HEAL QA Plan prior to analyzing samples by this method.

1.3 The elements covered by this S.O.P. are summarized in Table 1.

Table 1: Element List

Element	Symbol	CAS Number	Element	Symbol	CAS Number
Aluminum	Al	7429-90-5	Manganese	Mn	7439-96-5
Antimony	Sb	7440-36-0	Molybdenum	Mo	7439-98-7
Arsenic	As	7440-38-2	Nickel	Ni	7440-02-0
Barium	Ba	7440-39-3	Potassium	K	7440-09-7
Beryllium	Be	7440-41-7	Selenium	Se	7782-49-2
Boron	B	7440-42-5	Silicon	Si	7440-21-3
Cadmium	Cd	7440-43-9	Silver	Ag	7440-22-4
Calcium	Ca	7440-70-2	Sodium	Na	7440-23-5
Chromium	Cr	7440-47-3	Strontium	Sr	7440-24-6
Cobalt	Co	7440-484	Thallium	Tl	7440-28-0
Copper	Cu	7440-50-8	Tin	Sn	7440-31-5
Iron	Fe	7439-89-6	Titanium	Ti	7440-32-6
Lead	Pb	7439-92-1	Uranium	U	7440-61-1
Magnesium	Mg	7439-954	Vanadium	V	7440-62-2
			Zinc	Zn	7440-66-6

1.1 Other elements may be analyzed by this method if performance at the concentrations of interest is demonstrated.

2.0 Summary of Method

2.1 This method describes multielemental determinations by ICP-OES using a simultaneous optical system and axial or radial viewing of the plasma.

2.2 The instrument, using optical spectrometry, measures characteristic emission spectra. Samples are first mixed with Yttrium (internal standard) then nebulized and transported through to the plasma torch. The radio-frequency inductively coupled plasma produces element-specific spectral emissions that are dispersed by a grating spectrometer, and simultaneously detected by wavelength specific photosensitive devices.

2.3 For each element 3 replicate intensity readings are taken at each wavelength, each reading is run through any correction algorithms if present (MSF, background correction point), and the average intensity from the three readings is used to quantitate the element concentration from the calibration curve. The quantitated result is then adjusted using an internal standard factor.

2.4 Table 2 summarizes the emission lines validated for this method.

2.5 Except for groundwater samples that have been pre-filtered and acidified, samples must be solubilized or digested using the appropriate sample preparation method.

Table 2: Validated Emission Lines

Element	Wavelength	Element	Wavelength
Aluminum	396.153	Manganese	257.610
Antimony	206.836	Molybdenum	202.031
Arsenic	188.979	Nickel	231.604
Barium	233.527	Potassium	766.490
Beryllium	313.107	Selenium	196.026
Boron	249.772	Silicon	212.412
Cadmium	228.802	Silver	328.068
Calcium	317.933	Sodium	589.592
Chromium	205.560	Strontium	421.552
Cobalt	228.616	Thallium	190.801
Copper	324.752	Tin	189.927
Iron	238.204	Titanium	334.940
Lead	220.353	Uranium	385.958
Magnesium	285.213	Vanadium	292.402
		Zinc	206.200

Confirmation Emission Lines

Element	Wavelength	Element	Wavelength
Arsenic	193.696	Cadmium	226.502
Chromium	267.716	Lead	217.000
Selenium	203.985	Boron	249.677

Internal Standard Emission Line

Element	Wavelength
Yttrium	371.029

3.0 Detection Limits

3.1 MDLs shall be empirically established annually. An MDL study must be conducted for each matrix type analyzed and are required for each preparatory/determinative method

combination utilized. MDLs are subject to change. See attached addendum for the current Detection Limits.

3.2 A summary of the current PQLs can be found in **Appendix A.**

4.0 Definitions

4.1 Calibration Blank (CalibBlank) – Blank Solution of 2% HNO3 used to "zero" the instrument.

4.2 Initial Calibration Verification (ICV) – Second source standard used to validate the calibration curve prior to sample analysis.

4.3 Initial Calibration Blank (ICB) – Analysis of Calibration blank solution after initial calibration to check the cleanliness of the instrument.

4.4 Continuing Calibration Verification (CCV) – Standard from the same source as calibration curve. Used to check the stability of the instrument.

4.5 Continuing Calibration Blank (CCB) – Analysis of Calibration Blank solution as a continuing verification of instrument cleanliness.

4.6 Interference Check Solution (ICS) – A solution of Fe, Al, Ca, and Mg. Run at the beginning of each analytical run to check correction factors.

4.7 Interference Check Solution plus Elements (ICS Elem) – A solution of Fe, Al, Ca, and Mg fortified with 0.5ug/mL of other elements of interest. Run at the beginning of each analytical run. Used to check correction factors.

4.8 Laboratory Control Spike (LCS) – Laboratory fortified blank.

4.9 Matrix Specific QC (MSpQC) –Consists of an MS, MSD, post digestion spike addition, and dilution test.

4.10 Matrix Spike (MS) – Laboratory fortified sample.

4.11 Matrix Spike Duplicate (MSD) – Laboratory fortified sample duplicate.

4.12 Method Blank (MB) – Laboratory blank.

4.13 Method Detection Limit (MDL) – The constituent concentration that when processed through the complete method, produces a signal with a 99% probability that it is different than the blank.

4.14 Practical Quantitation Limit (PQL)/Reporting Limit – The minimum limit to which an analyte can be routinely reported.

4.15 Relative Standard Deviation (RSD) – Measures the precision between the three replicate readings for each analyte.

4.16 Internal Standard (IS) – A solution of Yttrium (Y) that is added to all samples and standards. Used to correct for physical interferences.

4.17 DI Water (DI) – De-ionized water/reagent free water. Water that is passed through a purification system and is free from interferences.

5.0 Interferences

5.1 Spectral interferences are caused by background emission from continuous or recombination phenomena, stray light from the line emission of high concentration elements, spectral line overlap from another element, or unresolved molecular band spectra overlap.

5.2 Physical interferences are effects associated with the nebulization of the sample and the physical transport of the sample to the nebulizer. Changes in sample viscosity, surface tension, and high levels of dissolved solids or acids can cause significant inaccuracies.

5.3 Memory interferences occur when high analyte levels in a previous sample contribute to intensities measured in the new sample. These can occur from high levels of a particular analyte building up on the torch and spray chamber, nebulizer, injector, and from analytes not being completely rinsed from pump tubing. Extreme care must be taken to ensure that both the nebulizer and injector are clean. Pump tubing used in sample transfer to the nebulizer must also be clean and replaced frequently. Torch and spray chamber must also be cleaned regularly.

6.0 Safety

6.1 The toxicity or carcinogenicity of each reagent or standard has not been fully defined. For handling procedures and exposure information always refer to the MSDS. Exposure to reagents and standards must be kept to a minimum. Gloves, lab coats, and safety glasses must be worn when handling samples, standards, or reagents.

6.2 Refer to the Optima 4300DV operation manual for ICP safety considerations.

7.0 Apparatus and Materials

7.1 Inductively Coupled Plasma Spectrometer system

7.1.A Perkin Elmer Optima 4300 DV equipped with AS-93 Plus Autosampler, recirculator, and Winlab32 Instrument Control Software. See maintenance logbook for the most current hardware and software.

7.1.B Operating Conditions

7.1.B.1 Instrument settings are detailed in **Appendix D**.

7.1.C Always allow at least 30 minutes for the instrument to warm up after plasma ignition. The autosampler should be in the rinse position and rinse solution should be pumping to the nebulizer.

7.1.D Low-Flow Gem Cone type nebulizer with a Cyclonic spray chamber.

7.1.E 160 Liter Liquid Argon dewar with compressed argon back up tanks.

7.1.F Air Compressor

7.2 Volumetric Flasks, Class A 50ml – 200mL.

7.3 Rainin Handheld Repipettors 100uL, 1mL, 5mL, and 10mL sizes.

7.4 VWR Handheld Repipettors 100uL, and 1mL sizes.

7.5 Centrifuge Tubes – Polypropylene 15mL and 50mL (w/ skirted bottoms)

7.6 Certified clean plastic bottles – 125mL, 250mL, and 500mL sizes.

8.0 Reagents and Standards

8.1 All acids are trace metals grade or equivalent

8.2 DI Water

8.3 Nitric Acid, concentrated, trace metals grade.

8.4 5% Nitric Acid rinse solution-add 650mL of reagent grade nitric acid and volume to 13L with DI water. Every time new solution is prepared, it must be noted in the maintenance logbook and assigned a chemical ID in LIMS.

8.5 2% Nitric Acid blank solution-add 400mL of trace metals grade nitric acid and volume up to 20L with DI water.

8.5.A The utmost care must be taken to ensure that the 2% HNO3 blank solution is clean for this will be used as the calibration blank, ICB, the MB for dissolved samples, and it will also be used to make calibration standards and spikes. The solution needs to be logged into the LIMS and assigned a bottle number in the chemical inventory which will be noted on all standards and spike logs.

8.6 Stock Standard Solutions

8.6.A A Calibration/Spike standard is purchased from Ultra Scentific containing Al, Sb, As, Ba, Be, B, Cd, Ca, Cr, Co, Cu, Fe, Pb, Mg, Mn, Mo, Ni, Se, Ag, Na, Th, Ti, V, Zn at 100ug/mL, K at 1000ug/mL, and Si at 50ug/mL.

8.6.B A separate kit containing the second source standard mixes is purchased from Environmental Express containing: Mix 1 - Sb, As, Be, Cd, Ca, Cr, Co, Cu, Fe, Pb, Mg, Mn, Mo, Ni, Se, Tl, Ti, V, Zn at 100ug/mL. Mix 2 – Si at 50ug/mL, K at 1000ug/mL, and Ag, Na, Al, Ba, and B at 100ug/mL. The third mix contains: Ca, K, Mg, and Na at 1000ug/mL, and is purchased from VHG Labs.

8.6.C The primary standard for Cations includes K, Mg, Na, and Ca @ 5000ug/mL and is purchased from Environmental Express.

8.6.D A custom "PQL" Solution containing all elements reported by this SOP is purchased from VHG Labs. Ca, Mg, K, Na @125; U @ 25; Si @ 20; Se @ 12.5; B @ 10; As, Fe @ 5; Al, Sb, Tl, Sn, V @ 2.5; Mo @ 2; Cr, Co, Cu @ 1.5; Ni, Pb, Ag, Sr, Ti, Zn @ 1.25; Ba, Be, Cd, Mn @ 0.5ug/mL.

8.6.E The U, Sn, Sr and Si spike solutions are made by the analyst from the stock standards and assigned a Spike Solution ID when logged into the LIMS. The Si spike is made in DI water, while the U, Sn, Sr spike is made in 2% HNO3.

8.6.F All standards are Ultra High Purity standards, designed specifically for trace level work. Standard must not contain HCl as a preservative. All stock standards must be preserved with HNO_3 (trace levels of HF and tartaric acid are also acceptable), with the exception of the 1000ug/mL Si standard which comes in H20. All standards are stored at room temperature and should be replaced when expired, or sooner if comparison with a check standard indicates a problem.

8.7 Calibration Standards

8.7.A Calibration standards are made on a routine basis. When they show signs of degradation, become expired, or if comparison with a check standard indicates a problem. All standards and spikes must be logged into the LIMS, assigned a solution ID, and be appropriately labeled.

8.7.B All calibration standards are made in either 2% HNO_3 (for dissolved aqueous and digestates of aqueous samples) or 10% HNO_3 (for soil and leachate digestates). Cations are calibrated separately from the other elements.

8.7.C Incorporating Strontium into the calibration standards requires a 10ppm solution to be made from the 1000ppm stock standard. The intermediate 10ppm solution is to be logged into LIMS and will be referenced as a "Stock Source" on the Standard Log. A summary is found in **Appendix B.**

8.8 Second Source Standards

8.8.A All second source standards are made in either 2% HNO_3 (for dissolved aqueous and digestates of aqueous samples) or 10% HNO_3 (for soil and leachate digestates), and appropriately labeled with a solution ID assigned by LIMS.

8.8.B Incorporating Strontium into the second source standards requires a 10ppm solution to be made from the 1000ppm second source standard. The intermediate 10ppm solution is to be logged into LIMS and will be referenced as a "Stock Source" on the Standard Log. A summary of second source standards can be found in **Appendix C.**

8.9 Internal Standard

8.9.A Yttrium @ 5ug/mL is used. Prepared in 2% HNO_3, logged into LIMS, and appropriately labeled.

8.10 Interference Check Standards

8.10.A Interference Check Standard – A solution prepared in 2% HNO_3 with Al, Ca, and Mg @ 50ug/mL, and Fe @ 20ug/mL. When fresh solution is prepared, it is to be logged into LIMS and labeled.

8.10.B Interference Check Standard plus Elements – A solution prepared in 2% HNO_3 that contains the primary same concentration of Al, Fe, Ca, and Mg as in the ICS plus all other reported elements at 0.5ug/mL. When fresh solution is prepared, it is to be logged into LIMS and labeled.

9.0 Sample Collection, Preservation, Shipment, and Storage

9.1 Aqueous samples must be collected in HNO_3 preserved certified clean plastic containers. (0.5mL concentrated HNO3/125mL sample).

9.2 Dissolved samples must be prefiltered with 0.45um Teflon filters before preservation.

9.3 Soil samples must be collected in certified clean glass sample jars.

9.4 Wipe samples are collected in Environmental Express digestion tubes.

9.5 Samples may be stored at room temperature, but HEAL protocol recommends samples be stored at 0-6°C until analysis/digestion.

9.6 Holding time 180 days.

9.7 Digestates are stored at room temperature until analysis. Holding time is 180 days.

9.7.A While Ag does not have a separate holding time. Samples that contain silver, when possible, should be analyzed as soon as possible after digestion.

10.0 Quality Control

10.1 Initial Calibration

10.1.A A 3-4 point calibration is run daily.

10.1.B An ICV, ICB, ICS, and ICS Element are run for all elements.

10.2 Continuing Calibration

10.2.A A CCV is run every ten injections for all elements.

10.2.B A CCB is run after the CCV, every 10 injections.

10.3 Batch QC Requirements

10.3.A For dissolved samples a MB, LCS, and MSpQC must be analyzed for every 20 samples.

10.3.B See the appropriate digestion method for Batch QC requirements.

10.3.C The post digestion spike addition and dilution test are only required when the spike recoveries in the MS, and/or MSD are unacceptable.

11.0 Calibration

11.1 Initial Calibration

11.1.A Instrument calibration is conducted each day for each element to be analyzed and is built into the instrument control method. The calibration is automatically loaded into the sample run list from the selected method.

11.1.B Initial calibration for all elements consists of a Calibration Blank, 3-4 calibration standards, ICV, ICB, ICS, and ICS Element.

11.1.C The calibration is based upon the corrected analyte intensity versus the analyte concentration. The calibration equations for all elements are linear, forced through zero using the equation:

$$Y = MX$$

Where:

X = concentration
Y = intensity
M = slope of calibration curve

Forcing the intercept to be zero and by using three calibration points M is calculated using the following equation:

$$M = \frac{\sum_{n}^{i=1}(X_i Y_i)}{\sum_{n}^{i=1}(X_i^2)}$$

Where:

X_i = concentration at interval i
Y_i = intensity at interval i
n = number of standards (including the blank)

11.1.D The calibration levels are summarized in **Appendix B**.

11.1.E As mentioned before, the utmost care must be taken to ensure that the calibration blank is clean and that the background intensity levels are at their appropriate levels. This is accomplished by comparing the raw background of a particular element line to historical background intensity levels. If the current background intensity levels are significantly different, action must be taken to bring them closer to the established historical levels. (Changing pump tubing, cleaning the nebulizer or injector, performing instrument maintenance, etc.).

11.1.F The calibration blank is a 2% HNO_3 solution.

11.2 Continuing Calibration

11.2.A A Continuing Calibration Verification (CCV) and Continuing Calibration Blank (CCB) must be analyzed every ten analysis and at the end of each analytical batch.

12.0 Sample Analysis

12.1 Sample Preparation/Digestion

12.1.A Dissolved samples are analyzed directly.

12.1.B Aqueous samples for total recoverable metals are digested using HEAL SOP for Method 3005A.

12.1.C Aqueous leachates from Method 1311 or method 1312 are digested using HEAL SOP for Method 3010A.

12.1.D Soil samples are digested using HEAL SOP for Method 3050B.

12.1.E Wipe samples are analyzed using the HEAL SOP for Metals Analysis on Environmental Express Ghost Wipe.

12.2 Analysis

12.2.A Use secure log in to open software.

12.2.B Load the appropriate method into the instrument control software. Each method contains all of the instrument control settings and QC limits.

12.2.C Place fresh standards into 50mL centrifuge tubes and place in the appropriate location.

12.2.C.1 Be sure to use the correct calibration standards. All aqueous samples (dissolved, 3005, and wipe digestates) are analyzed using the 2% nitric standards. All soil or leachate digestates (3010 and 3050) are analyzed using the 10% nitric standards.

12.2.D Open Sample Information file and list all samples to be analyzed. Be sure to include their location on the autosampler. Save Sample Information file.

12.2.E Open a new Results table. The instrument will automatically place the curve and continuing and closing standards in their appropriate places based upon the settings in the method.

12.2.F Samples (dissolved or digestates) are placed into a clean 15mL-centrifuge tube and placed in autosampler.

12.2.G Click Analyze All. The instrument will automatically run the curve, initial calibration QC, all samples in the sample information file, and all continuing calibration and closing QC required.

13.0 Calculations

13.1 Dissolved – The result is taken directly from the instrument data sheet and multiplied by any dilution factors if applicable. And reported in mg/L.

13.2 Aqueous (or aqueous extract) – The result is taken directly form the instrument data sheet and multiplied by dilution factors if applicable. And reported in mg/L. (The initial volume is the same as the final volume). Calculation is as follows:

$$mg/L = A \times \frac{F}{I} \times D$$

Where:

A = Result from instrument data sheet
F = Final volume in mL
I = Initial volume in mL
D = Dilution factor

13.3 Soil – 1 g of sample is used 50 mL is the final volume. The result from the instrument data sheet is multiplied times 50 and by any dilution factors if applicable. And reported in mg/kg. Calculation is as follows:

$$mg/kg = A \times \frac{F}{I} \times \frac{1000g}{1kg} \times \frac{1L}{1000mL} \times D$$

Where:

A = Result from instrument data sheet
F = Final volume in mL
I = Initial volume in g
D = Dilution factor

13.4 Wipe – One wipe is used, 50mL is the final volume. The result from instrument data sheet times 50 divided by 1000. Results are reported in mg/100cm^2.

$$mg/100cm^2 = A \times \frac{F}{I} \times \frac{1L}{1000mL} \times D$$

Where:

A = Result from instrument data sheet
F = Final volume in mL
I = Initial volume (1 wipe or 100cm^2)
D = Dilution factor

13.5 Hardness (as CaCO3) by calculation —Results for Ca and Mg used in the calculation must fall within the calibration range. If analyte requires dilution, diluted final result is used. Results are reported in mg/L. Calculation is as follows:

$$mg/L = (2.497 \times A) + (4.118 \times B)$$

Where:

A = On-scale Ca result in mg/L
B = On-scale Mg result in mg/L

13.5 Sodium Absorption Ratio(SAR)—Ratio of soluble and exchangeable Na ions in a saturated soil paste, or an aqueous sample using total concentrations of Ca, Mg, and Na. Results are unit less. See **Appendix H** for procedure. Calculation is as follows:

$$SAR = \frac{A \times D \times 0.0435^*}{\sqrt{\left[\frac{(B \times D \times 0.0499^*) + (C \times D \times 0.08229^*)}{2}\right]}}$$

Where:

A = Result for Na from instrument data sheet in mg/L
B = Result for Ca from instrument data sheet in mg/L
C = Result for Mg from instrument data sheet in mg/L
D = Dilution factor
* = Constant used to convert mg/L to meq/L (specific to each cation)

14.0 Method Performance/Estimation of Uncertainty

14.1 Method Performance

14.1.A PT or "blind study" samples for various matrix types are routinely analyzed to check the accuracy of the metals analysis.

14.2 Estimation of Uncertainty

14.2.A Estimation of uncertainty is to be conducted for all metals in each matrix type on an annual basis.

14.2.B Collect the most recent 20 LCS results for each element. Calculate the standard deviation (s) and the 95% confidence interval (2 times the standard deviation). Report the uncertainty as ±2(s).

14.2.C Refer to the HEAL QA Plan for the formulas to calculate the standard deviation (s) and 95% confidence interval.

14.3 Demonstration of Capability

14.3.A Demonstration of Capability (DOC) samples are analyzed before an analyst is able to independently generate data. DOCs for metals must be generated for each preparatory method utilized. A DOC consists of four LCS spikes. The average percent recovery and relative standard deviation are then calculated and must pass within ±20% and ≤20% respectively.

14.3.B Annual Documentation of Continued Proficiency (ADOCP) is performed annually after acceptable DOC has been established. An ADOCP consists of either four consecutive LCS spikes, or an acceptable proficiency.

14.3.C A detailed description of DOCs and ADOCPs can be found in the HEAL QA Plan.

15.0 Pollution Prevention

15.1 The Perkin Elmer ICP 4300DV is a simultaneous instrument. It is able to analyze many elements without requiring much sample. Generally only 4-8mL is required. Smaller sample containers are used on the auto sampler.

15.2 Small sample size used during analysis also contributes to savings in calibration, second source, and continuing calibration standards.

15.3 Smaller sample size for the instrument also allows small digestion volumes. The volume of sample required for digestion has been lowered to 50mL for aqueous samples. Soils only require 1g for digestion. The volume of sample waste has been cut in half.

16.0 Data Assessment and Acceptance Criteria for Quality Control Measures

16.1 Wavelength Selection, Spectral Corrections, and Multicomponent Spectral Fitting (MSF)

16.1.A The analyte is visually represented by a spectrum. Each element has a response peak centered on the selected wavelength (if the analyte response is high enough). For more complicated spectra or elements with lower intensities, this response peak is not easily seen. Especially at lower analyte concentrations. The software instead "generates" a response peak from the correction algorithms used. (For this SOP MSF is the most common correction used).

16.1.B The selection of a particular wavelength line is accomplished by examining emission intensity and the background emission intensity at a region surrounding the selected wavelength. The off-line background emission intensity from other known interfering elements at high concentrations must also be examined to determine the off-line effects of the selected analyte line. From this information an appropriate correction scheme is selected. Selected analyte scanning ranges and the correction schemes are summarized in **Appendix D**.

16.1.C All elements reported by this SOP fall into two types of correction schemes. Those analytes with very high intensities use peak area with two-point background correction points, while those analytes that are susceptible to off-line interferences, have lower analyte intensities, or contain complex spectra use MSF. See **Appendix D**.

16.1.D MSF utilizes a multiple linear least-squares model. Using the pure analyte, the calibration blank, and the matrix (or off-line interference) spectra to create interference corrected mathematical model. MSF is used when scanned spectral regions contain not only the analyte peak but also other spectral peaks or baseline shifts. Elements using MSF do not need to be examined for correct analyte peak integration. MSF examines the entire spectral region surrounding the selected spectral line and mathematically "filters" the effects of matrix, blank, and pure analyte. MSF generates a mathematically corrected spectrum that appears in the software as a peak or group of peaks below the uncorrected spectra. The MSF "model" that is created improves detection limits and accuracy considerably. **Appendix E** contains a summary of the MSF analyte model used for analysis.

16.1.E Two-Point background correction is used when the intensities for the selected element are great enough, and the spectral region around selected wavelength do not contain any interferences peaks or baseline shifts that could interfere with the baseline integration of the analyte peak. Each background correction point is selected at the left and right peak minima, and a baseline is drawn between the two points. The area under the curve is used to quantitate the result.

16.1.F Alternate spectral lines are chosen if there is a direct overlap at a selected analyte wavelength from an off-line interference.

16.2 Data Assessment

16.2.A Any analyte outside of the linear calibration range must be re-analyzed at dilution. Regardless of the sample type, all dilutions are into 2% HNO_3.

16.2.B Internal standard recovery must fall between 80-120% for the sample to be considered valid. Dilute the sample if necessary.

16.2.C See **Appendix I** for a summary of the Calibration, Continuing Calibration, batch QC requirements, acceptance criteria, and corrective actions.

16.3 Acceptance Criteria for Quality Control Measures

16.3.A Initial Calibration

16.3.A.1 Three to four point calibration curve forced through zero must have an $r^2 \geq 0.9995$.

16.3.A.2 ICV recovery must fall between 90%-110%, and the RSD from the three replicate readings of each analyte must be ≤5%.

16.3.A.3 ICB analyte concentration must fall below the PQL.

16.3.A.4 ICS - All elements (not including Fe, Ca, Mg, and Al) must fall below 2 times the PQL.

16.3.A.5 ICS Element – All elements (not including Fe, Ca, Mg, and Al) must be recovered within ±20% of true value.

16.3.B Continuing Calibration

16.3.B.1 CCV recovery must fall between 90%-110%, and the RSD from the three replicate readings of each analyte must be ≤5%.

16.3.B.2 CCB is from the same source as the Calibration Blank and must not contain target analytes above the PQL.

16.3.C Batch QC

16.3.C.1 For a MB to be considered valid it must not contain analytes above the established PQL or be less than 5% of the lowest sample concentration for each analyte.

16.3.C.2 An LCS should contain the analytes at a level that falls between the low and high standards of the calibration curve. For an LCS to be considered valid the spike recovery must fall within 80%-120%.

16.3.C.3 Matrix Specific QC

16.3.C.3.1 As the primary component of the MSpQC, the MS/MSD should contain the same level of analyte as that of the LCS. For the MS/MSD to be considered valid the spike recovery must fall between 75%-125%, have passing Internal Standard recoveries, and analytes must be on-scale.

16.3.C.3.2 The post digestion spike addition as a second component of the MSpQC, requires that the sample from which the MS, and MSD were prepared from be spiked, post digestion, and be recovered within 80-120%. An assessment of the data can only be made if the concentration of the analyte in the post spike falls within

the range of 10 to 100 times the PQL. See **Appendix G** for post spike ranges of quantitation, specific to test.

16.3.C.3.3 As the final step in the MSpQC process, the dilution test says that analysis of the sample at a 1:5 dilution should generate results that agree within ±10%. Only analytes that yield concentrations >10 times the PQL will be assessed, and any analyte(s) that have concentration above the high standard will not be reported.

16.3.C.3.4 The RPD between the MS and MSD must be less than 20%, for analytes whose concentrations are greater than 10 times the PQL.

16.3.C.3.5 See **Appendix F** for MSpQC flow chart.

17.0 Corrective Actions For Out-of-Control Data

17.1 Initial Calibration

17.1.A If the r^2 is unacceptable the failing analyte(s) must be recalibrated.

17.1.B If ICV or ICB is unacceptable, reanalyze standards up to two additional times. Recalibrate and reanalyze, if needed. (Failing elements may not be reported).

17.1.C If ICS is unacceptable, reanalyze standard up to two additional times. Unacceptable elements in the ICS may only be reported in samples that are ND for the failing elements (the exceptions being B, Co and U - see section 17.1.E). Otherwise recalibrate and reanalyze.

17.1.D If ICS Element is unacceptable, reanalyze standard up to two additional times. Elements that are failing low may not be reported, recalibrate the instrument. Elements that are failing high may only be reported if samples are non-detect for those elements (the exceptions being B, Co and U – see section 17.1.E). Otherwise the instrument must be recalibrated.

17.1.E Both B and U have significant Fe interferences. Both analytes will fail for ICS/ICS Element during the initial analysis due to the amount of Fe present in those standards. As a result, U must be evaluated using spectral overlay to determine if U is actually present in the sample. At this time samples requiring U must be evaluated on a case-by-case basis using spectral overlay. The only method currently employed to minimize Fe interference is dilution of the sample.

17.1.F B and Co are reprocessed using alternative MSF models that better represent the matrices analyzed. Both B and Co will always be reported from reprocessed data.

17.2 Continuing Calibration

17.2.A If CCV or CCB is unacceptable, reanalyze standard up to two additional times. If still unacceptable, the failing analyte(s) may not be reported in the 10 samples

preceding and following the CCV or CCB. Recalibrate, if necessary, and reanalyze affected samples.

17.3 Batch QC, MSpQC

17.3.A If the MB contains elements above the reporting limit or is greater than 5% of the lowest sample concentration, reanalyze MB. If MB contains elements above the PQL but is less than 5% of the lowest sample concentration, generate corrective action and report result. If MB continues to fail re-prep and reanalyze analytical batch for failing elements.

17.3.B If the LCS is not acceptable reanalyze. If it continues to fail re-prep and reanalyze analytical batch for failing elements.

17.3.C When recoveries in the primary component of the MSpQC (MS, MSD) fall outside of the 75-125% acceptable recovery range, an assessment of the analytes of interest must be performed. If the concentration of the analyte is greater than 100 mg/L, no spike recovery is needed. If less than 100 mg/L, a series of tests must be completed and detailed in a corrective action. These include: a post digestion spike addition to the sample and, if necessary, a dilution test which includes analysis of the sample at a 1:5 dilution. See **Appendix F** for an informative MSpQC flow chart.

17.3.C.1 The post digestion spike is only considered valid if it has a recovery within 80-120%. A dilution of the sample may be necessary for analytes that generate results above the high standard after the post spike addition. The only data reported will be analytes whose concentrations fall within 10 to 100 times the PQL. See **Appendix G** for a range of quantitation specific to test.

17.3.C.2 A failure in the MS, MSD and in the post spike confirms matrix interference(s). If recovery is acceptable, and the concentration does not exceed the high standard, the analyte can be reported.

17.3.C.3 If spike recovery falls outside of the ±20% criterion, a corrective action must be generated and a dilution test must be performed. The dilution test consists of an analysis of a 1x and a 5x dilution. Where at a minimum, concentrations of the analytes are 10 times the PQL after dilution, the analysis should agree within ±10%. Any analyte(s) whose concentration(s) exceed the high standard at the 5x dilution will not be considered or reported.

17.3.C.4 If analyte passes dilution test, no physical or chemical interferences can be confirmed. If analytes do not agree within ±10%, a chemical, physical and/or matrix related interference is suspected and the end user should be notified.

17.3.C.5 Analytes that are susceptible to off-line interferences, have low intensities, or contain complex spectra, will most likely require a post spike addition. These elements include, but are not limited to: As, Se, Sb, and Tl.

17.3.D If the RPD between MS and MSD is greater than 20, reanalyze and report reanalysis. Only analytes that are >10 times the PQL require assessment. If

failure continues, generate corrective action, flag result, and report (for soil samples reanalyze/narrate as possible matrix inconsistency).

18.0 Contingencies For Handling Out of Control Data

18.1 A corrective action is generated and all out of control data is flagged with the appropriate qualifier.

18.2 The client is notified about the out-of-control data.

19.0 Waste management

19.1 Analytical results are used to characterize their respective sample contamination level(s) so that the proper disposal can be performed. These wastes will be disposed of according to their hazard as well as their type and level of contamination. Refer to the Hall Environmental Analysis Laboratory Chemical Hygiene Plan for details regarding waste disposal.

20.0 References

20.1 U.S. EPA Method 6010B, Revision 2, December 1996.

20.2 Concepts, Instrumentation and Techniques in Inductively Coupled Plasma Optical Emission Spectrometry, Third Edition. Charles B. Boss and Kenneth J. Fredeen. Perkin Elmer, 2004.

20.3 U.S. EPA Method 3005A, Revision 1, July 1992.

20.4 HEAL SOP S-3005C-4.

20.5 U.S. EPA Method 3010A, Revision 1, July 1992.

20.6 HEAL SOP S-3010C-4.

20.7 U.S. EPA Method 3050B, Revision 2, December 1996.

20.8 HEAL SOP S-3050C-4.

20.9 Hall Environmental Quality Assurance Plan – Revision 8.2.

20.10 Hall Environmental Chemical Hygiene Plan - Version 2.0.1.

20.11 Sample Preparation for Pb on Environmental Express Ghost Wipe, using Environmental Express HotBlock.

20.12 SW-846, Chapter 1, Revision 1, July 1992.

20.13 Standard Methods for the Examination of Water and Wastewater, 18th Edition, 1992.

20.14 USDA Handbook No.60, pg. 26.

Appendix A

Method 6010B - PQLs

	Dissolved Metals mg/L	Total Metals mg/L	Soil Metals mg/kg	TCLP Metals mg/L	Wipe Metals mg/100cm²
Aluminum	0.02	0.02	3		
Antimony	0.05	0.05	2.5		0.01
Arsenic	0.02	0.02	2.5	5	0.01
Barium	0.02	0.01	0.1	100	0.001
Beryllium	0.003	0.003	0.15		
Boron	0.04	0.04	2		0.01
Cadmium	0.002	0.002	0.1	1	0.001
Calcium	1	0.5	25		
Chromium	0.006	0.006	0.3	5	0.001
Cobalt	0.006	0.006	0.3		
Copper	0.006	0.006	0.3		
Iron	0.02	0.05	1		
Lead	0.005	0.005	0.25	5	0.001
Magnesium	1	0.5	25		
Manganese	0.002	0.002	0.1		
Molybdenum	0.008	0.008	0.4		
Nickel	0.01	0.01	0.5		
Potassium	1	1	50		
Selenium	0.05	0.05	2.5	1	0.005
Silicon	0.08	0.5	25		
Silver	0.005	0.005	0.25	5	0.001
Sodium	1	0.5	25		
Strontium	0.006	0.01			
Thallium	0.05	0.05	2.5		
Tin	0.02	0.01	1		
Titanium	0.005	0.005	0.25		
Uranium	0.1	0.1	5		
Vanadium	0.05	0.05	2.5		
Zinc	0.05	0.02	2.5		
Silica	0.16	1.083			

Appendix B

Calibration Summary

Calibration standard		St1			St2			CCV St3		St4	
		VHG Custom Standard 601			Ultra QCS Spike Mix						
Aluminum	0.02	2.5	0.80	0.010	100	0.10	0.05	1.0	0.50	2.0	1.0
Antimony	0.005	2.5	0.80	0.010	100	0.10	0.05	1.0	0.50	2.0	1.0
Arsenic	0.02	5	0.80	0.020	100	0.10	0.05	1.0	0.50	2.0	1.0
Barium	0.002	0.5	0.80	0.002	100	0.10	0.05	1.0	0.50	2.0	1.0
Beryllium	0.003	0.5	0.80	0.002	100	0.10	0.05	1.0	0.50	2.0	1.0
Boron	0.04	10	0.80	0.040	100	0.10	0.05	1.0	0.50	2.0	1.0
Cadmium	0.002	0.5	0.80	0.002	100	0.10	0.05	1.0	0.50	2.0	1.0
Calcium	1	125	0.80	0.500	100	0.10	0.05	1.0	0.50	2.0	1.0
Chromium	0.006	1.5	0.80	0.006	100	0.10	0.05	1.0	0.50	2.0	1.0
Cobalt	0.006	1.5	0.80	0.006	100	0.10	0.05	1.0	0.50	2.0	1.0
Copper	0.006	1.5	0.80	0.006	100	0.10	0.05	1.0	0.50	2.0	1.0
Iron	0.02	5	0.80	0.020	100	0.10	0.05	1.0	0.50	2.0	1.0
Lead	0.005	1.25	0.80	0.005	100	0.10	0.05	1.0	0.50	2.0	1.0
Magnesium	1	125	0.80	0.500	100	0.10	0.05	1.0	0.50	2.0	1.0
Manganese	0.002	0.5	0.80	0.002	100	0.10	0.05	1.0	0.50	2.0	1.0
Molybdenum	0.008	2	0.80	0.008	100	0.10	0.05	1.0	0.50	2.0	1.0
Nickel	0.01	1.25	0.80	0.005	100	0.10	0.05	1.0	0.50	2.0	1.0
Potassium	1	125	0.80	0.500	1000	0.10	0.50	1.0	5.00	2.0	10.0
Selenium	0.02	12.5	0.80	0.050	100	0.10	0.05	1.0	0.50	2.0	1.0
Silicon (VHG)		20	0.80	0.080							
Silicon (Ultra)					50	0.10	0.025	1.0	0.25	2.0	0.5
Silicon (single)					1000	0.10	0.500	0.2	1.00	1.0	5.0
Silicon conc.	0.08			0.080			0.525		1.250		5.500
Silver	0.005	1.25	0.80	0.005	100	0.10	0.05	1.0	0.50	2.0	1.0
Sodium	1	125	0.80	0.500	100	0.10	0.05	1.0	0.50	2.0	1.0
Thallium	0.01	2.5	0.80	0.010	100	0.10	0.05	1.0	0.50	2.0	1.0
Titanium	0.005	1.25	0.80	0.005	100	0.10	0.05	1.0	0.50	2.0	1.0
Vanadium	0.05	2.5			100	0.10	0.05	1.0	0.50	2.0	1.0
Zinc	0.005	1.25	0.80	0.005	100	0.10	0.05	1.0	0.50	2.0	1.0
		VHG Custom Standard 601			Uranium 1000 ppm Stock						
Uranium	0.1	25	0.80	0.100	1000	0.05	0.25	0.1	0.50	1.0	5.0
		VHG Custom Standard 601			Tin 1000 ppm						
Tin	0.01	2.5	0.80	0.010	1000	0.05	0.25	0.1	0.50	0.2	1.0
		VHG Custom Standard 601			Intermediate 10ppm Standard						
Strontium	0.006	1.25	0.8	0.005	10	0.50	0.025	2.0	0.10		
Strontium (VHG)					Strontium 1000 ppm Stock						
					1000					0.05	0.25

Appendix B

Calibration Summary

			St6 Cat5		CCVCAT St7 Cat50		St8 Cat200	
Cation Spike Mix @ 5000ug/mL Environmental Express								
Calcium	1	5000	0.2	5.00	2.0	50.00	4.0	100.00
Magnesium	1	5000	0.2	5.00	2.0	50.00	4.0	100.00
Potassium	1	5000	0.2	5.00	2.0	50.00	4.0	100.00
Sodium	1	5000	0.2	5.00	2.0	50.00	4.0	100.00

Appendix C

SECOND SOURCE STANDARDS

ICV

[redacted]

QC Standard #1 @ 100ug/mL Environmental Express

Antimony	100	1.0	0.50
Arsenic	100	1.0	0.50
Beryllium	100	1.0	0.50
Cadmium	100	1.0	0.50
Calcium	100	1.0	0.50
Chromium	100	1.0	0.50
Cobalt	100	1.0	0.50
Copper	100	1.0	0.50
Iron	100	1.0	0.50
Lead	100	1.0	0.50
Magnesium	100	1.0	0.50
Manganese	100	1.0	0.50
Molybdenum	100	1.0	0.50
Nickel	100	1.0	0.50
Selenium	100	1.0	0.50
Thallium	100	1.0	0.50
Titanium	100	1.0	0.50
Vanadium	100	1.0	0.50
Zinc	100	1.0	0.50

QC Standard #2 @ 1000/100/50ug/mL Environmental Express

	Silicon	50	1.0	0.25
	Silicon	1000	0.5	2.50
*	Silicon Conc.			2.75
	Silver	100	1.0	0.50
	Sodium	100	1.0	0.50
	Aluminum	100	1.0	0.50
	Barium	100	1.0	0.50
	Boron	100	1.0	0.50
	Potassium	1000	1.0	5.00

Uranium 1000ppm Stock

Uranium	1000	0.1	0.50

Tin 1000ppm

Tin	1000	0.1	0.50

Intermediate Strontium 10ppm

Strontium	10	2.0	0.10

ICVCAT

Cation Spike Mix @ 5000ug/mL VHG Custom Standard 801

Calcium	5000	2.0	50.00
Magnesium	5000	2.0	50.00
Potassium	5000	2.0	50.00
Sodium	5000	2.0	50.00

Appendix D

INSTRUMENT SETTINGS AND METHOD PARAMETERS

Spectrometer Parameters
Read Delay: 15 sec
Replicates: 3
Read Time: Auto
min: 1 sec/max: 5 sec

Wash Parameters
Wash Frequency: Between Samps.
Wash Flow Rate: 2.00 mL/min
Wash Time: 60 sec

Peristaltic Pump Parameters
Sample Flow Rate: 2.00 mL/min
Sample Flush Time: 15 sec

Plasma Parameters
Source Equilibration Delay: 30 sec
Plasma Flow: 15 L/min
Aux Gas Flow: 0.2 L/min
Nebulizer Gas Flow: 0.55 L/min
RF Power: 1500 watts
View Distance: 15

Plasma View, Spectral Windows, and Correction Type

[redacted]	[redacted]	[redacted]	[redacted]	[redacted]	[redacted]
Aluminum	396.153	Axial	396.055	396.276	MSF
Antimony	206.836	Axial	206.794	206.889	MSF
Arsenic	188.979	Axial	188.939	189.025	MSF
Arsenic*	193.696	Axial	193.655	193.743	MSF
Barium	233.527	Axial	233.423	233.588	Area 2pt BGC
Beryllium	313.107	Axial	312.979	313.199	MSF
Boron	249.772	Axial	249.588	249.845	MSF
Boron*	249.667	Axial	249.588	249.845	Area 2pt BGC
Cadmium	226.502	Axial	226.453	226.612	MSF
Cadmium*	228.802	Axial	228.753	228.858	MSF
Calcium	317.933	Radial	317.885	318.011	Area 2pt BGC
Chromium	205.560	Axial	205.509	205.602	Area 2pt BGC
Chromium*	267.716	Axial	267.632	267.786	MSF
Cobalt	228.616	Axial	228.567	228.672	MSF
Copper	324.752	Axial	324.684	324.834	MSF
Iron	238.204	Radial	238.153	238.262	Area 2pt BGC
Lead*	217.000	Axial	216.953	217.052	MSF
Lead	220.353	Axial	220.255	220.451	MSF
Magnesium	285.213	Radial	285.152	285.283	Area 2pt BGC
Manganese	257.610	Axial	257.554	257.798	Area 2pt BGC
Molybdenum	202.031	Axial	201.987	202.079	MSF
Nickel	231.604	Axial	231.554	231.661	MSF
Potassium	766.490	Radial	766.309	766.718	Area 2pt BGC
Selenium	196.026	Axial	195.984	196.074	MSF
Selenium*	203.985	Axial	203.941	204.035	MSF
Silicon	212.412	Axial	212.367	212.464	Area 2pt BGC
Silver	328.068	Axial	327.998	328.148	MSF
Sodium	589.592	Radial	589.425	589.800	Area 2pt BGC
Strontium	421.552	Axial	421.351	421.782	Area 2pt BGC
Thallium	190.801	Axial	190.743	190.865	MSF
Tin	189.927	Axial	189.881	190.015	MSF
Titanium	334.940	Axial	334.840	335.158	MSF
Uranium	385.958	Axial	385.855	386.086	MSF
Vanadium	292.402	Axial	292.340	292.475	MSF
Zinc	206.200	Axial	206.101	206.246	MSF

Yttrium	371.028	Axial	370.949	371.120	Area 2pt BGC
Yttrium	371.029	Radial	370.915	371.154	Area 2pt BGC

* These wavelengths are used for analyte confirmation only.

Appendix E

MSF MODEL SUMMARY

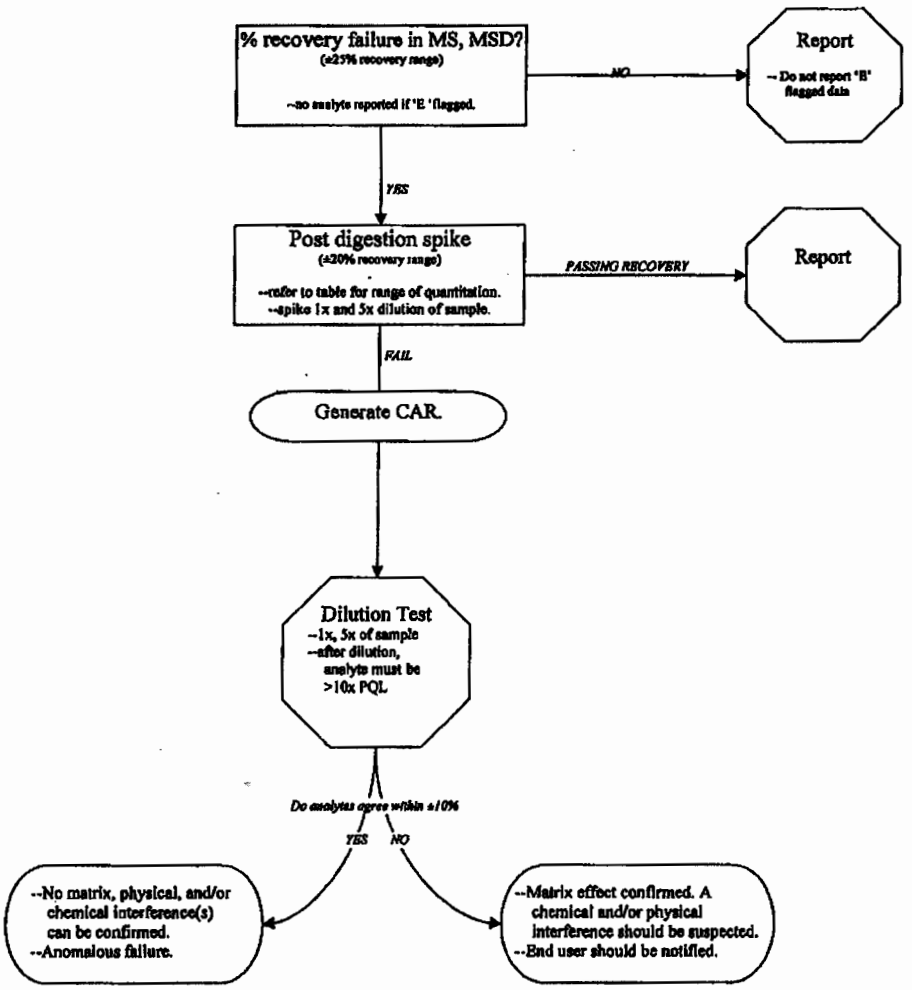
Aluminum	396.153	MSF	blank	0.5	Ca @ 500
Antimony	206.836	MSF	blank	1.0	Al @ 100, Ca @ 500, Cr @ 10, Fe @ 200, Ti @ 10
Arsenic	188.979	MSF	blank	1	Ca @ 500, Cr @ 10, Ti @ 10
Arsenic*	193.696	MSF	blank	1	Ca @ 500, Fe @ 200, V @ 10
Barium	233.527	Area 2pt BGC	-	-	
Beryllium	313.107	MSF	blank	0.5	Ti @ 10
Boron	249.772	MSF	blank	0.5	Ca @ 500, Fe @ 200, Mg @ 200
Boron*	249.677	Area 2pt BGC	-		
Cadmium*	226.502	MSF	blank	1	Fe @ 200
Cadmium	228.802	MSF	blank	1	As @ 1, Fe @ 200
Calcium	317.933	Area 2pt BGC	-	-	
Chromium	205.560	Area 2pt BGC	-	-	
Chromium*	267.716	MSF	blank	1	Fe @ 200, Mg @ 200
Cobalt	228.616	MSF	blank	1	Ba @ 200, Ti @ 10
Copper	324.752	MSF	blank	0.5	
Iron	238.204	Area 2pt BGC	-	-	
Lead*	217.000	MSF	blank	1	Al @ 100, Ca @ 500, Fe @ 200
Lead	220.353	MSF	blank	1	Ca @ 500, Fe @ 200
Magnesium	285.213	Area 2pt BGC	-	-	
Manganese	257.610	Area 2pt BGC	-	-	
Molybdenum	202.031	MSF	blank	1	Al @ 100, Ca @ 500, Fe @ 200
Nickel	231.604	MSF	blank	1	Fe @ 200
Potassium	766.490	Area 2pt BGC	-	-	
Selenium	196.026	MSF	blank	1	Ca @ 500, Fe @ 200
Selenium*	203.985	MSF	blank	1	Al @ 100, Ca @ 500, Cr @ 10, Fe @ 200, Mn @ 10
Silicon	212.412	Area 2pt BGC	-	-	
Silver	328.068	MSF	blank	0.5	Fe @ 200, Ti @ 10, V @ 10, Y @ 5
Sodium	589.592	Area 2pt BGC	-	-	
Strontium	421.552	Area 2pt BGC	-	-	
Thallium	190.801	MSF	blank	1	Ca @ 200, Fe @ 200, V @ 10
Tin	189.927	MSF	blank	1	Ca @ 500, Mg @ 200, Ti @ 10
Titanium	334.940	MSF	blank	0.5	Ca @ 500
Uranium	385.958	MSF	blank	1	Ca @ 500, Fe @ 200
Vanadium	292.402	MSF	blank	0.5	Fe @ 200
Zinc	206.200	MSF	blank	1	Cr @ 10

Yttrium	371.029	Area 2pt BGC	-	-	
Yttrium	371.029	Area 2pt BGC	-	-	

* These wavelengths are used for confirmation only.

Appendix F

Matrix Specific QC Flowchart



Appendix G

RANGE OF QUANTITATION FOR POST SPIKE

	Dissolved Metals			Total Metals			Soil Metals		
	PQL mg/L			PQL mg/L			PQL mg/kg		
Aluminum	0.02	0.2	2	0.02	0.2	2	3	30	300
Antimony	0.05	0.5	5	0.05	0.5	5	2.5	25	250
Arsenic	0.02	0.2	2	0.02	0.2	2	2.5	25	250
Barium	0.02	0.2	2	0.01	0.1	1	0.1	1	10
Beryllium	0.003	0.03	0.3	0.003	0.03	0.3	0.15	1.5	15
Boron	0.04	0.4	4	0.04	0.4	4	2	20	200
Cadmium	0.002	0.02	0.2	0.002	0.02	0.2	0.1	1	10
Calcium	1	10	100	0.5	5	50	25	250	2500
Chromium	0.006	0.06	0.6	0.006	0.06	0.6	0.3	3	30
Cobalt	0.006	0.06	0.6	0.006	0.06	0.6	0.3	3	30
Copper	0.006	0.06	0.6	0.006	0.06	0.6	0.3	3	30
Iron	0.02	0.2	2	0.05	0.5	5	1	10	100
Lead	0.005	0.05	0.5	0.005	0.05	0.5	0.25	2.5	25
Magnesium	1	10	100	0.5	5	50	25	250	2500
Manganese	0.002	0.02	0.2	0.002	0.02	0.2	0.1	1	10
Molybdenum	0.006	0.06	0.6	0.006	0.06	0.6	0.4	4	40
Nickel	0.01	0.1	1	0.01	0.1	1	0.5	5	50
Potassium	1	10	100	1	10	100	50	500	5000
Selenium	0.05	0.5	5	0.05	0.5	5	2.5	25	250
Silicon	0.08	0.8	8	0.5	5	50	25	250	2500
Silver	0.005	0.05	0.5	0.005	0.05	0.5	0.25	2.5	25
Sodium	1	10	100	0.5	5	50	25	250	2500
Strontium	0.006	0.06	0.6	0.01	0.1	1			
Thallium	0.05	0.5	5	0.05	0.5	5	2.5	25	250
Tin	0.02	0.2	2	0.01	0.1	1	1	10	100
Titanium	0.005	0.05	0.5	0.005	0.05	0.5	0.25	2.5	25
Uranium	0.1	1	10	0.1	1	10	5	50	500
Vanadium	0.05	0.5	5	0.05	0.5	5	2.5	25	250
Zinc	0.05	0.5	5	0.02	0.2	2	2.5	25	250
Silica	0.16	1.6	16	1.083	10.83	108.3			

Appendix H

SAR Procedure

Soil

1. Prep soil by making a saturated paste using DI water and allowing to sit at room temperature for 24 ± 2 hours. (See section 11.3.1 of Resistivity SOP for guidance on the preparation of a saturated paste.) Enough paste must be made so that there is enough liquid after centrifugation to perform the analysis.

2. Move paste into centrifuge tube(s) and centrifuge at a minimum of 1500rpm for 15 min.

3. Pipette off supernatant and analyze at dilution for Ca, Mg, and Na on ICP-OES.

4. On-scale results and any dilutions are entered into spreadsheet located at:
   H:\MS Office\Inorganics\USDA Methods\SAR Calculation.xls

Aqueous

1. No prep required, samples are analyzed directly.

2. On-scale results and any dilutions are entered into spreadsheet mentioned above.

Appendix I

Inductively Coupled Plasma Atomic Emmision Spectrometry Metals Analysis Using SW-846 6010B
Calibration Specification And Corrective Action Procedures

Analytical Method	Parameter	QC Element	Frequency	Acceptance Criteria	Corrective Action
SW-846 6010B	Metals	Intial Calibration (3-4 points + blank minimum)	Daily prior to analysis	Correlation coefficient $r \geq 0.9995$	1. Recalibrate
		Intial Calibration Verification (ICV)	After calibration, prior to sample analysis	Results ±10% of assigned value	1. Reanalyze ICV 2 additional times 2. Recalibrate if ICV still out
		Initial Calibration Blank (ICB)	After calibration, prior to sample analysis	< Practical quantitation limit (PQL)	1. Reanalyze 2 additional times 2. Reanalyze calibration blank and rerun ICV and ICB 3. Clean system
		Interference Check Standard + Elements (ICS Elem)	At the beginning of each run.	Results ± 25% of assigned value	1. Reanalyze 2 additional times 2. Elements that are failing low may not be reported for entire run. Recalibrate and reanalyze. 3. Elements that are failing high may be reported if samples are ND for those elements. Otherwise recalibrate and reanalyze.
		Interference Check Standard (ICS)	At the beginning of each run.	< 2 times the PQL	1. Reanalyze 2 additional times 2. Elements that are failing high may be reported if samples are ND for those elements. Otherwise recalibrate and reanalyze.
		Method Blank (MB)	1 per preperation/analysis batch (≤ 20 samples)	< PQL or must not contain analytes >5% of the lowest reported sample concentration.	1. Reanalyze 2. If MB contains elements above the PQL but less than 5% of lowest sample concentration fill out corrective action and report results. 3. Reprep and reanalyze analytical batch as appropriate
		Laboratory Control Sample (LCS)	1 per preperation/analysis batch (≤ 20 samples)	% Recovery ±20% of assigned value	1. Reanalyze LCS 2. Reprep/reanalyze LCS and affected samples
		Matrix Spike (MS)	1 per preperation/analysis batch (≤ 20 samples)	% Recovery ±25% of assigned value	1. Dilute sample and MS and reanalyze 2. Asses data. If analyte >100 mg/L no recovery needed, if <100 mg/L complete tests and CAR. 3. Refer to section 17.3.D in the SOP for details on MSpQC criteria.
		Matrix Spike Duplicate (MSD)	1 per preperation/analysis batch (≤ 20 samples)	% Recovery ±25% of assigned value Relative percent difference (RPD) ≤20%	1. Dilute sample and MS and reanalyze 2. Asses data. If analyte >100 mg/L no recovery needed, if <100 mg/L complete tests and CAR. 3. Refer to section 17.3.D in the SOP for details on MSpQC criteria. 4. If RPD >20 reanalyze and report reanalysis. Fill out corrective action and report.

Continued Next Page

Appendix I

Analytical Method	Parameter	QC Element	Frequency	Acceptance Criteria	Corrective Action
SW-846 6010B	Metals	Continuing Calibration Verification (CCV)	Every 10 samples and the end of the analytical sequence	Results ±10% of assigned value	1. Reanalyze 2 additional times 2. Reanalyze affected samples preceding and following the failed CCV.
		Continuing Calibration Blank (CCB)	Every 10 samples after each CCV and at the end of the run	< PQL	1. Reanalyze 2 additional times 2. Reanalyze affected samples preceding and following the failed CCB.

Hall Environmental Analysis Laboratory, Inc.

Corrective Action Report

Date Initiated: 20-Jun-11 **Corrective Action Report ID:** 3659

Initiated By: Ian Cameron **Department:** ME

Corrective Action Description

CAR Summary: CAR will be used in place of an active SOP for SAR-ESP prep, analysis, and calculation.

Description of Nonconformance:

Preparation
-Prepare saturated paste, per Resistivity SOP.
-Prepare MB (DI water), LCS, and LCSD (0.5mL cation Spike to 50mL FV DI)
-Allow sample to sit overnight.
-Transfer to 2-4 50mL centrifuge tubes
-Centrifuge for 15min on #6 speed setting.
-Remove supernatant with polypropylene transfer pipet and place in 15mL tube.

Analysis
-Analyze cations by DISS/6010

Calculation
-Calculate SAR and ESP using the ESP-SAR spreadsheet on server.

Prep/Analysis/Calculation is from:
Diagnosis and Improvement of Saline and Alkaline Soils, Agriculture Handbook No. 60, USDA, Issued February 1954.

Description of Corrective Action: Training to procedure performed by Ian Cameron 6/24/11
Trained: Mario Roybal 6/24/11, Rose Galbraith-Saunders 6/29/11, Eric Suing 6/24/11

Performed By: Ian Cameron **Completion Date:** 20-Jun-11

Client Notification

Client Notification Required: No **Notified By:**

Comment:

Quality Assurance Review

Nonconformance Type: Deficiency

Approval and Closure

Technical Director / Deputy Tech. Dir.: _____ Close Date: 6/29/11

QA Officer Approval: _____ QA Date:

Hall Environmental Analysis Laboratory, Inc.

Corrective Action Report

Date Initiated: 20-Jun-11 **Corrective Action Report ID:** 3659

Initiated By: Ian Cameron **Department:** ME

Further Action required by QA:

Approval and Closure

Technical Director / Deputy Tech. Dir.: _____ Close Date:

QA Officer Approval: _____ QA Date:

Last Updated BY Ian **Updated:** 24-Jun-2011 5:33 PM **Reported:** 24-Jun-2011 5:33 PM

2-1

Hall Environmental Analysis Laboratory

Common Name: EPA Method 6010B	S.O.P. Number: S-6010-8
Method: Inductively Coupled Plasma-Optical Emission Spectrometry	
Effective Date: 31 August 2011	Supersedes: S-6010-7
Laboratory Manager:	Quality Assurance/Quality Control Officer:

1.0 Scope and Application

1.1 Inductively coupled plasma-optical emission spectrometry (ICP-OES) is used to determine trace elements in solution. This S.O.P. is applicable to dissolved aqueous samples and aqueous digestates of aqueous, leachate, soil, and Ghost Wipe samples.

1.2 Method 6010 is an optical emission spectrometric technique. This method is restricted to use by intensively trained and closely supervised analysts experienced in the use of optical emission spectrometers for metals analysis, and skilled in the interpretation of optical emission spectra and their use as a quantitative tool. A detailed description of instrument operation is beyond the scope of this SOP

1.3 The elements covered by this S.O.P. are summarized in Table 1.

Table 1: Element List

Element	Symbol	CAS Number	Element	Symbol	CAS Number
Aluminum	Al	7429-90-5	Manganese	Mn	7439-96-5
Antimony	Sb	7440-36-0	Molybdenum	Mo	7439-98-7
Arsenic	As	7440-38-2	Nickel	Ni	7440-02-0
Barium	Ba	7440-39-3	Potassium	K	7440-09-7
Beryllium	Be	7440-41-7	Selenium	Se	7782-49-2
Boron	B	7440-42-5	Silicon	Si	7440-21-3
Cadmium	Cd	7440-43-9	Silver	Ag	7440-22-4
Calcium	Ca	7440-70-2	Sodium	Na	7440-23-5
Chromium	Cr	7440-47-3	Strontium	Sr	7440-24-6
Cobalt	Co	7440-484	Thallium	Tl	7440-28-0
Copper	Cu	7440-50-8	Tin	Sn	7440-31-5
Iron	Fe	7439-89-6	Titanium	Ti	7440-32-6
Lead	Pb	7439-92-1	Uranium	U	7440-61-1
Magnesium	Mg	7439-954	Vanadium	V	7440-62-2
			Zinc	Zn	7440-66-6

1.1 Other elements may be analyzed by this method if performance at the concentrations of interest is demonstrated.

2.0 Summary of Method

2.1 This method describes multielemental determinations by ICP-OES using a simultaneous optical system and axial or radial viewing of the plasma.

2.2 The instrument, using optical spectrometry, measures characteristic emission spectra. Samples are first mixed with Yttrium (internal standard) then nebulized and transported through to the plasma torch. The radio-frequency inductively coupled plasma produces element-specific spectral emissions that are dispersed by a grating spectrometer, and simultaneously detected by wavelength specific photosensitive devices.

2.3 For each element 3 replicate intensity readings are taken at each wavelength, each reading is run through any correction algorithms if present (MSF, background correction point), and the average intensity from the three readings is used to quantitate the element concentration from the calibration curve. The quantitated result is then adjusted using an internal standard factor. Quantitated results are never blank corrected.

2.4 Table 2 summarizes the emission lines validated for this method.

2.5 Except for groundwater samples that have been pre-filtered and acidified, samples must be solubilized or digested using the appropriate sample preparation method.

Table 2: Validated Emission Lines

Element	Wavelength	Element	Wavelength
Aluminum	396.153	Manganese	257.610
Antimony	206.836	Molybdenum	202.031
Arsenic	188.979	Nickel	231.604
Barium	233.527	Potassium	766.490
Beryllium	313.107	Selenium	196.026
Boron	249.772	Silicon	212.412
Cadmium	228.802	Silver	328.068
Calcium	317.933	Sodium	589.592
Chromium	205.560	Strontium	421.552
Cobalt	228.616	Thallium	190.801
Copper	324.752	Tin	189.927
Iron	238.204	Titanium	334.940
Lead	220.353	Uranium	424.167
Magnesium	285.213	Vanadium	292.402
		Zinc	206.200

Confirmation Emission Lines

Element	Wavelength	Element	Wavelength
Arsenic	193.696	Cadmium	226.502
Chromium	267.716	Lead	217.000
Selenium	203.985	Boron	249.677
		Uranium	385.958

Internal Standard Emission Line

Element	Wavelength
Yttrium	371.029

3.0 Detection Limits

3.1 MDLs shall be empirically established annually. An MDL study must be conducted for each matrix type analyzed and are required for each preparatory/determinative method combination utilized. MDLs are subject to change.

3.2 See the LIMS for the current Detection Limits. Refer to the HEAL QAP for MDL procedure and calculations.

3.3 A summary of the current PQLs can be found in **Appendix A**.

4.0 Definitions

4.1 Calibration Blank (CalibBlank) – Blank Solution of 2% HNO3 used to "zero" the instrument.

4.2 Initial Calibration Verification (ICV) – Second source standard used to validate the calibration curve prior to sample analysis.

4.3 Initial Calibration Blank (ICB) – Analysis of Calibration blank solution after initial calibration to check the cleanliness of the instrument.

4.4 Continuing Calibration Verification (CCV) – Standard from the same source as calibration curve. Used to check the stability of the instrument.

4.5 Continuing Calibration Blank (CCB) – Analysis of Calibration Blank solution as a continuing verification of instrument cleanliness.

4.6 Interference Check Solution (ICS) – A solution of Fe, Al, Ca, and Mg. Run at the beginning of each analytical run to check correction factors.

4.7 Interference Check Solution plus Elements (ICS Elem) – A solution of Fe, Al, Ca, and Mg fortified with 0.5ug/mL of other elements of interest. Run at the beginning of each analytical run. Used to check correction factors.

4.8 Laboratory Control Spike (LCS) – Laboratory fortified blank.

4.9 Laboratory Control Spike Duplicate (LCSD) – Laboratory fortified blank duplicate.

4.10 Matrix Specific QC (MSpQC) –Consists of an MS, MSD, post digestion spike addition, and dilution test.

4.11 Matrix Spike (MS) – Laboratory fortified sample.

4.12 Matrix Spike Duplicate (MSD) – Laboratory fortified sample duplicate.

4.13 Method Blank (MB) – Laboratory blank.

4.14 Method Detection Limit (MDL) – The constituent concentration that when processed through the complete method, produces a signal with a 99% probability that it is different than the blank.

4.15 Practical Quantitation Limit (PQL)/Reporting Limit – The minimum limit to which an analyte can be routinely reported.

4.16 Relative Standard Deviation (RSD) – Measures the precision between the three replicate readings for each analyte.

4.17 Internal Standard (IS) – A solution of Yttrium (Y) that is added to all samples and standards. Used to correct for physical interferences.

4.18 DI Water (DI) – De-ionized water/reagent free water. Water that is passed through a purification system and is free from interferences.

5.0 Interferences

5.1 Spectral interferences are caused by background emission from continuous or recombination phenomena, stray light from the line emission of high concentration elements, spectral line overlap from another element, or unresolved molecular band spectra overlap.

5.2 Physical interferences are effects associated with the nebulization of the sample and the physical transport of the sample to the nebulizer. Changes in sample viscosity, surface tension, and high levels of dissolved solids or acids can cause significant inaccuracies.

5.3 Memory interferences occur when high analyte levels in a previous sample contribute to intensities measured in the new sample. These can occur from high levels of a particular analyte building up on the torch and spray chamber, nebulizer, injector, and from analytes not being completely rinsed from pump tubing. Extreme care must be taken to ensure that both the nebulizer and injector are clean. Pump tubing used in sample transfer to the nebulizer must also be clean and replaced frequently. Torch and spray chamber must also be cleaned regularly.

6.0 Safety

6.1 The toxicity or carcinogenicity of each reagent or standard has not been fully defined. For handling procedures and exposure information always refer to the MSDS. Exposure to reagents and standards must be kept to a minimum. Gloves, lab coats, and safety glasses must be worn when handling samples, standards, or reagents.

6.2 Refer to the Optima 4300DV operation manual for ICP safety considerations.

7.0 Apparatus and Materials

7.1 Inductively Coupled Plasma Spectrometer system

7.1.A Perkin Elmer Optima 4300 DV equipped with AS-93 Plus Autosampler, recirculator, and Winlab32 Instrument Control Software. See maintenance logbook for the most current hardware and software.

7.1.B Operating Conditions

7.1.B.1 Instrument settings are detailed in **Appendix D**.

7.1.C Always allow at least 30 minutes for the instrument to warm up after plasma ignition. The autosampler should be in the rinse position and rinse solution should be pumping to the nebulizer.

7.1.D Low-Flow Gem Cone type nebulizer with a Cyclonic spray chamber.

7.1.E 160 Liter Liquid Argon Dewar with compressed argon back-up tanks.

7.1.F Air Compressor

7.2 Volumetric Flasks, Class A 50ml – 1000mL.

7.3 Rainin Handheld Repipetters 100µL, 1mL, 5mL, and 10mL sizes.

7.4 VWR Handheld Repipetters 100µL, and 1mL sizes.

7.5 Centrifuge Tubes – Polypropylene 15mL and 50mL (w/ skirted bottoms)

7.6 Certified clean plastic bottles – 125mL, 250mL, 500mL, and 1000mL sizes.

7.7 Analytical Balance.

7.8 Hamilton Glass GASTIGHT Syringes - 1000µL, 250µL, and 100µL sizes.

8.0 Reagents and Standards/Spikes

8.1 All acids are trace metals grade or equivalent

8.2 DI Water

8.3 Nitric Acid, concentrated, trace metals grade.

8.4 5% Nitric Acid rinse solution-add 650mL of reagent grade HNO_3 and volume to 13L with DI water. Every time new solution is prepared, it must be noted in the maintenance logbook and assigned a chemical ID and expiration date in LIMS.

8.5 2% Nitric Acid blank solution-add 400mL of trace metals grade HNO_3 and volume up to 20L with DI water.

8.5.A The utmost care must be taken to ensure that the 2% HNO_3 blank solution is clean for this will be used as the calibration blank, ICB, the MB for dissolved samples, and it will also be used to make calibration standards and spikes. The solution needs to be logged into the LIMS and assigned a bottle number in the chemical inventory which will be noted on all standards and spike logs.

8.6 Stock Standard Solutions

8.6.A All standards are Ultra High Purity standards, designed specifically for trace level work. Standards cannot contain HCl as a preservative. All stock standards must be preserved with HNO_3 (trace levels of HF and tartaric acid are also acceptable), with the exception of the 1000µg/mL Si standard which comes in H_20. All standards are stored at room temperature and should be replaced when expired, or sooner if comparison with a check standard indicates a problem.

8.7 Calibration Standards/Spikes

8.7.A A custom "PQL" Solution containing all elements reported by this SOP is purchased from VHG Labs and contains: Ca, Mg, K, Na @125µg/mL; U @ 25µg/mL; Si @ 20µg/mL; Se @ 12.5µg/mL; B @ 10µg/mL; As, Fe @ 5µg/mL; Al, Sb, Tl, Sn, V @ 2.5µg/mL; Mo @ 2µg/mL; Cr, Co, Cu @ 1.5µg/mL; Ni, Pb, Ag, Sr, Ti, Zn @ 1.25µg/mL; Ba, Be, Cd, Mn @ 0.5µg/mL.

8.7.B The Calibration/Spike standards are purchased from VHG Labs and contain:

8.7.B.1 Mix 1 - Al, Sb, As, Ba, Be, B, Cd, Cr, Co, Cu, Fe, Pb, Mn, Mo, Ni, Se, Ag, Tl, Sn, Ti, U, V, Zn at 100µg/mL, Si at 500µg/mL, and Sr at 25µg/mL.

8.7.B.2 Mix 2 - K, Mg, Na, and Ca at 5000µg/mL

8.7.C Calibration standards are made on a routine basis, when they show signs of degradation, become expired, or if comparison with a check standard indicates a problem. The standards are made using either Eppendorf Pipettes or the Hamiliton Glass GASTIGHT syringes.

8.7.D All standards and spikes must be logged into the LIMS, and appropriately labeled with a standard ID, and expiration date. If Eppendorf Pipettes are used to make the calibration standards, a pipette calibration sticker must be completed for each volume dispensed and placed on the appropriate Standard/Spike Log.

8.7.E All calibration standards are made in either 2% HNO_3 (for dissolved aqueous and digestates of aqueous samples) or 10% HNO_3 (for soil and leachate digestates). A summary can be found in **Appendix B**.

8.7.E.1 Cations are calibrated separately from the other elements and are always made in 2% HNO_3.

8.8 Second Source Standards

8.8.A A separate kit containing the second source standard mixes is purchased from Environmental Express containing:

8.8.A.1 Mix 1 - Sb, As, Be, Cd, Ca, Cr, Co, Cu, Fe, Pb, Mg, Mn, Mo, Ni, Se, Tl, Ti, V, Zn at 100µg/mL. Mix 2 – Si at 50µg/mL, K at 1000µg/mL, and Ag, Na, Al, Ba, and B at 100µg/mL.

8.8.A.2 Mix 2 - Ca, K, Mg, and Na at 5000µg/mL.

8.8.B Uranium and Tin are added to the second source standard from individual standards both at 1000µg/mL purchased from Environmental Express.

8.8.C Incorporating Strontium into the second source standards requires a 10ppm solution to be made from the 1000ppm second source standard. The intermediate 10ppm solution is to be logged into LIMS and will be referenced as a "Stock Source" on the Standard Log.

8.8.D All second source standards must be logged into the LIMS, and appropriately labeled with a standard ID, and expiration date.

8.8.E Second Source standards are made using either an Eppendorf Pipette or a Hamilton Glass GASTIGHT Syring. If a pipette is used to make the standard, a pipette calibration sticker must be completed for the volume dispensed and placed on the Standard/Spike Log.

8.8.F All second source standards are made in either 2% HNO_3 (for dissolved aqueous and digestates of aqueous samples) or 10% HNO_3 (for soil and leachate digestates). A summary can be found in **Appendix C**.

8.8.F.1 The second source cation standard will always be made in 2% HNO_3

8.9 Internal Standard

8.9.A Yttrium @ 5µg/mL is used. It is prepared in 2% HNO_3, logged into LIMS, and appropriately labeled. The assigned standard ID will be recorded on the ICP Analysis Summary sheet included in every data packet.

8.10 Interference Check Standards

8.10.A The stock standard used in both the interference check standards is purchased from Environmental Express and contains: Al, Ca, Mg @5000µg/mL, and Fe @ 2000µg/mL.

8.10.B Interference Check Standard (ICS) – A solution prepared in 2% HNO_3 with Al, Ca, and Mg @ 50µg/mL, and Fe @ 20µg/mL. When fresh solution is prepared, it is to be logged into LIMS and labeled.

8.10.C Interference Check Standard plus Elements (ICS ELEM)– A solution prepared in 2% HNO_3 that contains the same concentration of Al, Fe, Ca, and Mg as in the ICS plus all other reported elements at 0.5µg/mL. When fresh solution is prepared, it is to be logged into LIMS and labeled.

9.0 Sample Collection, Preservation, Shipment, and Storage

9.1 Aqueous samples must be collected in HNO_3 preserved certified clean plastic containers. (0.5mL concentrated HNO_3/125mL sample). Samples must be preserved to a pH<2.

9.2 Dissolved samples must be prefiltered with 0.45um Teflon filters before preservation.

9.3 Soil samples must be collected in certified clean glass sample jars.

9.4 Wipe samples are collected in Environmental Express digestion tubes (results are calculated based on 1 wipe per digestion tube).

9.5 Samples may be stored at room temperature, but HEAL protocol recommends samples be stored at 0-6°C until analysis/digestion.

9.6 Holding time for HNO_3 preserved samples is 180 days.

9.7 Digestates are stored at room temperature until analysis. Holding time is 180 days.

9.7.A While Ag does not have a separate holding time, samples that contain silver, when possible, should be analyzed as soon as possible after digestion.

10.0 Quality Control

10.1 Initial Calibration

10.1.A A 3-4 point calibration is run daily.

10.1.B An ICV, ICB, ICS, and ICS Element are run for all elements.

10.2 Continuing Calibration

10.2.A A CCV is run every ten injections for all elements.

10.2.B A CCB is run after the CCV, every 10 injections.

10.3 Batch QC Requirements

10.3.A For dissolved samples a MB, LCS, and MS, MSD must be analyzed for every 20 samples.

10.3.A.1 If there is insufficient sample volume for an MSD to be prepared, a LCSD must be prepared and analyzed.

10.3.B For wipe samples, and soil SAR/ESP samples a MB, LCS, and LCSD must be analyzed for every 20 samples.

10.3.C See the appropriate digestion method for Batch QC requirements.

10.3.D The post digestion spike addition and dilution test are only required for digested samples, when the spike recoveries in the MS, and/or MSD are unacceptable.

11.0 Calibration

11.1 Initial Calibration

11.1.A Calibration is conducted each day for each element to be analyzed and is built into the instrument control method. The calibration is automatically loaded into the sample run list from the selected method and is included in each data packet.

11.1.B Initial calibration for all elements consists of a Calibration Blank, 3-4 calibration standards, ICV, ICV Cat, ICB, ICS, and ICS Element.

11.1.C The calibration is based upon the corrected analyte intensity versus the analyte concentration. The calibration equations for all elements are linear, forced through zero using the equation:

$$Y = MX$$

Where:

X = concentration
Y = intensity
M = slope of calibration curve

Forcing the intercept to be zero and by using at least three calibration points M is calculated using the following equation:

$$M = \frac{\sum_{i=1}^{n}(X_i Y_i)}{\sum_{i=1}^{n}(X_i^2)}$$

Where:

X_i = concentration at interval i
Y_i = intensity at interval i
n = number of standards (including the blank)

11.1.D The calibration levels are summarized in **Appendix B**.

11.1.E As mentioned before, the utmost care must be taken to ensure that the calibration blank is clean and that the background intensity levels are at their appropriate levels. This is accomplished by comparing the raw background of a particular element line to historical background intensity levels. If the current background intensity levels are significantly different, action must be taken to bring them closer to the established historical levels. (Changing pump tubing, cleaning the nebulizer or injector, performing instrument maintenance, etc.).

11.1.F The calibration blank is a 2% HNO_3 solution.

11.2 Continuing Calibration

11.2.A A Continuing Calibration Verification (CCV) and Continuing Calibration Blank (CCB) must be analyzed for each analyte every ten analysis and at the end of each analytical batch.

12.0 Sample Analysis

12.1 Sample Preparation/Digestion

12.1.A Dissolved samples are analyzed directly.

12.1.A.1 To each 15mL tube add 10mL of well mixed sample.

12.1.A.1.1 For QC samples: MB, and LCS/LCSD will contain 2% HNO_3.

12.1.A.2 Check the pipette calibration using DI water and the analytical balance according to the HEAL Pipette Check SOP. Record the calibration check information on the ICP Analysis Summary sheet and include in data packet.

12.1.A.3 Spike 100µL of the Custom Spike and 100µL of the Cation Mix into the LCS/LCSD and MS, MSD samples.

12.1.B Aqueous samples for total recoverable metals are digested using HEAL SOP for Method 3005A.

12.1.C Aqueous leachates from Method 1311 or method 1312 are digested using HEAL SOP for Method 3010A.

12.1.D Soil samples are digested using HEAL SOP for Method 3050B.

12.1.E Wipe samples are digested using the Environmental Express method for Sample Preparation for Lead Analysis on the Ghost Wipe.

12.1.F Directions for SAR and/or ESP sample prep can be found in **Appendix H**.

12.2 Analysis

12.2.A Use secure log in to open software.

12.2.B Load the appropriate method into the instrument control software. Each method contains all of the instrument control settings and QC limits.

12.2.C Place fresh standards into 50mL centrifuge tubes and place in the appropriate location. Record the standard ID information on the ICP Analysis Summary sheet.

12.2.C.1 Be sure to use the correct calibration standards. All aqueous samples (dissolved, 3005, wipe digestates, and SAR/ESP samples) are analyzed using the 2% nitric standards. All soil or leachate digestates (3010 and 3050) are analyzed using the 10% nitric standards.

12.2.D Open Sample Information file and list all samples to be analyzed. Be sure to include their location on the autosampler. Save Sample Information file.

12.2.E Open a new Results table. The instrument will automatically place the curve and continuing and closing standards in their appropriate places based upon the settings in the method.

12.2.F Samples (dissolved or digestates) are placed into a clean 15mL-centrifuge tube and placed on the autosampler.

12.2.G Click Analyze All. The instrument will automatically run the curve, initial calibration QC, all samples in the sample information file, and all continuing calibration and closing QC required.

13.0 Calculations

13.1 Dissolved – The result is taken directly from the instrument data sheet and multiplied by any dilution factors if applicable. And reported in mg/L.

13.2 Aqueous (or aqueous extract) – The result is taken directly form the instrument data sheet and multiplied by dilution factors if applicable. And reported in mg/L. (The initial volume is the same as the final volume). Calculation is as follows:

$$mg/L = A \times \frac{F}{I} \times D$$

Where:

A = Result from instrument data sheet
F = Final volume in mL
I = Initial volume in mL
D = Dilution factor

13.3 Soil – 1 g of sample is used 50 mL is the final volume. The result from the instrument data sheet is multiplied times 50 and by any dilution factors if applicable. And reported in mg/kg. Calculation is as follows:

$$mg/kg = A \times \frac{F}{I} \times \frac{1000g}{1kg} \times \frac{1L}{1000mL} \times D$$

Where:

A = Result from instrument data sheet
F = Final volume in mL
I = Initial volume in g
D = Dilution factor

13.4 Wipe – One wipe is used, 50mL is the final volume. The result from instrument data sheet times 50 divided by 1000. Results are reported in mg/100cm^2.

$$mg/100cm^2 = A \times \frac{F}{I} \times \frac{1L}{1000mL} \times D$$

Where:

A = Result from instrument data sheet
F = Final volume in mL
I = Initial volume (1 wipe or 100cm^2)
D = Dilution factor

13.5 Hardness (as $CaCO_3$) by calculation —Results for Ca and Mg used in the calculation must fall within the calibration range. If analyte requires dilution, diluted final result is used. Results are reported in mg/L. Calculation is as follows:

$$mg/L = (2.497 \times A) + (4.118 \times B)$$

Where:

A = On-scale Ca result in mg/L
B = On-scale Mg result in mg/L

13.6 Sodium Adsorption Ratio (SAR)—Ratio of soluble and exchangeable Na ions in a saturated soil paste, or an aqueous sample using total concentrations of Ca, Mg, and Na. Results are unit less. See **Appendix H** for procedure. Calculation is as follows:

$$SAR = \frac{A \times D \times 0.0435*}{\sqrt{\left[\frac{(B \times D \times 0.0499*) + (C \times D \times 0.08229*)}{2}\right]}}$$

Where:

A = Result for Na from instrument data sheet in mg/L
B = Result for Ca from instrument data sheet in mg/L
C = Result for Mg from instrument data sheet in mg/L
D = Dilution factor
* = Constant used to convert mg/L to meq/L (specific to each cation)

13.7 Exchangeable Sodium Percentage (ESP) – The degree of saturation of sodium with the soil exchange complex. Results are reported as a percentage. See **Appendix H** for procedure. Calculation is as follows:

$$ESP = 100 \times \frac{(-0.0126 + 0.01475S)}{1 + (-0.0126 + 0.01475S)}$$

Where:

S = Calculated SAR result (unitless)

14.0 Method Performance/Estimation of Uncertainty

14.1 Method Performance

14.1.A PT or "blind study" samples for various matrix types are analyzed every 6 months to check the accuracy of the metals analysis.

14.2 Estimation of Uncertainty

14.2.A Estimation of uncertainty is to be conducted for all metals in each matrix type on an annual basis.

14.2.B See HEAL QAP for details on the calculation of uncertainty.

14.3 Demonstration of Capability

14.3.A Initial Demonstration of Capability (IDOC) samples are analyzed before an analyst is able to independently generate data. DOCs for metals must be generated for each preparatory method utilized. Annual Documentation of Continued Proficiency (ADOCP) is performed annually after acceptable DOC has been established.

14.3.B IDOCs consist of four replicate LCS spikes carried through the preparatory process to analysis. The average percent recovery and relative standard deviation are then calculated and must pass within ±20% and ≤20% respectively. ADOCPs consist of one of the following items, an "acceptable" PT result, a blind spike, or the successful analysis of four replicate LCS samples.

14.3.C A detailed description of IDOCs and ADOCPs can be found in the HEAL QA Plan.

15.0 Pollution Prevention

15.1 The Perkin Elmer ICP 4300DV is a simultaneous instrument. It is able to analyze many elements without requiring much sample. Generally only 4-8mL is required. Smaller sample containers are used on the auto sampler.

15.2 Small sample size used during analysis also contributes to savings in calibration, second source, and continuing calibration standards.

15.3 Smaller sample size for the instrument also allows small digestion volumes. The volume of sample required for digestion has been lowered to 50mL for aqueous samples. Soils only require 1g for digestion. The volume of sample waste has been cut in half.

16.0 Data Assessment and Acceptance Criteria for Quality Control Measures

16.1 Wavelength Selection, Spectral Corrections, and Multicomponent Spectral Fitting (MSF)

16.1.A The analyte is visually represented by a spectrum. Each element has a response peak centered on the selected wavelength (if the analyte response is high enough). For more complicated spectra or elements with lower intensities, this response peak is not easily seen. Especially at lower analyte concentrations. The software instead "generates" a response peak from the correction algorithms used. (For this SOP MSF is the most common correction used).

16.1.B The selection of a particular wavelength line is accomplished by examining emission intensity and the background emission intensity at a region surrounding the selected wavelength. The off-line background emission intensity from other known interfering elements at high concentrations must also be examined to determine the off-line effects of the selected analyte line. From this information an appropriate correction scheme is selected. Selected analyte scanning ranges and the correction schemes are summarized in **Appendix D**.

16.1.C All elements reported by this SOP fall into two types of correction schemes. Those analytes with very high intensities use peak area with two-point background correction points, while those analytes that are susceptible to off-line interferences, have lower analyte intensities, or contain complex spectra use MSF. See **Appendix D**.

16.1.D MSF utilizes a multiple linear least-squares model. Using the pure analyte, the calibration blank, and the matrix (or off-line interference) spectra to create interference corrected mathematical model. MSF is used when scanned spectral regions contain not only the analyte peak but also other spectral peaks or baseline shifts. Elements using MSF do not need to be examined for correct analyte peak integration. MSF examines the entire spectral region surrounding the selected spectral line and mathematically "filters" the effects of matrix, blank, and pure analyte. MSF generates a mathematically corrected spectrum that appears in the software as a peak or group of peaks below the uncorrected spectra. The MSF "model" that is created improves detection limits and accuracy considerably. **Appendix E** contains a summary of the MSF analyte model used for analysis.

16.1.E Two-Point background correction is used when the intensities for the selected element are great enough, and the spectral region around selected wavelength do not contain any interferences peaks or baseline shifts that could interfere with the baseline integration of the analyte peak. Each background correction point is selected at the left and right peak minima, and a baseline is drawn between the two points. The area under the curve is used to quantitate the result.

16.1.F Alternate spectral lines are chosen if there is a direct overlap at a selected analyte wavelength from an off-line interference.

16.2 Data Assessment

16.2.A Any analyte outside of the linear calibration range must be re-analyzed at dilution. Regardless of the sample type, all dilutions are into 2% HNO_3.

16.2.B Internal standard recovery must fall between 80-120% for the sample to be considered valid. Dilute the sample if necessary.

16.2.C See **Appendix I** for a summary of the Calibration, Continuing Calibration, batch QC requirements, acceptance criteria, and corrective actions.

16.3 Acceptance Criteria for Quality Control Measures

16.3.A Initial Calibration

16.3.A.1 Three to four point calibration curve forced through zero must have an $r^2 \geq 0.9995$.

16.3.A.2 ICV recovery must fall between 90%-110%, and the RSD from the three replicate readings of each analyte must be ≤5%.

16.3.A.3 ICB analyte concentration must fall below the PQL, and must be greater than or equal to the negative of the PQL.

16.3.A.4 ICS - All elements (not including Fe, Ca, Mg, and Al) must fall below 2 times the PQL.

16.3.A.5 ICS Element – All elements (not including Fe, Ca, Mg, and Al) must be recovered within ±20% of their true value.

16.3.B Continuing Calibration

16.3.B.1 CCV recovery must fall between 90%-110%, and the RSD from the three replicate readings of each analyte must be ≤5%.

16.3.B.2 CCB is from the same source as the Calibration Blank and therefore must not contain target analytes above the PQL or have an analyte concentration below the negative of the PQL.

16.3.C Batch QC

16.3.C.1 For the MB to be considered valid it must not contain analytes above the established PQL or be less than 5% of the lowest sample concentration for each analyte. In addition, the analyte concentration must not be less than 2 times the negative of the PQL.

16.3.C.2 An LCS should contain the analytes at a level that falls between the low and high standards of the calibration curve. For an LCS to be considered valid the spike recovery must fall within 80%-120%.

NOTE: LCS recoveries listed on the raw data are only estimates and should only be used to decide whether the QC sample requires reanalysis.

16.3.C.3 Matrix Specific QC

16.3.C.3.1 As the primary component of the MSpQC, the MS, MSD should contain the same level of analyte as that of the LCS. For the MS, MSD to be considered valid the spike recovery must fall between 75%-125%, have passing

Internal Standard recoveries, and analytes must be on-scale.

16.3.C.3.2 The post digestion spike addition as a second component of the MSpQC, for digested samples, requires that the sample from which the MS, and MSD were prepared from be spiked, post digestion, and be recovered within 80-120%. An assessment of the data can only be made if the concentration of the analyte in the post spike falls within the range of 10 to 100 times the PQL. See **Appendix G** for post spike ranges of quantitation, specific to test.

16.3.C.3.3 As the final step in the MSpQC process, the dilution test says that analysis of the sample at a 1:5 dilution should generate results that agree within ±10%. Only analytes that yield concentrations >10 times the PQL will be assessed, and any analyte(s) that have concentration above the high standard will not be reported.

16.3.C.3.4 The RPD between the MS and MSD must be less than 20%, for analytes whose concentrations are greater than 10 times the PQL.

16.3.C.3.5 See **Appendix F** for MSpQC flow chart and **Appendix I** for a summary of Corrective Action Procedures.

17.0 Corrective Actions For Out-of-Control Data

17.1 Initial Calibration

17.1.A If the r^2 is unacceptable the failing analyte(s) must be recalibrated.

17.1.B If ICV or ICB is unacceptable, reanalyze standards up to one additional time. Recalibrate and reanalyze, if needed. (Failing elements may not be reported).

17.1.C If ICS is unacceptable, reanalyze standard up to one additional time. Unacceptable elements in the ICS may only be reported in samples that are ND for the failing elements (the exceptions being B, and Co - see section 17.1.E). Otherwise recalibrate and reanalyze.

17.1.D If ICS Element is unacceptable, reanalyze standard up to one additional time. Elements that are failing low may not be reported, recalibrate the instrument. Elements that are failing high may only be reported if samples are non-detect for those elements (the exceptions being B, and Co – see section 17.1.E). Otherwise the instrument must be recalibrated.

17.1.E Boron has a significant Fe interference. The analyte will fail in the ICS/ICS Element during the initial analysis due to the amount of Fe present in those standards.

17.1.F B and Co are reprocessed using alternative MSF models that better represent the matrices analyzed. Both B and Co will always be reported from reprocessed data.

17.2 Continuing Calibration

17.2.A If CCV or CCB is unacceptable, reanalyze standard up to one additional time. If still unacceptable, the failing analyte(s) may not be reported in the 10 samples preceding and following the CCV or CCB. Recalibrate and reanalyze affected samples.

17.3 Batch QC, MSpQC

17.3.A If the MB contains elements above the reporting limit or is greater than 5% of the lowest sample concentration, reanalyze MB. If MB contains elements above the PQL but is less than 5% of the lowest sample concentration, generate corrective action and report result. If the concentration of the element in the MB is less than 2 times the negative of the PQL, reanalyze, If MB continues to fail the lower limit re-prep and reanalyze analytical batch for failing elements.

17.3.B If the LCS is not acceptable reanalyze. If it continues to fail re-prep and reanalyze analytical batch for failing elements.

17.3.C When recoveries in the primary component of the MSpQC (MS, MSD) fall outside of the 75-125% acceptable recovery range, an assessment of the analytes of interest must be performed. If the concentration of the analyte is greater than 100 mg/L, no spike recovery is needed. If less than 100 mg/L, a series of tests must be completed and detailed in a corrective action. These include: a post digestion spike addition to the sample and, if necessary, a dilution test which includes analysis of the sample at a 1:5 dilution. See **Appendix F** for an informative MSpQC flow chart.

17.3.C.1 The post digestion spike is only considered valid if it has a recovery within 80-120%. A dilution of the sample may be necessary for analytes that generate results above the high standard after the post spike addition. The only data reported will be analytes whose concentrations fall within 10 to 100 times the PQL. See **Appendix G** for a range of quantitation specific to test.

17.3.C.2 A failure in the MS, MSD and in the post spike confirms matrix interference(s) and should be noted in a CAR. If recovery is acceptable, and the concentration does not exceed the high standard, the analyte can be reported.

17.3.C.3 If spike recovery falls outside of the ±20% criterion, a corrective action must be generated and a dilution test must be performed. The dilution test consists of an analysis of a 1x and a 5x dilution. Where at a minimum, concentrations of the analytes are 10 times the PQL after dilution, the analysis should agree within ±10%. Any analyte(s) whose concentration(s) exceed the high standard at the 5x dilution will not be considered or reported.

17.3.C.4 If analyte passes dilution test, no physical or chemical interferences can be confirmed. If analytes do not agree within ±10%, a chemical, physical and/or matrix related interference is suspected and the end user should be notified.

17.3.C.5 Analytes that are susceptible to off-line interferences, have low intensities, or contain complex spectra, will most likely require a post spike addition. These elements include, but are not limited to: As, Se, Sb, and Tl.

17.3.D If the RPD between MS and MSD is greater than 20, reanalyze and report reanalysis. Only analytes that are >10 times the PQL require assessment. If failure continues, generate corrective action, flag result, and report (for soil samples reanalyze/narrate as possible matrix inconsistency).

17.3.E See **Appendix I** for a summary of Corrective Action Procedures.

18.0 Contingencies For Handling Out of Control Data

18.1 A corrective action is generated and all out of control data is flagged with the appropriate qualifier.

18.2 The client is notified about the out-of-control data.

19.0 Waste management

19.1 Analytical results are used to characterize their respective sample contamination level(s) so that the proper disposal can be performed. These wastes will be disposed of according to their hazard as well as their type and level of contamination. Refer to the Hall Environmental Analysis Laboratory Chemical Hygiene Plan for details regarding waste disposal.

20.0 References

20.1 U.S. EPA Method 6010B, Revision 2, December 1996.

20.2 Concepts, Instrumentation and Techniques in Inductively Coupled Plasma Optical Emission Spectrometry, Third Edition. Charles B. Boss and Kenneth J. Fredeen. Perkin Elmer, 2004.

20.3 U.S. EPA Method 3005A, Revision 1, July 1992.

20.4 Current revision of HEAL 3005 SOP.

20.5 U.S. EPA Method 3010A, Revision 1, July 1992.

20.6 Current revision of HEAL 3010 SOP.

20.7 U.S. EPA Method 3050B, Revision 2, December 1996.

20.8 Current revision of HEAL 3050 SOP.

20.9 Current revision of HEAL Quality Assurance Plan (QAP).

20.10 Current revision of HEAL Chemical Hygiene Plan (CHP).

20.11 Current revision of HEAL Variable Volume Pipette Check SOP

20.12 Sample Preparation for Pb on Environmental Express Ghost Wipe, using Environmental Express Hot Block.

20.13 SW-846, Chapter 1, Revision 1, July 1992.

20.14 Standard Methods for the Examination of Water and Wastewater, 18th Edition, 1992.

20.15 USDA Handbook No.60.

Appendix A
Method 6010B - PQLs

	Dissolved Metals mg/L	Total Metals mg/L	Soil Metals mg/kg	TCLP Metals mg/L	Wipe Metals mg/100cm²
Aluminum	0.02	0.02	3	0.05	0.0025
Antimony	0.05	0.05	2.5	0.05	0.0025
Arsenic	0.02	0.02	2.5	5	0.005
Barium	0.02	0.02	0.1	100	0.001
Beryllium	0.003	0.003	0.15	0.002	0.001
Boron	0.04	0.04	2	0.04	
Cadmium	0.002	0.002	0.1	1	0.001
Calcium	1	1	25	5	
Chromium	0.006	0.006	0.3	5	0.002
Cobalt	0.006	0.006	0.3	0.008	0.002
Copper	0.006	0.006	0.3	0.008	
Iron	0.02	0.05	1	0.05	
Lead	0.005	0.005	0.25	5	0.002
Magnesium	1	1	25	0.5	
Manganese	0.002	0.002	0.1	0.05	0.001
Molybdenum	0.008	0.008	0.4	0.008	0.002
Nickel	0.01	0.01	0.5	0.005	0.0025
Potassium	1	1	50	5.5	
Selenium	0.05	0.05	2.5	1	0.02
Silicon	0.08	0.5			
Silver	0.005	0.005	0.25	5	0.002
Sodium	1	1	25		
Strontium	0.008	0.01			
Thallium	0.05	0.05	2.5	0.05	0.0025
Tin	0.02	0.01	1	0.025	
Titanium	0.005	0.005	0.25		0.01
Uranium	0.1	0.1	5		
Vanadium	0.05	0.05	2.5	0.01	0.003
Zinc	0.02	0.02	2.5	0.05	
Silica	0.16	1.083			

Appendix B
Calibration Summary

		Std1 - 200mL FV			Std2 - 200mL FV		CCV Std3 - 200mL FV		Std4 - 200mL FV	
	Custom Standard Mix 601 VHG			Custom Calibration Mix VHG						
Aluminum	2.5	0.8	0.01	100	0.1	0.05	1	0.5	2	1
Antimony	2.5	0.8	0.01	100	0.1	0.05	1	0.5	2	1
Arsenic	5	0.8	0.02	100	0.1	0.05	1	0.5	2	1
Barium	0.5	0.8	0.002	100	0.1	0.05	1	0.5	2	1
Beryllium	0.5	0.8	0.002	100	0.1	0.05	1	0.5	2	1
Boron	10	0.8	0.04	100	0.1	0.05	1	0.5	2	1
Cadmium	0.5	0.8	0.002	100	0.1	0.05	1	0.5	2	1
Calcium	125	0.8	0.5							
Chromium	1.5	0.8	0.006	100	0.1	0.05	1	0.5	2	1
Cobalt	1.5	0.8	0.006	100	0.1	0.05	1	0.5	2	1
Copper	1.5	0.8	0.006	100	0.1	0.05	1	0.5	2	1
Iron	5	0.8	0.02	100	0.1	0.05	1	0.5	2	1
Lead	1.25	0.8	0.005	100	0.1	0.05	1	0.5	2	1
Magnesium	125	0.8	0.5							
Manganese	0.5	0.8	0.002	100	0.1	0.05	1	0.5	2	1
Molybdenum	2	0.8	0.008	100	0.1	0.05	1	0.5	2	1
Nickel	1.25	0.8	0.005	100	0.1	0.05	1	0.5	2	1
Potassium	125	0.8	0.5							
Selenium	12.5	0.8	0.05	100	0.1	0.05	1	0.5	2	1
Silver	1.25	0.8	0.005	100	0.1	0.05	1	0.5	2	1
Sodium	125	0.8	0.5							
Strontium	1.25	0.8	0.005	25	0.1	0.0125	1	0.125	2	0.25
Thallium	2.5	0.8	0.01	100	0.1	0.05	1	0.5	2	1
Tin	2.5	0.8	0.01	100	0.1	0.05	1	0.5	2	1
Titanium	1.25	0.8	0.005	100	0.1	0.05	1	0.5	2	1
Uranium	25	0.8	0.1	100	0.1	0.05	1	0.5	2	1
Vanadium	2.5	0.8	0.01	100	0.1	0.05	1	0.5	2	1
Zinc	1.25	0.8	0.005	100	0.1	0.05	1	0.5	2	1
Silicon	20	0.8	0.08	500	0.1	0.25	1	2.5	2	5

Appendix B
Calibration Summary

				CCV_CAT			
		Std6 - 200mL FV		Std7 - 200mL FV		Std8 - 200mL FV	
	Custom Standard 801 VHG						
Aluminum							
Antimony							
Arsenic							
Barium							
Beryllium							
Boron							
Cadmium							
Calcium	5000	0.2	5	2	50	4	100
Chromium							
Cobalt							
Copper							
Iron							
Lead							
Magnesium	5000	0.2	5	2	50	4	100
Manganese							
Molybdenum							
Nickel							
Potassium	5000	0.2	5	2	50	4	100
Selenium							
Silver							
Sodium	5000	0.2	5	2	50	4	100
Strontium							
Thallium							
Tin							
Titanium							
Uranium							
Vanadium							
Zinc							
Silicon							

Appendix C
Second Source Standards

	ICV - 200mL FV		
	IQC100-19 Std. Env. Express		
Antimony	100	1.00	0.500
Arsenic	100	1.00	0.500
Beryllium	100	1.00	0.500
Cadmium	100	1.00	0.500
Calcium	100	1.00	0.500
Chromium	100	1.00	0.500
Cobalt	100	1.00	0.500
Copper	100	1.00	0.500
Iron	100	1.00	0.500
Lead	100	1.00	0.500
Magnesium	100	1.00	0.500
Manganese	100	1.00	0.500
Molybdenum	100	1.00	0.500
Nickel	100	1.00	0.500
Selenium	100	1.00	0.500
Thallium	100	1.00	0.500
Titanium	100	1.00	0.500
Vanadium	100	1.00	0.500
Zinc	100	1.00	0.500
	IQC100-7 Std. Env. Express		
Aluminum	100	1.00	0.500
Barium	100	1.00	0.500
Boron	100	1.00	0.500
Potassium	1000	1.00	5.000
Silver	100	1.00	0.500
Sodium	100	1.00	0.500
Silicon (EE)	50	1.00	0.250
Silicon (EE)	1000	0.50	2.500
Silicon			2.750
	U @ 1000ppm Std. Env. Express		
Uranium	1000	0.10	0.500
	Sn @ 1000ppm Std. Env. Express		
Tin	1000	0.10	0.500
	Sr @ 10ppm - Intermediate Std.		
Strontium	10	2	0.1

	ICV CAT - 200mL FV		
	ICL500-1 Std. Env. Express		
Calcium	5000	2.00	50.0
Magnesium	5000	2.00	50.0
Potassium	5000	2.00	50.0
Sodium	5000	2.00	50.0

Appendix D
Instrument Setting and Method Parameters

Spectrometer Parameters

Read Delay: 15
Replicates: 3
Read Time: Auto
min: 1 sec/max: 5 sec

Peristaltic Pump Parameters

Sample Flow Rate: 2.00 mL/min
Sample Flush Time: 15 sec

Wash Parameters

Wash Frequency: Between samples + extra time if needed.
Wash Flow Rate: 2.00 mL/min
Wash Time: 60 sec

Plasma Parameters

Source Equilibration Delay: 30 sec
Plasma Flow: 15 L/min
Aux Gas Flow: 0.2 L/min
Nebulizer Gas Flow: 0.55 L/min
RF Power: 1500 watts
View Distance: 15

Plasma View, Spectral Windows, and Correction Type

[illegible]	[illegible]	[illegible]	[illegible]	[illegible]	[illegible]
Aluminum	396.153	Axial	396.055	396.276	MSF
Antimony	206.836	Axial	206.794	206.889	MSF
Arsenic	188.979	Axial	188.939	189.025	MSF
Arsenic*	193.696	Axial	193.655	193.743	MSF
Barium	233.527	Axial	233.423	233.588	Area 2pt BGC
Beryllium	313.107	Axial	312.979	313.199	MSF
Boron	249.772	Axial	249.588	249.845	MSF
Boron*	249.667	Axial	249.588	249.845	Area 2pt BGC
Cadmium	226.502	Axial	226.453	226.612	MSF
Cadmium*	226.802	Axial	226.753	226.858	MSF
Calcium	317.933	Radial	317.865	318.011	Area 2pt BGC
Chromium	205.560	Axial	205.508	205.602	Area 2pt BGC
Chromium*	267.716	Axial	267.532	267.786	MSF
Cobalt	228.616	Axial	228.567	228.672	MSF
Copper	324.752	Axial	324.684	324.834	MSF
Iron	238.204	Radial	238.153	238.262	Area 2pt BGC
Lead*	217.000	Axial	216.953	217.052	MSF
Lead	220.353	Axial	220.255	220.451	MSF
Magnesium	285.213	Radial	285.152	285.283	Area 2pt BGC
Manganese	257.610	Axial	257.554	257.798	Area 2pt BGC
Molybdenum	202.031	Axial	201.987	202.079	MSF
Nickel	231.604	Axial	231.554	231.661	MSF
Potassium	766.490	Radial	766.309	766.718	Area 2pt BGC
Selenium	196.026	Axial	195.984	196.074	MSF
Selenium*	203.985	Axial	203.941	204.035	MSF
Silicon	212.412	Axial	212.367	212.464	Area 2pt BGC
Silver	328.068	Axial	327.998	328.148	MSF
Sodium	589.592	Radial	589.425	589.800	Area 2pt BGC
Strontium	421.552	Axial	421.351	421.782	Area 2pt BGC
Thallium	190.801	Axial	190.743	190.865	MSF
Tin	189.927	Axial	189.881	190.015	MSF
Titanium	334.940	Axial	334.840	335.158	MSF
Uranium*	385.958	Axial	385.855	386.086	MSF
Vanadium	292.402	Axial	292.340	292.475	MSF
Zinc	206.200	Axial	206.101	206.246	MSF
Uranium	424.167	Axial	423.976	424.386	Area 2pt BGC

Yttrium	371.029	Axial	370.949	371.120	Area 2pt BGC
Yttrium	371.029	Radial	370.915	371.154	Area 2pt BGC

* These wavelengths are used for analyte confirmation only.

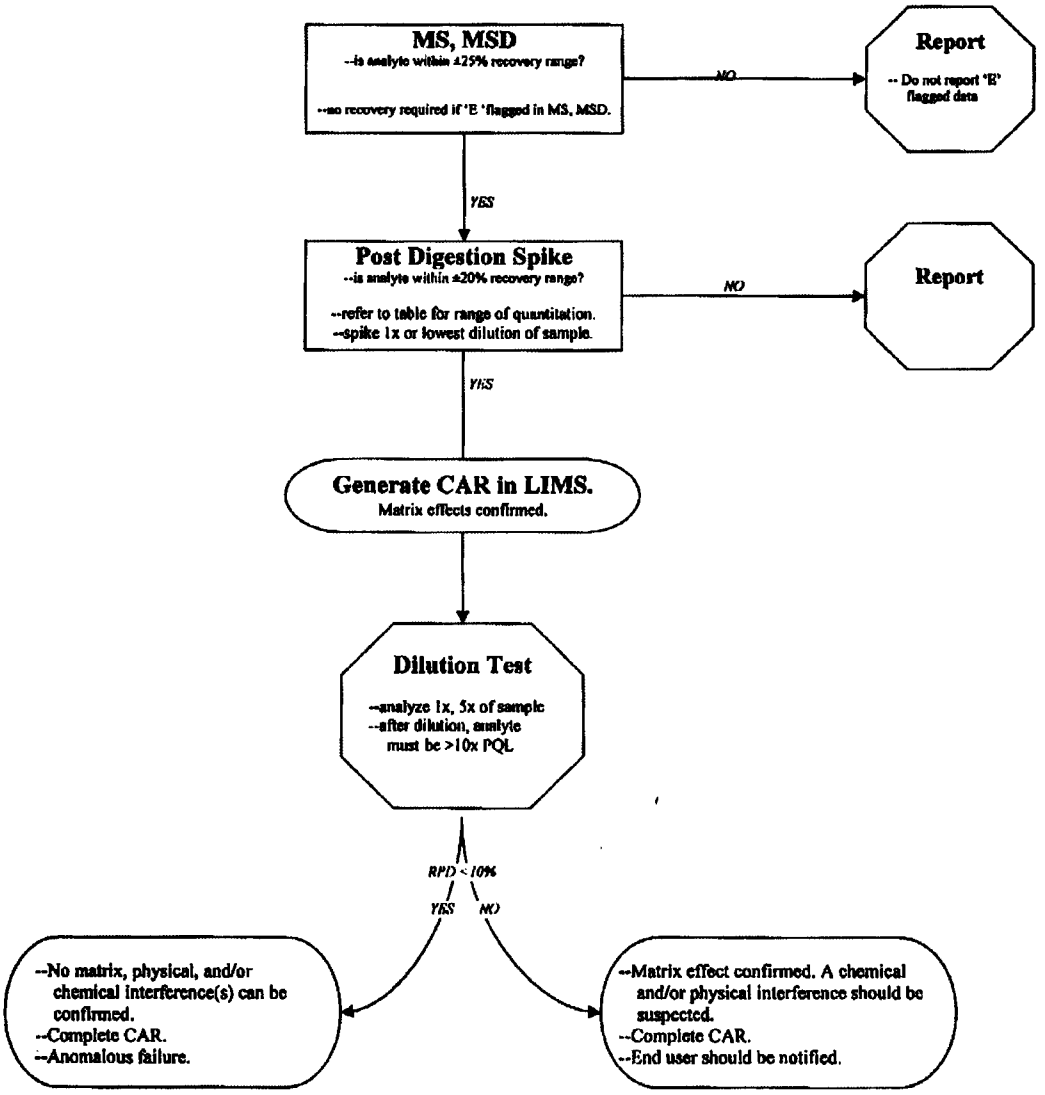
Appendix E
MSF Model Summary

[illegible]	[illegible]	[illegible]	[illegible]	[illegible]	[illegible]
Aluminum	396.153	MSF	blank	0.5	Ca @ 500
Antimony	206.836	MSF	blank	1.0	Al @ 100, Ca @ 500, Cr @ 10, Fe @ 200, Ti @ 10
Arsenic	188.979	MSF	blank	1	Ca @ 500, Cr @ 10, Ti @ 10
Arsenic*	193.696	MSF	blank	1	Ca @ 500, Fe @ 200, V @ 10
Barium	233.527	Area 2pt BGC	-	-	
Beryllium	313.107	MSF	blank	0.5	Ti @ 10
Boron	249.772	MSF	blank	0.5	Ca @ 500, Fe @ 200, Mg @ 200
Boron*	249.677	Area 2pt BGC	-		
Cadmium*	226.502	MSF	blank	1	Fe @ 200
Cadmium	228.802	MSF	blank	1	As @ 1, Fe @ 200
Calcium	317.933	Area 2pt BGC	-	-	
Chromium	205.560	Area 2pt BGC	-	-	
Chromium*	267.716	MSF	blank	1	Fe @ 200, Mg @ 200
Cobalt	228.616	MSF	blank	1	Ba @ 200, Ti @ 10
Copper	324.752	MSF	blank	0.5	
Iron	238.204	Area 2pt BGC	-	-	
Lead*	217.000	MSF	blank	1	Al @ 100, Ca @ 500, Fe @ 200
Lead	220.353	MSF	blank	1	Ca @ 500, Fe @ 200
Magnesium	285.213	Area 2pt BGC	-	-	
Manganese	257.610	Area 2pt BGC	-	-	
Molybdenum	202.031	MSF	blank	1	Al @ 100, Ca @ 500, Fe @ 200
Nickel	231.604	MSF	blank	1	Fe @ 200
Potassium	766.490	Area 2pt BGC	-	-	
Selenium	196.026	MSF	blank	1	Ca @ 500, Fe @ 200
Selenium*	203.985	MSF	blank	1	Al @ 100, Ca @ 500, Cr @ 10, Fe @ 200, Mn @ 10
Silicon	212.412	Area 2pt BGC	-	-	
Silver	328.068	MSF	blank	0.5	Fe @ 200, Ti @ 10, V @ 10, Y @ 5
Sodium	589.592	Area 2pt BGC	-	-	
Strontium	421.552	Area 2pt BGC	-	-	
Thallium	190.801	MSF	blank	1	Ca @ 200, Fe @ 200, V @ 10
Tin	189.927	MSF	blank	1	Ca @ 500, Mg @ 200, Ti @ 10
Titanium	334.940	MSF	blank	0.5	Ca @ 500
Uranium	385.958	MSF	blank	1	Ca @ 500, Fe @ 200
Vanadium	292.402	MSF	blank	0.5	Fe @ 200
Zinc	206.200	MSF	blank	1	Cr @ 10
Uranium	424.167	Area 2pt BGC	-	-	

Yttrium	371.029	Area 2pt BGC	-	-	
Yttrium	371.029	Area 2pt BGC	-	-	

* These wavelengths are used for confirmation only.

Appendix F
Matrix Specific QC Flowchart



Appendix G
Range of Quantitation for Post Spike

	Dissolved Metals			Total Metals			Soil Metals		
	PQL mg/L	Range of quantitation for post spike mg/L		PQL mg/L	Range of quantitation for post spike mg/L		PQL mg/kg	Range of quantitation for post spike mg/kg	
Aluminum	0.02	0.2	2	0.02	0.2	2	3	30	300
Antimony	0.05	0.5	5	0.05	0.5	5	2.5	25	250
Arsenic	0.02	0.2	2	0.02	0.2	2	2.5	25	250
Barium	0.02	0.2	2	0.02	0.2	2	0.1	1	10
Beryllium	0.003	0.03	0.3	0.003	0.03	0.3	0.15	1.5	15
Boron	0.04	0.4	4	0.04	0.4	4	2	20	200
Cadmium	0.002	0.02	0.2	0.002	0.02	0.2	0.1	1	10
Calcium	1	10	100	1	10	100	25	250	2500
Chromium	0.006	0.06	0.6	0.006	0.06	0.6	0.3	3	30
Cobalt	0.006	0.06	0.6	0.006	0.06	0.6	0.3	3	30
Copper	0.006	0.06	0.6	0.006	0.06	0.6	0.3	3	30
Iron	0.02	0.2	2	0.05	0.5	5	1	10	100
Lead	0.005	0.05	0.5	0.005	0.05	0.5	0.25	2.5	25
Magnesium	1	10	100	1	10	100	25	250	2500
Manganese	0.002	0.02	0.2	0.002	0.02	0.2	0.1	1	10
Molybdenum	0.008	0.08	0.8	0.008	0.08	0.8	0.4	4	40
Nickel	0.01	0.1	1	0.01	0.1	1	0.5	5	50
Potassium	1	10	100	1	10	100	50	500	5000
Selenium	0.05	0.5	5	0.05	0.5	5	2.5	25	250
Silicon	0.08	0.8	8	0.5	5	50			
Silver	0.005	0.05	0.5	0.005	0.05	0.5	0.25	2.5	25
Sodium	1	10	100	1	10	100	25	250	2500
Strontium	0.006	0.06	0.6	0.01	0.1	1			
Thallium	0.05	0.5	5	0.05	0.5	5	2.5	25	250
Tin	0.02	0.2	2	0.01	0.1	1	1	10	100
Titanium	0.005	0.05	0.5	0.005	0.05	0.5	0.25	2.5	25
Uranium	0.1	1	10	0.1	1	10	5	50	500
Vanadium	0.05	0.5	5	0.05	0.5	5	2.5	25	250
Zinc	0.02	0.2	2	0.02	0.2	2	2.5	25	250
Silica	0.16	1.6	16	1.083	10.83	108.3			

Appendix H

SAR Procedure

Soil

1.0 Prep soil by making a saturated paste with DI water and allowing it to sit at room temperature for 24 ± 2 hours. (See section 11.3.1 of the HEAL Resistivity SOP for guidance on the preparation of a saturated paste.) Enough paste must be made so that there is enough liquid after centrifugation to perform the analysis.

1.1 Prepare QC for every 10 samples at the same time the saturated paste is being prepared. An MB, LCS, and LCSD are required for each batch. For the MB, volume up a 50mL digestion tube with DI water and cap. The LCS and LCSD are prepared by filling a 50mL digestion tube and spiking each with 500µL of the Cation Spike.

2.0 Move paste into centrifuge tube(s) and centrifuge at a minimum of 1500rpm for 15 min.

2.1 QC does not need to be placed in the centrifuge.

3.0 Pipette off supernatant and analyze at dilution for Ca, Mg, and Na on ICP-OES.

4.0 On-scale results and any dilutions are entered into the spreadsheet located at:

H:\MS Office\Inorganics\USDA Methods\SAR Calculation.xls

Aqueous

1.0 No prep required. Samples are analyzed directly.

2.0 On-scale results and any dilutions are entered into spreadsheet mentioned above.

Appendix I

ICP-OES Metals Analysis Using SW-846 6010B Calibration Specification and Corrective Action Procedures

Analytical Method	Parameter	QC Element	Frequency	Acceptance Criteria	Corrective Action
SW-846 6010B	Metals	Calibration (3-4 points + blank minimum)	Daily prior to analysis	Correlation coefficient r ≥ 0.9995	1. Recalibrate.
		Initial Calibration Verification (ICV)	After calibration, prior to sample analysis	Results ±10% of assigned value RSD ≤ 5%	1. Reanalyze 1 additional time; report re-run. 2. Recalibrate if ICV still out.
		Initial Calibration Blank (ICB)	After calibration, prior to sample analysis	< Practical quantitation limit (PQL), ≥ -PQL	1. Reanalyze 1 additional time; report re-run. 2. Failing analytes cannot be reported. Recalibrate and reanalyze. 3. If still failing, clean system, recalibrate.
		Interference Check Standard + Elements (ICS Elem)	At the beginning of each run.	Results ± 25% of assigned value	1. Reanalyze 1 additional time; report re-run. 2. Analytes that are failing low cannot be reported for entire run. Recalibrate and reanalyze. 3. Analytes that are failing high may be reported if samples are ND for those analytes. Otherwise recalibrate and reanalyze.
		Interference Check Standard (ICS)	At the beginning of each run.	< 2 times the PQL	1. Reanalyze 1 additional time; report re-run. 2. Analytes that are failing high may be reported if samples are ND for those analytes. Otherwise recalibrate and reanalyze.
		Method Blank (MB)	1 per preparation/analysis batch (≤ 20 samples)	< PQL or must not contain analytes, > 5% of the lowest reported sample concentration, ≥ 2 times -PQL	1. Reanalyze MB and associated samples. 2. If MB contains analytes above the PQL and the associated samples are ND, the analyte can be reported for those samples. 3. If MB contains analytes above the PQL but less than 5% of lowest sample concentration fill out corrective action and report results. 4. Any analytes with a concentration less than 2x the negative PQL cannot be reported. 5. Reprep and reanalyze analytical batch as appropriate
		Laboratory Control Spike (LCS)	1 per preparation/analysis batch (≤ 20 samples)	% Recovery ±20% of assigned value	1. Reanalyze LCS and associated samples. 2. Analytes that are failing high may be reported if samples are ND for those analytes. Otherwise recalibrate and reanalyze. 3. Reprep and reanalyze analytical batch as appropriate.
		Laboratory Control Spike Duplicate (LCSD)	1 per preparation/analysis batch for SAR/ESPs or when sample volume does not allow an MSD to be prepped. (≤ 20 samples)	% Recovery ±20% of assigned value Relative percent difference (RPD) ≤ 20%	1. Reanalyze LCSD and associated samples. 2. Analytes that are failing high may be reported if samples are ND for those analytes. Otherwise recalibrate and reanalyze. 3. If RPD >20 reanalyze and report reanalysis. Fill out corrective action and report. 4. Reprep and reanalyze analytical batch as appropriate.

Continued on next page

Appendix I

ICP-OES Metals Analysis Using SW-846 6010B Calibration Specification and Corrective Action Procedures

Analytical Method	Parameter	QC Element	Frequency	Acceptance Criteria	Corrective Action
SW-846 6010B	Metals	Matrix Spike (MS)	1 per preparation/analysis batch (≤ 20 samples)	% Recovery ±25% of assigned value	1. Asses data. If analyte >100 mg/L in the sample, no recovery needed. 2. If <100 mg/L perform post spike on lowest reported dilution.
		Matrix Spike Duplicate (MSD)	1 per preparation/analysis batch (≤ 20 samples)	% Recovery ±25% of assigned value Relative percent difference (RPD) ≤20%	1. Asses data. If analyte >100 mg/L in the sample, no recovery needed. 2. If <100 mg/L perform post spike on lowest reported dilution. 3. If RPD >20 reanalyze and report reanalysis. Fill out corrective action and report for analytes >10x PQL.
		Post Spike (PS)	1 per analysis batch if MS, MSD fails. (See Appendix F)	% Recovery ±20% of assigned value	1. Open CAR for failing analytes whose concentrations fall within 10 to 100x their PQL. 2. Perform dilution test, report analyte(s).
		Dilution Test (DL)	1 per analysis batch if post spike fails. (See Appendix F)	RPD <10% for 1:5 dilution	1. Report and include in CAR.
		Continuing Calibration Verification (CCV)	Every 10 samples and the end of the analytical sequence	Results ±10% of assigned value RSD ≤ 5%	1. Reanalyze 1 additional time; report re-run. 2. Analytes that are failing high may be reported if samples are ND for those analytes. Otherwise reanalyze affected samples. 3. Analytes failing low cannot be reported, reanalyze affected samples preceding and following the failed CCV. 4. If RSD fails, reanalyze affected samples.
		Continuing Calibration Blank (CCB)	Every 10 samples after each CCV and the end of the analytical sequence	< PQL, ≥ -PQL	1. Reanalyze 1 additional time; report re-run. 2. Analytes that are failing high may be reported if samples are ND for those analytes. Otherwise reanalyze affected samples. 3. Analytes failing low cannot be reported, reanalyze affected samples preceding and following the failed CCB.

Hall Environmental Analysis Laboratory

SOP Acknowledgement Sheet
SOP Name-300.0
SOP Number- S-300.0-9

I state that I have read the above referenced SOP. I understand all portions of the document as they apply to me and my responsibilities at Hall Environmental Analysis Laboratory. I further recognize this as the most current application of the SOP and that it supersedes any previous documents that I may have utilized.

Effective 4/20/11-8/31/11

Name	Signature	Date	Supervisor Initials	QA Officer Initials
Ian Cameron		4/25/11		
Tiffany Shaw		4/28/11		
Lavonda Begay		4/21/11		
Stacey McCoy	Stacey McCoy	4/22/11		

Hall Environmental Analysis Laboratory

Common Name: EPA Method 300.0	S.O.P. Number: S-300.0-9
Method: Determination of Inorganic Anions by Ion Chromatography	
Effective Date: April 20, 2011	Supersedes: S-300.0-8
Laboratory Manager	Quality Assurance/Quality Control Officer:

1. Scope and Application

1.1. Ion chromatography (IC) is used to determine the concentration of inorganic anions summarized in Table 1. This Standard Operating Procedure (SOP) is applicable to anions in drinking water, groundwater, domestic and industrial waste waters, reagent waters, and aqueous extracts of solids.

Table 1: Anion List

Anion	Chemical Formula
Fluoride	F
Chloride	Cl
Nitrite as N	NO_2-N
Bromide	Br
Nitrate as N	NO_3-N
Phosphate as P	PO_4-P
Sulfate	SO_4

1.2 Method 300.0 is a liquid chromatographic (LC) technique. This method is restricted to use by, or under the supervision of analysts experienced in the use of LC, and skilled in the interpretation of chromatographs. Each analyst must demonstrate the ability to generate acceptable results with this method. Each analyst must establish a Demonstration of Capability (DOC) in accordance with the HEAL QA Plan.

2. Summary of Method

2.1. A 25uL sample aliquot is injected into an Ion Chromatograph system using an auto, self-regenerating conductivity suppression system. A carbonate-bicarbonate solution is used as the mobile phase.

2.2. The sample is swept onto a guard column first and then onto the analytical column for the separation of the anions. Anions are separated into discrete bands based on their affinity for the exchange sites of the stationary phase of the analytical column. The separated anions in their acid forms are measured using an electrical-conductivity cell. Anions are identified based on their retention times compared to known standards.

2.3. An extraction procedure must be performed prior to analysis of solid samples.

3. Detection Limits

3.1. Method Detection Limits (MDL) shall be performed every six months. An MDL study is to be conducted for both aqueous samples and aqueous extracts of solid samples. To view the current MDLs, see attached addendum. See **Appendix A** for a summary of the current PQLs.

3.2. MDLs must also be completed as part of a new analysts Initial Demonstration of Capability (IDOC).

4. Definitions

4.1. De-ionized (DI) Water – De-ionized water/reagent free water. Water that is passed through a purification system and is free from interferences.

4.2. Initial Calibration Verification (ICV) – Second source standard used to validate a new calibration immediately after calibration.

4.3. Initial Calibration Blank (ICB) – Analysis of a blank solution (DI water) after the initial calibration and ICV to check that the blank solution is free of target analytes.

4.4. Continuing Calibration Verification (CCV) – Standard from the same source as the calibration curve. Used prior to sample analysis on days when calibrations are not performed and periodically, there after, to check instrument stability.

4.5. Continuing Calibration Blank (CCB) – Analysis of a blank solution (DI water). Used as a periodic check that instrument is free of target analytes.

4.6. Laboratory Control Sample (LCS) - An aliquot of DI water to which known amounts of method analytes are added prior to sample preparation and analysis. The LCS is treated like a sample and used to determine whether the methodology is in control, and whether the laboratory is capable of making accurate and precise measurements.

4.7. Matrix Spike (MS) - An aliquot of field sample to which known amounts of method analytes are added prior to sample preparation and analysis. The MS is treated like a sample and used to determine whether sample matrix contributes bias to the analytical results. The background concentration in the un-spiked sample matrix must be determined separately. The MS concentrations are corrected for the background concentrations.

4.8. Matrix Spike Duplicate (MSD) – Treated exactly the same as the MS sample. Also used to examine the precision of the analytical process.

4.9. Method Blank (MB) - An aliquot of DI water or other blank matrices that are treated exactly the same as a sample including exposure to all glassware, equipment, solvents and reagents that are used in the preparation and analysis of field samples. The MB is to determine if any target analytes are present in the laboratory environment, the reagents, or the apparatus.

4.10. Method Detection Limit (MDL) – The constituent concentration that when processed through the complete method, produces a signal with a 99% probability that it is different than the blank.

4.11. Practical Quantitation Limit (PQL)/Reporting Limit – The minimum limit to which an analyte can be routinely reported.

4.12. Linear Calibration Range (LCR) – The linearity of the calibration curves (or LCR) for all analytes must be demonstrated during initial set-up of the instrument, when a calibration model is changed, or when a significant change in instrument response has occurred. Linearity must also be verified at least every six months.

5. Interferences

5.1. Interference can be caused by substances with retention times that are similar to and overlap those of the anion of interest. Large amounts of an anion can also interfere with the peak resolution of an adjacent anion. Sample dilution can be used to resolve most interference problems associated with retention times and elevated sample concentrations.

5.2. Method interference may be caused by contaminants in the DI water, sample processing chemicals or analysis apparatus.

5.3. Samples that contain large particles, large amounts of sediment or are abnormally discolored should be diluted prior to being analyzed to prevent damage to the instrument columns and flow systems.

5.4 Any anion that is not retained by the column or only slightly retained will elute quickly in the analysis and may interfere with early eluting peaks. Co-elution is caused by carbonates or other small organic anions and most significantly affect lower target analyte concentration.

6. Safety

6.1. The toxicity or carcinogenicity of each reagent or standard has not been fully defined. For handling procedures and exposure information always refer to the MSDS. Exposure to reagents and standards must be kept to a minimum. Gloves, lab coats, and safety glasses must be worn when handling samples, standards, or reagents. Refer to current HEAL Health and Safety SOP for more details.

7. Apparatus and Materials

7.1. Ion chromatograph Systems-

7.1.1. Dionex ICS-1000 IC systems

7.1.1.1. IC systems with conductivity detection.

7.1.1.2. AS-40 autosampler.

7.1.1.3. Dionex IonPac AG14A 4 x 50mm guard column.

7.1.1.4. Dionex IonPac AS14A 4 x 250mm analytical column.

7.1.1.5. Dionex Anion Self-Regenerating Suppressor, ASRS-ultra, 4mm.

7.1.1.6. Four liter eluent bottle with step-down regulator.

7.1.1.7. Dionex 5mL Auto sampler vials and filter caps. (pt. no. 38141).

7.1.2. Dionex ICS-1500 IC System

7.1.2.1. Dionex AS-50 autosampler, configured for use with 2mL vials.

7.1.2.2. 2mL polypropylene 2mL autosampler vials with split-top silicon/Teflon septum.

7.1.2.3. Dionex IonPac AS22-Fast 4 x 150mm analytical column.

7.1.2.4. Dionex IonPack AG22-Fast guard column.

7.1.2.5. Dionex Anion Self-Regenerating Suppressor, ASRS-ultra, 4mm.

7.1.2.6. Four liter eluent bottle with step-down regulator.

7.2. Operating Conditions/Integration Settings

7.2.1. Operating conditions are detailed in **Appendix B**.

7.2.2. Integration settings are detailed in **Appendix D**.

7.3. Analytical balance capable of weighing to the nearest 0.0001g.

7.4. Top loading balance capable of weighing to the nearest 0.01g.

7.5. Handheld repipettors and tips, 100uL, 1000uL, 5mL, and 10mL.

7.6. Volumetric flasks of assorted volumes.

7.7. Volumetric pipettes of assorted volumes.

7.8. Glass and plastic syringes of assorted volumes.

7.9. Auto-pipettor – 50ml.

7.10. Horizontal shaker.

7.11. Centrifuge tubes, skirted bottoms – 50mL

7.12. Centrifuge.

8. Reagents and Standards

8.1. DI water.

8.2. Sodium Bicarbonate, ACS Grade 'Baker Instra-Analyzed' – JT Baker (pt. no. 3506-01) or equivalent.

8.3. Sodium Carbonate, Anhydrous, Powder, Flux Grade 'Baker Instra-Analyzed' – JT Baker (pt. no. 4502-01) or equivalent.

8.4. Mobile Phase (Eluent) Preparation

8.4.1. Mobile Phase Preparation for ICS-1000 using AS/AG-14A analytical/guard columns

8.4.1.1. Carbonate/Bicarbonate stocks

8.4.1.1.1. 1600 mM Sodium Carbonate (Na_2CO_3) – Dissolve approximately 42.3941g sodium carbonate salt into 250 mL DI water. Transfer to a clean 250 mL plastic bottle and store at room temperature when not in use. Remake every 6 months.

8.4.1.1.2. 400 mM Sodium Bicarbonate ($NaHCO_3$) – Dissolve approximately 16.7808g of sodium bicarbonate salt into 500 mL DI. Transfer to a clean 500 mL plastic bottle and store at room temperature when not in use. Remake every 6 months.

8.4.1.2. Preparation of Mobile Phase (Eluent)

8.4.1.2.1. Close eluent regulator valve to release pressure on the eluent bottle.

8.4.1.2.2. Remove 4L eluent bottle from the instrument.

8.4.1.2.3. Pour out remaining eluent. Dilute down the drain.

8.4.1.2.4. Using 20mL volumetric pipette add 20mL of 1600 mM Na_2CO_3 stock solution to a 1L volumetric flask. To the same 1L flask, using a 10mL volumetric pipette add 10mL of 400 mM $NaHCO_3$ stock solution. Bring flask to 1L final volume.

8.4.1.2.5. Transfer the contents of the 1L flask to the 4L eluent bottle.

8.4.1.2.6. Using the same 1L volumetric flask, fill to the 1L mark with DI and transfer to the eluent bottle. Repeat with two more successive 1L additions. Total volume of eluent should be 4L.

8.4.1.2.7. Cap eluent bottle, shake well, and reconnect to instrument.

8.4.1.2.8. Using the regulator, adjust the bottle pressure to 7psi.

8.4.1.2.9. On the instrument control screen, click the pump settings button and drag eluent bottle arrow to 4L.

8.4.1.2.10. Every time the eluent is changed the instrument must be primed. Open the pump head valve by turning it clockwise. Insert the 10ml syringe and click the ON button on the pump settings screen. Remove approximately 35ml of eluent, click the OFF button and close the valve. Open the secondary pump valve, click the PRIME button to prime the pump until there are no bubbles exiting the waste line and close the valve.

8.4.2. Mobile Phase Preparation for ICS-1500 using AS/AG22-Fast Analytical/Guard Columns

8.4.2.1. Carbonate/Bicarbonate stock

8.4.2.1.1. 450 mM Sodium Carbonate (Na_2CO_3)/140 mM Sodium Bicarbonate ($NaHCO_3$)– Dissolve approximately 23.8522g sodium carbonate salt and 5.8825g of sodium bicarbonate salt into 500mL DI water. Transfer to a clean 500 mL plastic bottle and store at room temperature when not in use. Remake every 6 months.

8.4.2.2. Preparation of Mobile Phase (Eluent)

8.4.2.2.1. Prepare the mobile following the same steps as the ICS-1000s using the AG/AS-14A column set with the following exception.

8.4.2.2.2. Close eluent regulator valve to release pressure on the eluent bottle.

8.4.2.2.3. Remove 4L eluent bottle from the instrument.

8.4.2.2.4. Pour out remaining eluent. Dilute down the drain.

8.4.2.2.5. Prepare the eluent using a 10mL volumetric pipette add 10mL of 450 mM Na_2CO_3/140 mM $NaHCO_3$ stock solution to a 1L volumetric flask. Bring flask to 1L final volume.

8.4.2.2.6. Transfer the contents of the 1L flask to the 4L eluent bottle.

8.4.2.2.7. Using the same 1L volumetric flask, fill to the 1L mark with DI and transfer to the eluent bottle. Repeat with two more successive 1L additions. Total volume of eluent should be 4L.

8.4.2.2.8. Cap eluent bottle, shake well, and reconnect to instrument.

8.4.2.2.9. Using the regulator, adjust the bottle pressure to 7psi.

8.4.2.2.10. On the instrument control screen, click the pump settings button and drag eluent bottle arrow to 4L.

8.4.2.2.11. Every time the eluent is changed the instrument must be primed. Open the pump head valve by turning it clockwise. Insert the 10ml syringe and click the ON button on the pump settings screen. Remove approximately 35ml of eluent, click the OFF button and close the valve. Open the secondary pump valve, click the PRIME button to prime the pump until there are no bubbles exiting the waste line and close the valve.

8.5. Stock Standard Solutions

8.5.1. Calibration standards are purchased as both individual anions and custom mixed standards. Each standard is NIST traceable. See **Appendix C** for details. Calibration standards include:

8.5.1.1 Custom Mixed Stock Standard containing F (40mg/L), Cl (200mg/L), Br (200mg/L), Nitrate as N (120mg/L), Phosphate (200mg/L), and Sulfate (500mg/L).

8.5.1.2 Custom Stock Standard – Low Level containing F (10mg/L), Cl (50mg/L), Bromide (10mg/L), Nitrate (10mg/L), Phosphate (50mg/L), Sulfate (50mg/L).

8.5.1.3 Nitrite as N 1000mg/L standard.

8.5.2. Second source standards are purchased from a separate vendor as that of the primary calibration standards. All second source standards are purchased as individual anions. Each standard is NIST traceable at 1000mg/L. See **Appendix C** for details.

8.6. Calibration Standards

8.6.1. Calibration standards are made on a routine basis; every 3 months, when they show signs of degradation or if a comparison with a check standard indicates a problem.

8.6.2. Both stock and working standards must be stored at 0-6°C when they are not in use. They may be left out on the counter during the day while the instrument is being loaded but should be stored in the refrigerator overnight.

8.6.3. All 1000mg/L stock standards and custom stock standards **must** first be brought to room temperature before preparation of the working standards.

8.6.4. Working standards can be used directly from the refrigerator. They do not need to be at room temperature.

8.6.5. All calibration standards are prepared in DI water using volumetric pipettes, volumetric flasks, and hand-held repipettors. A summary of the calibration procedure is outlined in **Appendix C**.

8.6.6. A minimum of 5 calibration standards must be analyzed for each calibration.

8.7. Second Source Standards

8.7.1. An ICV standard is made on a routine basis; every 3 months, when it shows signs of degradation, or if a comparison with a check standard indicates a problem.

8.7.2. All 1000mg/L second source stock standards **must** first be brought to room temperature before preparation of the working ICV standard.

8.7.3. See **Appendix C** for a preparation summary of ICV.

8.8. Spike Solution

8.8.1. Use the A10 (mixed anion stock) 2nd source standard as the spiking solution.

8.8.2. Aqueous Samples

8.8.2.1. For LCS spike 250uL of spike solution to 4.75mL DI for a final volume of 5mL. Cap and invert to mix.

8.8.2.2. For MS/MSD spike 250uL of spike solution to 4.75mL sample for a final volume of 5mL. Cap and invert to mix.

8.8.3. Soil Samples

8.8.3.1. For LCS spike 1.5mL of spike solution into the centrifuge tube. Add 28.5mL DI on top of the spike solution (prepare 28.5mL DI water by filling a clean centrifuge tube with 30mL DI water, remove 1.5mL with the

repipettor, and pour the 28.5mL of DI on top of the LCS). Final volume will be 30mL.

8.8.3.2. For MS/MSD samples, add 10g of soil to centrifuge tube, spike 1.5mL of spike solution on the dry soil, then add 28.5mL of DI on top of sample and spike (prepare 28.5mL DI water by filling a clean centrifuge tube with 30mL DI water, remove 1.5mL with the repipettor, and pour the 28.5mL of DI on top of the MS sample. Repeat with MSD sample). Final volume will be 30mL.

9. Sample Collection, Preservation, Shipment, and Storage

9.1. Aqueous Samples

9.1.1. All aqueous samples must be collected in clean plastic or glass sampling containers.

9.1.2. After login all aqueous samples must be stored at above freezing to 6°C until analysis.

9.1.3. Aqueous anions fluoride, chloride, bromide, and sulfate must be reported from the unpreserved container. Holding time for these anions is 28 days.

9.1.4. Aqueous nitrite (NO_2), nitrate (NO_3) and phosphate (PO_4) have a hold time of 48 hours. They are reported as individual species from the unpreserved container.

9.1.4.1. Sample preserved with sulfuric acid extend the holding time to 28days, NO_2 and NO_3 are reported as a combined result (i.e. NO_2/NO_3). This is just the sum of the nitrite and nitrate values from the chromatogram. All sulfuric acid preserved samples must be diluted by at least 5x before analysis to minimize the amount of SO_4 being loaded on the column.

9.1.4.2. The holding time for phosphate cannot be extended with the use of a preservative.

9.2. Soil Samples

9.2.1. All soil samples must be collected in glass soils jars or plastic bags/containers. Nitrite, nitrate, phosphate, and sulfate samples must be stored at above freezing to 6°C until analysis. Fluoride, Chloride, and Bromide samples can be stored at room temperature.

9.2.2. Soil samples do not have an established holding time. Current HEAL protocol recommends the soil be prepared within 28days.

9.2.3. Aqueous extract of the soil has a 28day holding time for fluoride, chloride, bromide, and sulfate; and a **48hr holding time** for nitrite, nitrate, and phosphate.

10. Quality Control

10.1. Calibration

10.1.1. Instrument calibration is performed on an as needed basis.

10.1.2. A seven point curve is analyzed on a routine basis.

10.1.3. An ICV and ICB are analyzed for all anions.

10.1.4. The calibration is based upon the analyte area versus the analyte concentration. The calibration models used are linear average response, linear with 1/X weighting, and linear with $1/X^2$ weighting. Refer to current HEAL Quality Assurance plan for calibration details.

10.1.5. Calibration levels are summarized in **Appendix C**.

10.2. Continuing Calibration

10.2.1. A CCV and CCB are run at the beginning of each analytical batch (if not calibrating), every 10 sample injections and at the end of each run. The concentration of the CCV must be varied throughout the course of each analytical batch.

10.3. Batch QC Requirements

10.3.1. For aqueous and soil samples a MB and LCS must be analyzed every 20 new client samples. A MS and MSD must be analyzed every 10 new client samples (Batch QC is not required for dilutions of previously analyzed samples).

11. Sample Preparation and Analysis

11.1. Aqueous Sample Preparation

11.1.1. Using 5mL Dionex auto sampler vials fill to the 5ml mark on the vial.

11.1.2. Using 2mL Autosampler Vials, fill to at least the 1mL mark on vial.

11.1.3. Dilute samples into DI water.

11.2. Extraction of soils

11.2.1. Use 50mL centrifuge tubes for sample preparation.

11.2.2. Set up Prep in LIMS.

11.2.3. An MB and LCS must be prepared for every 20 samples.

11.2.4. An MS and MSD must be prepared for every 10 samples.

11.2.5. Label centrifuge tubes with appropriate HEAL number.

11.2.6. Using tabletop balance, weigh 10.00g (+/- 0.05g) of sample into each centrifuge tube.

11.2.7. To the MB and LCS do not add anything. To the MS and MSD add 10.00g of the sample.

11.2.8. Spike the appropriate amount of spike into the LCS, MS, and MSD.

11.2.9. Prepare three centrifuge tubes with 28.5mL DI.

11.2.10. Pour 28.5mL DI water on top of the LCS, MS, and MSD samples.

11.2.11. Using the auto-pipettor add 30mL DI water to the remaining centrifuge tubes. Be sure to add 30mL DI to the MB tube.

11.2.12. Agitate the samples on a horizontal shaker for 15 minutes. Remove from shaker and centrifuge for 15 minutes at approximately1500rpm.

11.2.13. For analysis, pipette or pour the aqueous extract portion into a 5mL-sample vial. Dilute as needed into DI depending on the amount of soil and debris suspended in solution.

11.3. Analysis

11.3.1. Open Chromeleon and press the "Start-Up" button on the instrument control screen. This begins the IC pumping. Allow approximately 10-20 minutes for the conductivity to stabilize.

11.3.2. Prepare standards and QC samples in the 5mL Dionex or 2mL auto sampler vials.

11.3.3. Select previous day's sequence from the list of sequences on the left of the screen, or press both Control and Tab keys together to switch to the sequence screen.

11.3.4. In the File menu click "Save as" to save the sequence file as a new name. (Label as the date, i.e.: 06-26-06). Make sure "save raw data" box is checked to carry the calibration data over to the new sequence.

11.3.5. Delete all the sample files except the calibration files at the top of the sequence.

11.3.6. Add the samples in the sequence. Starting with a conditioning blank, CCV, CCB, MB, and LCS. Save sequence by clicking the disk icon in the upper left corner of the screen. Samples with short hold times may be analyzed before the MB and LCS.

11.3.7. For the AS-50 autosampler the location of the sample, QC, etc. must be specified in the analytical sequence.

11.3.8. For the AS-40 autosampler, place the conditioning blank, CCV, CCB, MB, and LCS in the auto sampler rack. Holding the rack with the open side toward you add the samples in the rack from left to right.

11.3.9. Place in the AS-40 auto sampler with the belt cogs (just flip the rack around) facing you. Once the rack is seated properly, the belt and the rack are lined up, the spring loaded rack holder will be securely pushing on the rack.

11.3.10. Press Hold/Run button on the front panel of the AS-40 auto sampler. The first sample should move into injection position.

11.3.11. Check the conductivity on the instrument control window to make sure it has stabilized. Record conductivity and pressure from the instrument panel screen into the maintenance logbook.

11.3.12. At the top of the screen click "Batch" and "Select Sequence" to get to the sequence selection box.

11.3.13. Next click the "Add" button on the right, and select the sequence(s) from the correct instrument to be analyze. Add the Shutdown method at the end of the list.

11.3.14. Click the "Start" button on the bottom of the sequence selection box to trigger the auto sampler to begin with the first injection.

11.3.15. It is possible to continue to add samples once the analysis has begun. Save any changes that are made to the analytical sequence.

11.4. Data Import into LIMS

11.4.1. See **Appendix E** for a detailed outline of the data importation from Chromeleon to LIMS.

12. Calculations

12.1. Calculations for Aqueous samples:

$$Sample\ Amount,\ mg/L = A \times D$$

Where:

A = result from chromatogram, mg/L
D = dilution factor

12.2. Calculations for Non-Aqueous samples:

$$Sample\ Amount,\ mg/kg = A \times \frac{F}{I} \times \frac{1000g}{1kg} \times \frac{1L}{1000mL} \times D$$

Where:

A= result from chromatogram, mg/L
F = final volume of extracted sample, mL
I = initial volume (mass) of sample, g
D = dilution factor

13. Method Performance/Estimation of Uncertainty/DOCs/LCR

13.1. Method Performance

13.1.1. PT or "blind study" samples are routinely analyzed to check the accuracy of the anions analysis.

13.2. Estimation of Measurement Uncertainty

13.2.1. Estimation of uncertainty is to be conducted for all anions in each matrix on an annual basis using LCS results. Refer to current HEAL Quality Assurance plan for measurement uncertainty details.

13.3. Demonstration of Capability DOC

13.3.1. IDOCs are to be analyzed by all new analysts before handling client samples. IDOCs must be prepared from the second source stock solution. The IDOC for 300.0 must also include an MDL study. Before a new analyst can begin reporting client samples, the IDOC as outlined in the HEAL QAP and the MDL study must be completed.

13.3.2. ADOCPs (Annual Documentation of Continued Proficiency) are to be analyzed every year by all analysts working on the instrument. If four sequential LCSs are to be used they must be prepared from a second source. See HEAL QAP for more information.

13.4. Linear Calibration Range (LCR)

13.4.1. The linearity of the calibration curves (or LCR) for all analytes must be demonstrated during initial set-up of the instrument, when a calibration model is changed, or when a significant change in instrument response has occurred. Linearity must also be verified at least every six months. See **Appendix F** for LCR determination procedure.

13.4.2. For the LCR to be acceptable the calibration model employed must be linear, the ***r*** value (correlation coefficient) must be acceptable, and the recovery of each calibration point, when assessed against the curve, must be within +/-10% of the true value of that point. Once the initial LCR is established, the bi-annual LCR verification must meet the requirements outlined in the method.

14. Pollution Prevention

14.1. Small sample volumes are used (1-5mL).

14.2. Mobile phase (eluent) is sodium carbonate and sodium bicarbonate solution. It is made in the lab from bulk materials.

14.3. Sulfuric acid preserved bottles are usually sampled in 125mL plastics or in sulfuric preserved plastics that require additional tests.

14.4. Self-regenerating auto suppression does not require external (chemical) suppression of conductivity. Thus eliminating disposal of chemicals.

15. Data Assessment and Acceptance Criteria for Quality Control Measures

15.1. See **Appendix G** for a summary of the calibration, continuing calibration, and batch QC requirements.

15.2. Examination of Chromatograms

15.2.1. Identification of unknown anions in samples is based upon their retention times relative to a known standard. Peak shape and levels of interfering anions also influence correct identification.

15.2.2. Retention times, detection parameters, and integration information is detailed in **Appendix D.**

15.2.3. Retention Times

15.2.3.1. The retention times of each anion have been previously determined by comparison to a single anion standard.

15.2.3.2. The retention time windows for each anion are calculated by taking +/- a percentage of the absolute retention time of a calibration standard. For example if the absolute retention time of Chloride is 5.000 minutes the relative retention time value is 10%, the window has the range of 5.000 minutes +/- 10%. Or 4min 30sec to 5min 30sec.

15.2.3.3. Even though the instrument automatically identifies the anions of interest, it is still important to examine closely eluting unidentified peaks. This is done by overlaying the chromatogram of the unknown with that of the nearest CCV standard.

15.2.3.4. Slight variations of analyte retention time occur with mobile phase concentration differences, backpressure on the system, ambient column temperature, and high levels of anions in samples. Be sure to include these slight variations in the overall assessment of analyte identification.

15.3. Acceptance Criteria for Quality Control Measures

15.3.1. Initial Calibration

15.3.1.1. A multiple point linear average response, weighted 1/X, or weighted $1/X^2$ calibration is used. Calibration must have a correlation coefficient percent ≥ 99.5.

15.3.1.2. ICV must fall between 90-110% of the assigned value.

15.3.1.3. ICB must not contain analytes above the PQL.

15.3.2. Continuing Calibration

15.3.2.1. At the beginning of every new batch if not calibrating.

15.3.2.1.1. CCV recovery must fall between 90-110% of assigned value.

15.3.2.1.2. CCB must not contain analytes above the PQL

15.3.2.2. CCV recoveries must fall between 90-110% of assigned value.

15.3.2.3. CCB must not contain analytes above the PQL.

15.3.3. Batch QC

15.3.3.1. MB must not contain analytes above the PQL.

15.3.3.2. LCS spike concentrations should fall between the high and low standards of the calibration curve. LCS recoveries should fall between 90-110% of assigned value to be considered valid.

15.3.3.3. MS/MSD spike concentrations are the same as the LCS. Recoveries must fall between established control limits. See LIMS for current limits. Limits are updated at least annually. See current HEAL QAP for more information and calculations.

15.3.3.4. The RPD for the MSD must be less than 20%.

15.3.4. Peak Shape

15.3.4.1. Occasionally baseline anomalies are identified as analytes of interest. Analyte peaks in the sample should have the same shape as those in the standard. A wide peak may be identified as an anion of interest by the Chromeleon software, reanalyze for confirmation, if necessary, at dilution.

15.3.5. Levels of Interfering Anions

15.3.5.1. High levels of interfering anions (chloride and sulfate) can effect the position of the other anions on the chromatogram or completely mask/overlap a closely eluting anion peak of interest.

15.3.5.2. Common problems occur with high levels of chloride interfering with nitrite and high levels of sulfate interfering with phosphate. To accurately report a result for nitrite or phosphate when high levels of interferents are present, overlay the sample with a chromatogram of the nearest CCV standard. If the analyte is present and the peak can be integrated, then no dilution is necessary. Otherwise, dilute the sample to minimize the effect of the interferent and to allow accurate quantitation of the anion of interest.

15.3.5.3. Manual integrations are generally performed on nitrite (when high levels of chloride are present). The baseline must be drawn from valley-to-valley and reported. In the event that the analyst must perform a manual integration, they are to consult HEALs Manual Integration SOP, and utilize the experience and expertise of technical directors and senior analysts in order to properly and effectively execute the manual integration. A "before" and "after" printout of all manual integrations must be initialed and dated by the analyst who performed the integration, and included in the data packet.

15.3.5.4. Anions of interest that do not elute closely to the interfering peak may still be affected by that interferent. If a shift in retention time occurs, overlay the sample with a CCV standard to check for retention times shifts. A dilution may be necessary to minimize interference in the sample.

15.4. Data Assessment

15.4.1. Any analyte that quantitates above the highest calibration standard for a particular anion must be diluted. All dilutions are in DI water.

15.4.2. Occasionally, it is necessary to manually adjust the integration baseline to accurately quantitate a particular anion.

15.4.2.1. For example, a nitrite peak that elutes at the tail end of a large chloride peak. In this case, it is necessary to adjust the integration baseline from the beginning of the nitrite peak to the end of the nitrite peak, not including the area from the tail end of the chloride peak.

15.4.2.2. As a rule <u>never</u> adjust peak integration. Always default to how the instrument integrates a peak. Only adjust if absolutely necessary. Any chromatogram that is modified by the analyst must be stamped with the "Adjust baseline . . ." stamp and initialed by the IC analyst. See 16.2.5.3 above, for further information.

15.4.2.3. All analytes for MS/MSD samples should be reported, at all times, unless the following issues are encountered:

15.4.2.3.1. Any analytes in the MS/MSD exceed the upper calibration range.

15.4.2.3.2. Analyte concentration in a sample exceeds 4x the spike concentration.

15.4.2.3.3. Matrix or analyte concentration inhibits the response of any analyte.

15.4.2.3.4. When all analytes fail to recovered established control limits, rerun the MS/MSD samples.

15.4.2.3.5. When analyte recovery in the MS/MSD samples fail, and batch and QC have shown the analyte is in control, the analyte is reported on the MS/MSD to demonstrate the effect on matrix on the spike recovery.

15.4.2.3.6. NO_3, NO_2 and PO_4 are reported on the MS/MSD samples when they are analyzed when 48 hours of sample collection.

16. Corrective Actions for Out-of-Control Data

16.1. Initial Calibration

16.1.1. If the correlation coefficient percent is unacceptable, the failing analyte(s) must be recalibrated. If correlation coefficient continues to fail, remake standards from stocks and recalibrate. Perform instrument maintenance if necessary.

16.1.2. If the ICV sample should fail, reanalyze the standard one additional time. If ICV sample continues to fail remake ICV standard from stocks and reanalyze. Analytes that are failing will not be reported for any samples using that calibration. If it continues to fail recalibrate instrument.

16.1.3. If ICB fails, reanalyze one additional time. If ICB continues to fail, failing anions will not be reported for any samples using that calibration. Recalibrate instrument and reanalyze ICB.

16.2. Continuing Calibration

16.2.1. Failing anion(s) on the opening CCV or CCB may not be reported until the next acceptable CCV or CCB.

16.2.2. Samples prior to and following a failing CCV or CCB must be reanalyzed with acceptable CCV or CCB.

16.2.3. All of the samples before an unacceptable closing CCV or CCB must be reanalyzed with acceptable CCV or CCB.

16.2.4. When a CCB has hit above the PQL or the CCV recovery greater than 110%, all samples associated with the failures that are non-detect for analytes of interest, can be reported.

16.3. Batch QC

16.3.1. If the MB contains anions above the PQL contamination may be present, reanalyze MB one additional time. If contamination is still present, all samples associated with that MB must be reanalyzed (or re-extracted if associated with a soil extraction batch).

16.3.2. If the LCS is unacceptable, reanalyze one time. If the LCS still fails, all samples associated with that LCS must be reanalyzed (or re-extracted if associated with a soil extraction batch.)

16.3.3. If the MS or MSD recoveries are unacceptable, evaluate using the 4x rule.

16.3.3.1. The 4x rule states that if the sample concentration is greater than 4 times the spike concentration then it is not necessary to recover the spike.

16.3.3.2. If the 4x rule does not apply, examine the recoveries of the other anions (even if those anions are not being reported). If other anion recoveries are acceptable, an un-identified interference is likely.

16.3.3.3. If the recovery of any analyte falls outside the designated control limits and the laboratory performance for that analyte is shown to be in control, the recovery problem encountered with the MS/MSD is judged to be either matrix or solution related, not system related; therefore, no corrective action is necessary beyond qualifying the data for the failure.

16.3.3.4. Interferences associated with the matrix most commonly occur with the more reactive anions (NO_2, NO_3, and PO_4). If analyst error is suspected re-extract the sample, the MS, and the MSD. Otherwise flag the unacceptable recoveries and report the data.

16.3.3.5. Do not report MS/MSD results for analytes outside the range of reportability as defined by the curve.

16.3.4. If the RPD for the MSD is > 20%. Possible matrix inconsistency is suspected. A corrective action form may need to be filled out and results qualified and reported.

17. Contingencies for Handling Out-of-Control Data

17.1. All out-of-control data is flagged, a corrective action report is generated, and results reported to the client.

17.2. To notify end user of any failure not defined on the QA/QC Report. Including: analyst errors , deviations from the SOP, instrument failures, or in response to any internal or external audit findings.

18. Waste Management

18.1. All standards waste generated by the ICs will be disposed of according to the HEAL Chemical Hygiene Plan.

19. References

19.1. USEPA Method 300.0 Determination of Inorganic Anions by Ion Chromatography, US EPA Revision 2.1, (1993).

19.2. Chemical Hygiene Plan, HEAL.

19.3. Health and Safety SOP, HEAL.

19.4. Quality Assurance Plan, HEAL.

19.5. Manual Integration SOP, HEAL.

Appendix A

Method 300.0 - PQLs

	Aqueous	Soil
Anion	PQL (mg/L)	PQL (mg/kg)
Fluoride	0.1	0.3
Chloride	0.5	1.5
Nitrite as N	0.1	0.3
Bromide	0.1	0.3
Nitrate as N	0.1	0.3
Phosphate as P	0.5	1.5
Sulfate	0.5	1.5
Nitrite/Nitrate	0.2	

Appendix B

Dionex ICS-1000 Operating Conditions

ICS-1000 w/ AS-40 autosampler using AG/AS-14A Guard and Analytical Columns

Instrument Control File

```
        On
        Pressure.LowerLimit =                    200
        Pressure.UpperLimit =                    5000
        &A.Equate =                              "&A"
        LoadPosition
        Data_Collection_Rate =                   5
        CellTemperature =                        35
        ColumnTemperature =                      30
        Suppressor_Type =                        ASRS_4mm
        ; Pump_ECD.Carbonate =                  8
        ; Pump_ECD.Bicarbonate =                1
        ; Pump_ECD.Hydroxide =                  0
        ; Pump_ECD.Tetraborate =                0
        ; Pump_ECD.Other eluent =               0
        ; Pump_ECD.Recommended Current =        43
        Suppressor_Current =                     43
        ECD_Total.Step =                         0.2
        ECD_Total.Average =                      Off
        Channel_Pressure.Step =                  0.2
        Channel_Pressure.Average =               Off
        Flow =                                   1
        Pump_ECD_Relay_1.Open
 -2.300 Pump_ECD_Relay_1.Closed

  0.000 Pump_ECD_Relay_1.Open

  0.000 Autozero
        ECD_1.AcqOn
        ECD_Total.AcqOn
        Channel_Pressure.AcqOn
        Pump_InjectValve.InjectPosition
 15.000 ECD_1.AcqOff
        ECD_Total.AcqOff
        Channel_Pressure.AcqOff

        End
```

Summary of Control File

Flow rate (mL/min):	1.00
Conductivity Cell Temperature (°C)	35
Column Temperature (°C):	30
Supressor Current (mA):	43
Pressure Upper Limit (psi):	5000
Pressure Lower Limit (psi):	200

ICS-1000 w/ AS-50 autosampler using AG/AS-22 Fast Guard and Analytical Columns

Instrument Control File

```
        Sampler.AquireExclusive Access
        Pressure.LowerLimit =                    200
        Pressure.UpperLimit =                    3000
        &A.Equate =                              "&A"
        Flush                                    Volume=1000
        Wait                                     FlushState
        Needle Height =                          3 [mm]
        CutSegmentVolume =                       10 [ul]
        SyringeSpeed =                           4
        CycleTime =                              0 [min]
        WaitForTemperature =                     FALSE
        Data_Collection_Rate =                   5.0 [Hz]
        CellTemperature.Nominal                  35.0 [oC]
        ColumnTemperature.Nominal =              30.0 [oC]
        Supressor_Type =                         ASRS_4mm
        ; Pump_ECD.Carbonate =                  4.5
        ; Pump_ECD.Bicarbonate =                1.4
        ; Pump_ECD.Hydroxide =                  0
        ; Pump_ECD.Tetraborate =                0
        ; Pump_ECD.Other eluent =               0
        ; Pump_ECD.Recommended Current =        31
        Supressor_Current =                      31 [mA]
        ECD_Total.Step =                         0.20 [s]
        ECD_Total.Average =                      Off
        Channel_Pressure.Step =                  0.20 [s]
        Channel_Pressure.Average =               Off
        Flow =                                   1.2 [ml/min]
        Wait

  0.000 Autozero
        Load
        Wait                                     CycleTimeState
        Inject
        Wait                                     InjectState
        ECD_1.AcqOn
        ECD_Total.AcqOn
        Channel_Pressure.AcqOn
        Sampler.ReleaseExclusiveAccess

 10.000 ECD_1.AcqOff
        ECD_Total.AcqOff
        Channel_Pressure.AcqOff

        End
```

Summary of Control File

Flow rate (mL/min):	1.20
Conductivity Cell Temperature (°C)	35
Column Temperature (°C):	30
Supressor Current (mA):	31
Pressure Upper Limit (psi):	3000
Pressure Lower Limit (psi):	200

Appendix C

Calibration Procedure

Stock Standards

Individual Stock Standard

	Stock conc ug/ml
Nitrite	1000

80 ug/ml. NO_2. Volume in mLs.

	Stock conc ug/ml.	Std Volume	Conc. of Std	Final Volume
Nitrite	1000	16.0	80.0	200

10 ug/ml. NO_2 - Low Level. Volume in mLs.

	Stock conc ug/ml.	Std Volume	Conc. of Std	Final Volume
Nitrite	1000	1.0	10.0	100

Custom Mixed Stock Standard

	Stock Std Conc. ug/ml.
Fluoride	40
Chloride	200
Bromide	200
Nitrate	120
Phosphate	200
Sulfate	600

Custom Stock Standard - Low Level

	Stock Std Conc. ug/ml.
Fluoride	10
Chloride	50
Bromide	10
Nitrate	10
Phosphate	50
Sulfate	50

Calibration Standards

Volumes are in mLs. All calibration standards are made as dilution in the autosampler vials or small cups except A1 and A7.

A7 - 200 ml. total volume, Custom Stock and 80 mg/L NO_2.

	Custom Mixed Std. ug/ml	V1 Stock Volume	C2 Conc. of A7	V2 Final Volume
Fluoride	40	20.0	4	200
Chloride	200	20.0	20	200
Nitrite	80	20.0	8	200
Bromide	200	20.0	20	200
Nitrate	120	20.0	12	200
Phosphate	200	20.0	20	200
Sulfate	600	20.0	60	200

A1 - 200 ml. total volume, Low level custom stock and 10 mg/L NO_2.

	Low Level Custom ug/ml	V1 Stock Volume	C2 Conc. of A1	V2 Final Volume
Fluoride	10	2.00	0.10	200
Chloride	50	2.00	0.50	200
Nitrite	10	2.00	0.10	200
Bromide	10	2.00	0.10	200
Nitrate	10	2.00	0.10	200
Phosphate	50	2.00	0.50	200
Sulfate	50	2.00	0.50	200

A6 - A7 diluted 75% conc., 4.0 ml. total volume.

	C1 A7 ug/ml	V1 A7 Volume	C2 Conc. of A6	V2 Final Volume
Fluoride	4.00	3.0	3	4.0
Chloride	20.00	3.0	15	4.0
Nitrite	8.00	3.0	6	4.0
Bromide	20.00	3.0	15	4.0
Nitrate	12.00	3.0	9	4.0
Phosphate	20.00	3.0	15	4.0
Sulfate	60.00	3.0	37.5	4.0

A5 - A7 diluted x2, 4.0 ml. total volume.

	C1 A7 ug/ml	V1 A7 Volume	C2 Conc. of A5	V2 Final Volume
Fluoride	4.00	2.0	2	4.0
Chloride	20.00	2.0	10	4.0
Nitrite	8.00	2.0	4	4.0
Bromide	20.00	2.0	10	4.0
Nitrate	12.00	2.0	6	4.0
Phosphate	20.00	2.0	10	4.0
Sulfate	60.00	2.0	25	4.0

→

A3 - A5 diluted x8, 4.0 ml. total volume.

	C1 A5 ug/ml	V1 A5 Volume	C2 Conc. of A3	V2 Final Volume
Fluoride	2.0	0.5	0.25	4.0
Chloride	10.0	0.5	1.25	4.0
Nitrite	4.0	0.5	0.50	4.0
Bromide	10.0	0.5	1.25	4.0
Nitrate	6.0	0.5	0.75	4.0
Phosphate	10.0	0.5	1.25	4.0
Sulfate	25.0	0.5	3.125	4.0

A4 - A7 diluted x5, 5.0 ml. total volume.

	C1 A7 ug/ml	V1 A7 Volume	C2 Conc. of A4	V2 Final Volume
Fluoride	4.00	1.0	0.8	5.0
Chloride	20.00	1.0	4.0	5.0
Nitrite	8.00	1.0	1.6	5.0
Bromide	20.00	1.0	4.0	5.0
Nitrate	12.00	1.0	2.4	5.0
Phosphate	20.00	1.0	4.0	5.0
Sulfate	60.00	1.0	12.0	5.0

Appendix C

Calibration Procedure

A2 - A7 diluted x20, 5.0 mL total volume.

	C1 A7 ug/ml	V1 A7 Volume	C2 Conc. of A2	V2 Final Volume
Fluoride	4.00	0.25	0.2	5.0
Chloride	20.00	0.25	1.0	5.0
Nitrite	8.00	0.25	0.4	5.0
Bromide	20.00	0.25	1.0	5.0
Nitrate	12.00	0.25	0.5	5.0
Phosphate	20.00	0.25	1.0	5.0
Sulfate	50.00	0.25	2.5	5.0

Continuing Calibration Standards

Custom Mixed Stock Standard

	Stock conc ug/ml.
Fluoride	40
Chloride	200
Bromide	200
Nitrate	120
Phosphate	200
Sulfate	500

80 ug/ml. NO2

	Stock conc ug/ml.
Nitrite	80.0

A4 CCV - 200 mL total volume.

	Custom Mixed Std. ug/ml.	V1 CCV Std. Volume	C2 Conc. of A4 CCV	V2 Final Volume
Fluoride	40	4	0.8	200
Chloride	200	4	4.0	200
Nitrite	80	4	1.6	200
Bromide	200	4	4.0	200
Nitrate	120	4	2.4	200
Phosphat	200	4	4.0	200
Sulfate	500	4	10.0	200

A5 CCV - 200 mL total volume.

	Custom Mixed Std. ug/ml.	V1 CCV Std. Volume	C2 Conc. of A5 CCV	V2 Final Volume
Fluoride	40	10	2.0	200
Chloride	200	10	10.0	200
Nitrite	80	10	4.0	200
Bromide	200	10	10.0	200
Nitrate	120	10	6.0	200
Phosphat	200	10	10.0	200
Sulfate	500	10	25.0	200

A6 CCV - 200 mL total volume.

	Custom Mixed Std. ug/ml.	V1 CCV Std. Volume	C2 Conc. of A6 CCV	V2 Final Volume
Fluoride	40	15	3.0	200
Chloride	200	15	15.0	200
Nitrite	80	15	6.0	200
Bromide	200	15	15.0	200
Nitrate	120	15	9.0	200
Phosphat	200	15	15.0	200
Sulfate	500	15	37.5	200

Second Source Standards

	C1 Stock conc. ug/ml.	V1 Stock Volume	C2 Conc. of 2nd std.	V2 Final Volume
Fluoride	1000	2.0	10	200
Chloride	1000	20.0	100	200
Nitrite	1000	4.0	20	200
Bromide	1000	10.0	50	200
Nitrate	1000	10.0	50	200
Phosphate	1000	20.0	100	200
Sulfate	1000	40.0	200	200

IOV	C1 2nd Std. ug/ml.	V1 2nd Std. Volume	C2 Conc. of IOV	V2 Final Volume
Fluoride	10	20.0	1	200
Chloride	100	20.0	10	200
Nitrite	20	20.0	2	200
Bromide	50	20.0	5	200
Nitrate	50	20.0	5	200
Phosphate	100	20.0	10	200
Sulfate	200	20.0	20	200

LCS/MS	C1 2nd Std. ug/ml.	V1 2nd Std. Volume	C2 Conc. of LCS/MS	V2 Final Volume
Fluoride	10	0.25	0.5	5.0
Chloride	100	0.25	5.0	5.0
Nitrite	20	0.25	1.0	5.0
Bromide	50	0.25	2.5	5.0
Nitrate	50	0.25	2.5	5.0
Phosphate	100	0.25	5.0	5.0
Sulfate	200	0.25	10.0	5.0

Appendix D

Retention Times, Detection Parameters, and Intergration Information

Orion

Detection Table

No.	Retention Time (min)	Parameter	Value	Channel
1	0.000	Minimum Area	0.01 "[Signal]"min"	All Channels
2	0.000	Inhibit Integration	On	All Channels
3	2.800	Inhibit Integration	Off	All Channels
4	2.800	Frooting Sensitivity Factor	2	All Channels
5	4.110	Inhibit Integration	On	All Channels
6	4.120	Inhibit Integration	Off	All Channels
7	6.500	Minimum Area	0.001"[Signal]"min"	All Channels
8	9.000	Minimum Area	0.01 "[Signal]Min"	All Channels
9	9.001	Inhibit Integration	On	All Channels
10	9.002	Inhibit Integration	Off	All Channels

Peak Table

Use Recently Detected Retention Times: Off
Peak Retention Time Determination: Absolute
Delay Time of 2nd Detector: <None>
Delay Time of 3rd Detector: <None>

No.	Peak Name	Retention time (min)	Window	Standard	Integration Type	Calibration Type	Peak Type	Amount STD1	Amount STD2	Amount STD3	Amount STD4	Amount STD5	Amount STD6	Amount STD7
1	Fluoride	3.360	5.000 RG	External	Area	XLOff	Auto	0.10	0.20	0.25	0.80	2.00	3.00	4.00
2	Chloride	4.850	5.000 RG	External	Area	XLOff	Auto	0.50	1.00	1.25	4.00	10.00	15.00	20.00
3	Nitrite	5.700	5.000 RG	External	Area	XLOff	B-M-B	0.10	0.40	0.50	1.60	4.00	6.00	8.00
4	Bromide	7.100	5.000 RG	External	Area	XLOff	B-M-B	0.10	1.00	1.25	4.00	10.00	15.00	20.00
5	Nitrate	7.950	5.000 RG	External	Area	XXLOff	B-M-B	0.10	0.60	0.75	2.40	6.00	9.00	12.00
6	Phosphate	10.850	3.000 AF	External	Area	XLOff	Auto	0.50	1.00	1.25	4.00	10.00	15.00	20.00
7	Sulfate	12.900	5.000 RG	External	Area	XXLOff	Auto	0.50	2.50	3.13	10.00	25.00	37.50	50.00

Amount Table

Dimension of Amounts: mg/L
Reference volume for amounts: Use inject volume of first standard
Number of Amount Columns: 9
Sample column used for amount column assignment: Sample Name

No.	Peak Name	Retention time (min)	Response Factor	Amount STD1	Amount STD2	Amount STD3	Amount STD4	Amount STD5	Amount STD6	Amount STD7	Match Criteria	Threshold
1	Fluoride	3.360	1.000	0.10	0.20	0.25	0.80	2.00	3.00	4.00	Least Square	950
2	Chloride	4.850	1.000	0.50	1.00	1.25	4.00	10.00	15.00	20.00	Least Square	950
3	Nitrite	5.700	1.000	0.10	0.40	0.50	1.60	4.00	6.00	8.00	Least Square	950
4	Bromide	7.100	1.000	0.10	1.00	1.25	4.00	10.00	15.00	20.00	Least Square	950
5	Nitrate	7.950	1.000	0.10	0.60	0.75	2.40	6.00	9.00	12.00	Least Square	950
6	Phosphate	10.850	1.000	0.50	1.00	1.25	4.00	10.00	15.00	20.00	Least Square	950
7	Sulfate	12.900	1.000	0.50	2.50	3.13	10.00	25.00	37.50	50.00	Least Square	950

CAL TYPES: LOff = Linear ALOff = Linear Average Response XLOff = Linear 1/X Weighting XXLOff = Linear $1/X^2$ Weighting
Calibration Types and Retention Times will change as over time due to changes in the system.

Appendix D

Retention Times, Detection Parameters, and Intergration Information

Triton

Detection Table

No.	Retention Time (min)	Parameter	Value	Channel
1	0.000	Minimum Area	0.01 "[Signal]*min"	All Channels
2	0.000	Inhibit Integration	On	All Channels
3	2.950	Fronting Sensitivity Factor	2	All Channels
4	2.950	Inhibit Integration	Off	All Channels
5	4.110	Inhibit Integration	On	All Channels
6	4.120	Inhibit Integration	Off	All Channels
7	6.500	Minimum Area	0.001"[Signal]*min"	All Channels
8	8.500	Minimum Area	0.01 "[Signal]/Min"	All Channels

Peak Table

Use Recently Detected Retention Times: Off
Peak Retention Time Determination: Absolute
Delay Time of 2nd Detector: <None>
Delay Time of 3rd Detector: <None>

No.	Peak Name	Retention time (min)	Window	Standard	Integration Type	Calibration Type	Peak Type	Amount STD1	Amount STD2	Amount STD3	Amount STD4	Amount STD5	Amount STD6	Amount STD7
1	Fluoride	3.370	5.000 RG	External	Area	LOff	B-M-B	0.10	0.20	0.25	0.80	2.00	3.00	4.00
2	Chloride	4.780	5.000 RG	External	Area	XLOff	B-M-B	0.50	1.00	1.25	4.00	10.00	15.00	20.00
3	Nitrite	5.600	5.000 RG	External	Area	XLOff	B-M-B	0.10	0.40	0.50	1.60	4.00	6.00	8.00
4	Bromide	6.980	5.000 RG	External	Area	XLOff	B-M-B	0.10	1.00	1.25	4.00	10.00	15.00	20.00
5	Nitrate	7.850	5.000 RG	External	Area	XXLOff	B-M-B	0.10	0.60	0.75	2.40	6.00	9.00	12.00
6	Phosphate	10.420	10.000 RG	External	Area	XLOff	B-M-B	0.50	1.00	1.25	4.00	10.00	15.00	20.00
7	Sulfate	12.510	10.000 RG	External	Area	XLOff	B-M-B	0.50	2.50	3.13	10.00	25.00	37.50	50.00

Amount Table

Dimension of Amounts: mg/L
Reference volume for amounts: Use Inject volume of first standard
Number of Amount Columns: 9
Sample column used for amount column assignment: Sample Name

No.	Peak Name	Retention time (min)	Response Factor	Amount STD1	Amount STD2	Amount STD3	Amount STD4	Amount STD5	Amount STD6	Amount STD7	Match Criteria	Threshold
1	Fluoride	3.370	1.000	0.10	0.20	0.25	0.80	2.00	3.00	4.00	Least Square	950
2	Chloride	4.780	1.000	0.50	1.00	1.25	4.00	10.00	15.00	20.00	Least Square	950
3	Nitrite	5.600	1.000	0.10	0.40	0.50	1.60	4.00	6.00	8.00	Least Square	950
4	Bromide	6.980	1.000	0.10	1.00	1.25	4.00	10.00	15.00	20.00	Least Square	950
5	Nitrate	7.850	1.000	0.10	0.60	0.75	2.40	6.00	9.00	12.00	Least Square	950
6	Phosphate	10.420	1.000	0.50	1.00	1.25	4.00	10.00	15.00	20.00	Least Square	950
7	Sulfate	12.510	1.000	0.50	2.50	3.13	10.00	25.00	37.50	50.00	Least Square	950

CAL TYPES: LOff = Linear ALOff = Linear Average Response XLOff = Linear 1/X Weighting XXLOff = Linear $1/X^2$ Weighting
Calibration types and Retention Times will change as over time due to changes in the system.

Appendix D

Retention Times, Detection Parameters, and Intergration Information

Gemini

Detection Table

No.	Retention Time (min)	Parameter	Value	Channel
1	0.000	Minimum Area	0.005 "[Signal]*min"	All Channels
2	0.000	Void Volume Treatment	On	All Channels
3	0.000	Tailing Sensitivity Factor	6.0	All Channels
4	0.000	Inhibit Integration	On	All Channels
5	1.750	Inhibit Integration	Off	All Channels
6	2.480	Inhibit Integration	On	All Channels
7	2.500	Inhibit Integration	Off	All Channels
8	3.000	Void Volume Treatment	Off	All Channels
9	5.000	Inhibit Integration	On	All Channels
10	5.300	Inhibit Integration	Off	All Channels

Peak Table

Use Recently Detected Retention Times: Off
Peak Retention Time Determination: Absolute
Delay Time of 2nd Detector: <None>
Delay Time of 3rd Detector: <None>

No.	Peak Name	Retention time (min)	Window	Standard	Integration Type	Calibration Type	Peak Type	Amount STD1	Amount STD2	Amount STD3	Amount STD4	Amount STD5	Amount STD6	Amount STD7
1	Fluoride	1.957	5.000 RG	External	Area	XLOff	Auto	0.10	0.20	0.25	0.80	2.00	3.00	4.00
2	Chloride	2.809	5.000 RG	External	Area	XXLOff	Auto	0.50	1.00	1.25	4.00	10.00	15.00	20.00
3	Nitrite	3.384	5.000 RG	External	Area	XXLOff	Auto	0.10	0.40	0.50	1.60	4.00	6.00	8.00
4	Bromide	4.044	5.000 RG	External	Area	XLOff	Auto	0.10	1.00	1.25	4.00	10.00	15.00	20.00
5	Nitrate	4.602	5.000 RG	External	Area	XXLOff	Auto	0.10	0.60	0.75	2.40	6.00	9.00	12.00
6	Phosphate	5.932	5.000 RG	External	Area	XXLOff	Auto	0.50	1.00	1.25	4.00	10.00	15.00	20.00
7	Sulfate	6.770	5.000 RG	External	Area	XXLOff	Auto	0.50	2.50	3.13	10.00	25.00	37.50	50.00

Amount Table

Dimension of Amounts: mg/L
Reference volume for amounts: Use inject volume of first standard
Number of Amount Columns: 7
Sample column used for amount column assignment: Sample Name

No.	Peak Name	Retention time (min)	Response Factor	Amount STD1	Amount STD2	Amount STD3	Amount STD4	Amount STD5	Amount STD6	Amount STD7	Match Criteria	Threshold
1	Fluoride	1.957	1.000	0.10	0.20	0.25	0.80	2.00	3.00	4.00	Least Square	950
2	Chloride	2.809	1.000	0.50	1.00	1.25	4.00	10.00	15.00	20.00	Least Square	950
3	Nitrite	3.384	1.000	0.10	0.40	0.50	1.60	4.00	6.00	8.00	Least Square	950
4	Bromide	4.044	1.000	0.10	1.00	1.25	4.00	10.00	15.00	20.00	Least Square	950
5	Nitrate	4.602	1.000	0.10	0.60	0.75	2.40	6.00	9.00	12.00	Least Square	950
6	Phosphate	5.932	1.000	0.50	1.00	1.25	4.00	10.00	15.00	20.00	Least Square	950
7	Sulfate	6.770	1.000	0.50	2.50	3.13	10.00	25.00	37.50	50.00	Least Square	950

CAL TYPES: LOff = Linear ALOff = Linear Average Response XLOff = Linear 1/X Weighting XXLOff = Linear $1/X^2$ Weighting
Calibration types and Retention Times will change as over time due to changes in the system.

Appendix E

Procedure for Importing Data into LIMS

A. From previous days run, highlight samples from the ICV to the closing CCB.
B. Right click in the highlighted area, and choose batch report.
C. Check the printout box (if not already selected) and the export box.
   1. This will open a new window, select the Excel file format box, click next.
   2. In the new window, uncheck all except the summary option. Click finish, click ok to print and export.
D. Click OK when all have been printed and exported.
E. In the same days run, right click, choose print sequence.
   1. Click OK with boxes unchecked.
   2. Click OK to print.
F Open Microsoft Excel program.
   1. Go to File, Open.
   2. Find file from previous days run in G:/Omega/Data/300/Orion/MySequences or G:/Omega/Data/300/Triton/MySequences depending on which instrument is being imported.
   3. Open the run and double click on the last sample to retrieve all data from the run.
G. With the previous days run open, go to File, Open.
   1. Open the macro from G:/Omega/Data/300/19Macro3.
H. Return to the spreadsheet containing exported data .
   1. Go to Tools, Macro, Macros.
   2. Choose 'This Workbook' option from the 'Macros in:' drop-down list.
   3. Click Run.
I. From the 19Macro3 spreadsheet, copy anion headings (E1-L1). Paste to the spreadsheet that contains exported data (E1-L1). Data is now ready to be saved to the G:/Omega/Data/300 folder.
   1. Select Save As from File.
   2. Enter date and the instrument that data is from as the file name.
   3. Save, and keep open.
J. In the LIMS, open data entry.
   1. Enter Orion or Triton in Intstrument ID.
   2. Enter date that data is from in the Run Start Date box
   3. Select analyst who performed the run.
   4. Select Data Import
      a. In the Specification box, select Omega Excel File.
      b. Test Code will be 300_W, Sample type will be Samp (very important, do not leave either box empty or data will not import.)
      c. Import Data as: **Raw Result**.
      d. Click run import, Select the file that contains data of interest from the G:/Omega/Data/300 folder.
      e. A data range will need to be entered in the window that opens. The range will always begin with A1. To complete the range, refer to the excel spreadsheet that contains the imported data. The range will end with the AnalDate column and the row that contains the closing CCB. (ex. A1:L32).
      f. Click OK.

Appendix F

Procedure for Determining LCR

1. Perform instrument maintenance to ensure optimal instrument performance.

2. Calibrate instrument using all curve calibration points. Remember to analyze ICV and ICB.

3. Using the Arizona Calibration Workbook, enter all of the peak areas and standard concentrations into the "Initial Calibration" section of the spreadsheet.

4. The spreadsheet then calculates the results for linear average response, linear regression, and quadratic regression.

5. Copy and paste the concentrations and areas from the "Initial Calibration" section of the work sheet to the "Sample Calculations" section of the work sheet

6. As a final check include the ICV concentration and area in the "Sample Calculations" section.

7. In the "Sample Calculations" section, the spreadsheet details the results of four linear models (average response, equal weighing, 1/X weighing, and $1/X^2$ weighing). It also includes a non-liner quadratic calibration model.

8. For the LCR to be acceptable the calibration model employed must be linear, the *r* value (correlation coefficient) must be acceptable, and the recovery of each calibration point, when assessed against the curve, must be within +/-10% of the true value of that point.

Appendix G

Inorganic Anions Analysis Using EPA 300.0
Calibration Specification And Corrective Action Procedures

Analytical Method	Parameter	QC Element	Frequency	Acceptance Criteria	Corrective Action
EPA 300.0	Wet Chemistry	Initial Calibration (7-9 points)	Periodically; prior to sample analysis	Correlation coefficient r ≥ 99.5	1. Recalibrate 2. Remake calibration standards from stocks and recalibrate 3. Perform instrument maintenance, recalibrate
		Initial Calibration Verification (ICV)	After initial calibration, prior to sample analysis	Results ±10% of assigned value	1. Reanalyze ICV, one additional time. 2. Initial Calibration: recalibrate instrument, and reanalyze. 3. Initial Calibration: Remake ICV standards, recalibrate, and reanalyze.
		Initial Calibration Blank (ICB)	After calibration, prior to sample analysis	< Practical quantitation limit (PQL)	1. Reanalyze one additional time 2. Recalibrate instrument
		Method Blank (MB)	1 per preperation/analysis batch (≤ 20 samples)	< PQL	1. Reanalyze method blank, one additional time 2. Reprep/reanalyze analytical analytical batch as appropriate
		Laboratory Control Sample (LCS)	1 per preperation/analysis batch (≤ 20 samples)	% Recovery ± 10% of assigned value	1. Reanalyze LCS, one additional time 2. Reprep/reanalyze LCS and affected samples
		Matrix Spike (MS)	1 per preperation/analysis batch (≤ 10 samples)	% Recovery must fall between established limits	1. Asses data (4 x rule) 2. Narrate all outliers
		Matrix Spike Duplicate (MSD)	1 per preperation/analysis batch (≤ 10 samples)	% Recovery must fall between established limits Relative percent difference (RPD) <20%	1. Asses data (4 x rule) 2. Narrate all outliers 3. RPD: Narrate all outliers
		Continuing Calibration Verification (CCV)	At the beginning, every 10 samples, and at the end of the analytical sequence	Results ±10% of assigned value	1. Reanalyze affected samples back to the last acceptable CCV and to the next acceptable CCV
		Continuing Calibration Blank (CCB)	At the beginning, every 10 samples, and at the end of the analytical sequence	< Practical quantitation limit (PQL)	1. Reanalyze affected samples back to the last acceptable CCB and to the next acceptable CCB

3-1

Hall Environmental Analysis Laboratory

Common Name: EPA Method 300.0	S.O.P. Number: S-300.0-10
Method: Determination of Inorganic Anions by Ion Chromatography	
Effective Date: August 31, 2011	Supersedes: S-300.0-9
Laboratory Manager	Quality Assurance/Quality Control Officer:

1. Scope and Application

1.1. Ion chromatography (IC) is used to determine the concentration of inorganic anions summarized in Table 1. This Standard Operating Procedure (SOP) is applicable to anions in drinking water, groundwater, domestic and industrial waste waters, reagent waters, and aqueous extracts of solids.

Table 1: Anion List

Anion	Chemical Formula
Fluoride	F
Chloride	Cl
Nitrite as N	NO_2-N
Bromide	Br
Nitrate as N	NO_3-N
Phosphate as P	PO_4-P
Sulfate	SO_4

1.2 Method 300.0 is a liquid chromatographic (LC) technique. This method is restricted to use by, or under the supervision of analysts experienced in the use of LC, and skilled in the interpretation of chromatographs. Each analyst must demonstrate the ability to generate acceptable results with this method. Each analyst must establish a Demonstration of Capability (DOC) in accordance with the HEAL QA Plan.

2. Summary of Method

2.1. A 25uL sample aliquot is injected into an Ion Chromatograph system using an auto, self-regenerating conductivity suppression system. A carbonate-bicarbonate solution is used as the mobile phase.

2.2. The sample is swept onto a guard column first and then onto the analytical column for the separation of the anions. Anions are separated into discrete bands based on their affinity for the exchange sites of the stationary phase of the analytical column. The separated anions in their acid forms are measured using an electrical-conductivity cell. Anions are identified based on their retention times compared to known standards.

2.3. An extraction procedure must be performed prior to analysis of solid samples.

3. Detection Limits

3.1. Method Detection Limits (MDL) shall be performed every six months. An MDL study is to be conducted for both aqueous samples and aqueous extracts of solid samples. To view the current MDLs, see the LIMS. See **Appendix A** for a summary of the current PQLs.

3.2. MDLs must also be completed as part of a new analysts Initial Demonstration of Capability (IDOC).

4. Definitions

4.1. De-ionized (DI) Water – De-ionized water/reagent free water. Water that is passed through a purification system and is free from interferences.

4.2. Initial Calibration Verification (ICV) – Second source standard used to validate a new calibration immediately after calibration.

4.3. Initial Calibration Blank (ICB) – Analysis of a blank solution (DI water) after the initial calibration and ICV to check that the blank solution is free of target analytes.

4.4. Continuing Calibration Verification (CCV) – Standard from the same source as the calibration curve. Used prior to sample analysis on days when calibrations are not performed and periodically, there after, to check instrument stability.

4.5. Continuing Calibration Blank (CCB) – Analysis of a blank solution (DI water). Used as a periodic check that instrument is free of target analytes.

4.6. Laboratory Control Sample (LCS) - An aliquot of DI water to which known amounts of method analytes are added prior to sample preparation and analysis. The LCS is treated like a sample and used to determine whether the methodology is in control, and whether the laboratory is capable of making accurate and precise measurements.

4.7. Matrix Spike (MS) - An aliquot of field sample to which known amounts of method analytes are added prior to sample preparation and analysis. The MS is treated like a sample and used to determine whether sample matrix contributes bias to the analytical results. The background concentration in the un-spiked sample matrix must be determined separately. The MS concentrations are corrected for the background concentrations.

4.8. Matrix Spike Duplicate (MSD) – Treated exactly the same as the MS sample. Also used to examine the precision of the analytical process.

4.9. Method Blank (MB) - An aliquot of DI water or other blank matrices that are treated exactly the same as a sample including exposure to all glassware, equipment, solvents and reagents that are used in the preparation and analysis of field samples. The MB is to determine if any target analytes are present in the laboratory environment, the reagents, or the apparatus.

4.10. Method Detection Limit (MDL) – The constituent concentration that when processed through the complete method, produces a signal with a 99% probability that it is different than the blank.

4.11. Practical Quantitation Limit (PQL)/Reporting Limit – The minimum limit to which an analyte can be routinely reported.

4.12. Linear Calibration Range (LCR) – The linearity of the calibration curves (or LCR) for all analytes must be demonstrated during initial set-up of the instrument, when a calibration model is changed, or when a significant change in instrument response has occurred. Linearity must also be verified at least every six months.

5. Interferences

5.1. Interference can be caused by substances with retention times that are similar to and overlap those of the anion of interest. Large amounts of an anion can also interfere with the peak resolution of an adjacent anion. Sample dilution can be used to resolve most interference problems associated with retention times and elevated sample concentrations.

5.2. Method interference may be caused by contaminants in the DI water, sample processing chemicals or analysis apparatus.

5.3. Samples that contain large particles, large amounts of sediment or are abnormally discolored should be diluted prior to being analyzed to prevent damage to the instrument columns and flow systems.

5.4 Any anion that is not retained by the column or only slightly retained will elute quickly in the analysis and may interfere with early eluting peaks. Co-elution is caused by carbonates or other small organic anions and most significantly affect lower target analyte concentration.

6. Safety

6.1. The toxicity or carcinogenicity of each reagent or standard has not been fully defined. For handling procedures and exposure information always refer to the MSDS. Exposure to reagents and standards must be kept to a minimum. Gloves, lab coats, and safety glasses must be worn when handling samples, standards, or reagents. Refer to current HEAL Health and Safety SOP for more details.

7. Apparatus and Materials

7.1. Ion chromatograph Systems-

7.1.1. Dionex ICS-1000 IC systems

7.1.1.1. IC systems with conductivity detection.

7.1.1.2. AS-40 autosampler.

7.1.1.3. Dionex IonPac AG14A 4 x 50mm guard column.

7.1.1.4. Dionex IonPac AS14A 4 x 250mm analytical column.

7.1.1.5. Dionex Anion Self-Regenerating Suppressor, ASRS-ultra, 4mm.

7.1.1.6. Four liter eluent bottle with step-down regulator.

7.1.1.7. Dionex 5mL Auto sampler vials and filter caps. (pt. no. 38141).

7.1.2. Dionex ICS-1500 IC System

7.1.2.1. Dionex AS-50 autosampler, configured for use with 2mL vials.

7.1.2.2. 2mL polypropylene 2mL autosampler vials with split-top silicon/Teflon septum.

7.1.2.3. Dionex IonPac AS22-Fast 4 x 150mm analytical column.

7.1.2.4. Dionex IonPack AG22-Fast guard column.

7.1.2.5. Dionex Anion Self-Regenerating Suppressor, ASRS-ultra, 4mm.

7.1.2.6. Four liter eluent bottle with step-down regulator.

7.2. Operating Conditions/Integration Settings

7.2.1. Operating conditions are detailed in **Appendix B**.

7.2.2. Integration settings are detailed in **Appendix D**.

7.3. Analytical balance capable of weighing to the nearest 0.0001g.

7.4. Top loading balance capable of weighing to the nearest 0.01g.

7.5. Handheld repipettors and tips, 100uL, 1000uL, 5mL, and 10mL.

7.6. Volumetric flasks of assorted volumes.

7.7. Volumetric pipettes of assorted volumes.

7.8. Glass and plastic syringes of assorted volumes.

7.9. Auto-pipettor – 50ml.

7.10. Horizontal shaker.

7.11. Centrifuge tubes, skirted bottoms – 50mL

7.12. Centrifuge.

8. Reagents and Standards

8.1. DI water.

8.2. Sodium Bicarbonate, ACS Grade 'Baker Instra-Analyzed' – JT Baker (pt. no. 3506-01) or equivalent.

8.3. Sodium Carbonate, Anhydrous, Powder, Flux Grade 'Baker Instra-Analyzed' – JT Baker (pt. no. 4502-01) or equivalent.

8.4. Mobile Phase (Eluent) Preparation

8.4.1. Mobile Phase Preparation for ICS-1000 using AS/AG-14A analytical/guard columns

8.4.1.1. Carbonate/Bicarbonate stocks

8.4.1.1.1. 1600 mM Sodium Carbonate (Na_2CO_3) – Dissolve approximately 42.3941g sodium carbonate salt into 250 mL DI water. Transfer to a clean 250 mL plastic bottle and store at room temperature when not in use. Remake every 6 months.

8.4.1.1.2. 400 mM Sodium Bicarbonate ($NaHCO_3$) – Dissolve approximately 16.7808g of sodium bicarbonate salt into 500 mL DI. Transfer to a clean 500 mL plastic bottle and store at room temperature when not in use. Remake every 6 months.

8.4.1.2. Preparation of Mobile Phase (Eluent)

8.4.1.2.1. Close eluent regulator valve to release pressure on the eluent bottle.

8.4.1.2.2. Remove 4L eluent bottle from the instrument.

8.4.1.2.3. Pour out remaining eluent. Dilute down the drain.

8.4.1.2.4. Using 20mL volumetric pipette add 20mL of 1600 mM Na_2CO_3 stock solution to a 1L volumetric flask. To the same 1L flask, using a 10mL volumetric pipette add 10mL of 400 mM $NaHCO_3$ stock solution. Bring flask to 1L final volume.

8.4.1.2.5. Transfer the contents of the 1L flask to the 4L eluent bottle.

8.4.1.2.6. Using the same 1L volumetric flask, fill to the 1L mark with DI and transfer to the eluent bottle. Repeat with two more successive 1L additions. Total volume of eluent should be 4L.

8.4.1.2.7. Cap eluent bottle, shake well, and reconnect to instrument.

8.4.1.2.8. Label the eluent bottle with the date of preparation and the preparers initials.

8.4.1.2.9. Using the regulator, adjust the bottle pressure to 7psi.

8.4.1.2.10. On the instrument control screen, click the pump settings button and drag eluent bottle arrow to 4L.

8.4.1.2.11. Every time the eluent is changed the instrument must be primed. Open the pump head valve by turning it clockwise. Insert the 10ml syringe and click the ON button on the pump settings screen. Remove approximately 35ml of eluent, click the OFF button and close the valve. Open the secondary pump valve, click the PRIME button to prime the pump until there are no bubbles exiting the waste line and close the valve.

8.4.2. Mobile Phase Preparation for ICS-1500 using AS/AG22-Fast Analytical/Guard Columns

8.4.2.1. Carbonate/Bicarbonate stock

8.4.2.1.1. 450 mM Sodium Carbonate (Na_2CO_3)/140 mM Sodium Bicarbonate ($NaHCO_3$)– Dissolve approximately 23.8522g sodium carbonate salt and 5.8825g of sodium bicarbonate salt into 500mL DI water. Transfer to a clean 500 mL plastic bottle and store at room temperature when not in use. Remake every 6 months.

8.4.2.2. Preparation of Mobile Phase (Eluent)

8.4.2.2.1. Prepare the mobile following the same steps as the ICS-1000s using the AG/AS-14A column set with the following exception.

8.4.2.2.2. Close eluent regulator valve to release pressure on the eluent bottle.

8.4.2.2.3. Remove 4L eluent bottle from the instrument.

8.4.2.2.4. Pour out remaining eluent. Dilute down the drain.

8.4.2.2.5. Prepare the eluent using a 10mL volumetric pipette add 10mL of 450 mM Na_2CO_3/140 mM $NaHCO_3$ stock solution to a 1L volumetric flask. Bring flask to 1L final volume.

8.4.2.2.6. Transfer the contents of the 1L flask to the 4L eluent bottle.

8.4.2.2.7. Using the same 1L volumetric flask, fill to the 1L mark with DI and transfer to the eluent bottle. Repeat with two more successive 1L additions. Total volume of eluent should be 4L.

8.4.2.2.8. Cap eluent bottle, shake well, and reconnect to instrument.

8.4.2.2.9. Label the eluent bottle with the date of preparation and the preparers initials.

8.4.2.2.10. Using the regulator, adjust the bottle pressure to 7psi.

8.4.2.2.11. On the instrument control screen, click the pump settings button and drag eluent bottle arrow to 4L.

8.4.2.2.12. Every time the eluent is changed the instrument must be primed. Open the pump head valve by turning it clockwise. Insert the 10ml syringe and click the ON button on the pump settings screen. Remove approximately 35ml of eluent, click the OFF button and close the valve. Open the secondary pump valve, click the PRIME button to prime the pump until there are no bubbles exiting the waste line and close the valve.

8.4.2.3. The reagents IDs and volumes used to make the eluent are documented in the IC maintenance logbooks under the date the solution is made. A single letter instrument identifier is added after the date. This unique date-instrument ID is then documented on the bottle and all associated IC Analysis Summary Sheets (see Appendix H), so that the solution is traceable back to its origin. In addition, an expiration date of one month is assigned and documented in both the logbook and on the bottle.

8.5. Stock Standard Solutions

8.5.1. Calibration standards are purchased as both individual anions and custom mixed standards. Each standard is NIST traceable. See **Appendix C** for details. Calibration standards include:

8.5.1.1 Custom Mixed Stock Standard containing F (40mg/L), Cl (200mg/L), Br (200mg/L), Nitrate as N (120mg/L), Phosphate (200mg/L), and Sulfate (500mg/L).

8.5.1.2 Custom Stock Standard – Low Level containing F (10mg/L), Cl (50mg/L), Bromide (10mg/L), Nitrate (10mg/L), Phosphate (50mg/L), Sulfate (50mg/L).

8.5.1.3 Nitrite as N 1000mg/L standard.

8.5.2. Second source standards are purchased from a separate vendor as that of the primary calibration standards. All second source standards are purchased as individual anions. Each standard is NIST traceable at 1000mg/L. See **Appendix C** for details.

8.6. Calibration Standards

8.6.1. Calibration standards are made fresh on the day of calibration.

8.6.2. Both stock and working standards must be stored at 0-6°C when they are not in use. They may be left out on the counter during the day while the instrument is being loaded but should be stored in the refrigerator overnight.

8.6.3. All 1000mg/L stock standards and custom stock standards **must** first be brought to room temperature before preparation of the working standards.

8.6.4. Working standards can be used directly from the refrigerator. They do not need to be at room temperature.

8.6.5. All calibration standards are prepared in DI water using volumetric pipettes, volumetric flasks, and hand-held repipettors. A summary of the calibration procedure is outlined in **Appendix C**.

8.6.6. A minimum of 5 calibration standards must be analyzed for each calibration.

8.7. Second Source Standards

8.7.1. An ICV standard is made on a routine basis; every 3 months, when it shows signs of degradation, or if a comparison with a check standard indicates a problem.

8.7.2. All 1000mg/L second source stock standards **must** first be brought to room temperature before preparation of the working ICV standard.

8.7.3. See **Appendix C** for a preparation summary of ICV.

8.8. Spike Solution

8.8.1. Use the A10 (mixed anion stock) 2nd source standard as the spiking solution.

8.8.2. Aqueous Samples

8.8.2.1. For LCS spike 250uL of spike solution to 4.75mL DI for a final volume of 5mL. Cap and invert to mix.

8.8.2.2. For MS/MSD spike 250uL of spike solution to 4.75mL sample for a final volume of 5mL. Cap and invert to mix.

8.8.3. Soil Samples

8.8.3.1. For LCS spike 1.5mL of spike solution into the centrifuge tube. Add 28.5mL DI on top of the spike solution (prepare 28.5mL DI water by filling a clean centrifuge tube with 30mL DI water, remove 1.5mL with the repipettor, and pour the 28.5mL of DI on top of the LCS). Final volume will be 30mL.

8.8.3.2. For MS/MSD samples, add 10g of soil to centrifuge tube, spike 1.5mL of spike solution on the dry soil, then add 28.5mL of DI on top of sample and spike (prepare 28.5mL DI water by filling a clean centrifuge tube with 30mL DI water, remove 1.5mL with the repipettor, and pour the 28.5mL of DI on top of the MS sample. Repeat with MSD sample). Final volume will be 30mL.

9. Sample Collection, Preservation, Shipment, and Storage

9.1. Aqueous Samples

9.1.1. All aqueous samples must be collected in clean plastic or glass sampling containers.

9.1.2. After login all aqueous samples must be stored at above freezing to 6°C until analysis.

9.1.3. Aqueous anions fluoride, chloride, bromide, and sulfate must be reported from the unpreserved container. Holding time for these anions is 28 days.

9.1.4. Aqueous nitrite (NO_2), nitrate (NO_3) and phosphate (PO_4) have a hold time of 48 hours. They are reported as individual species from the unpreserved container.

9.1.4.1. Sample preserved with sulfuric acid extend the holding time to 28days, NO_2 and NO_3 are reported as a combined result (i.e. NO_2/NO_3). This is just the sum of the nitrite and nitrate values from the chromatogram. All sulfuric acid preserved samples must be diluted by at least 5x before analysis to minimize the amount of SO_4 being loaded on the column.

9.1.4.2. The holding time for phosphate cannot be extended with the use of a preservative.

9.2. Soil Samples

9.2.1. All soil samples must be collected in glass soils jars or plastic bags/containers. Nitrite, nitrate, phosphate, and sulfate samples must be stored at above freezing to 6°C until analysis. Fluoride, Chloride, and Bromide samples can be stored at room temperature.

9.2.2. Soil samples do not have an established holding time. Current HEAL protocol recommends the soil be prepared within 28days.

9.2.3. Aqueous extract of the soil has a 28day holding time for fluoride, chloride, bromide, and sulfate; and a **48hr holding time** for nitrite, nitrate, and phosphate.

10. Quality Control

10.1. Calibration

10.1.1. Instrument calibration is performed on an as needed basis.

10.1.2. A seven point curve is analyzed on a routine basis.

10.1.3. An ICV and ICB are analyzed for all anions.

10.1.4. The calibration is based upon the analyte area versus the analyte concentration. The calibration models used are linear average response, linear with 1/X weighting, and linear with $1/X^2$ weighting. Refer to current HEAL Quality Assurance plan for calibration details and formulas.

10.1.5. The calibration models are evaluated upon each calibration to determine the best fit on the instrument, under the current conditions. Weighting may change from one calibration to the next to improve the linear response of the calibration standards; however, it can not be updated in between calibrations and is not a tool to improve ICV recovery.

10.1.6. Calibration levels are summarized in **Appendix C**.

10.2. Continuing Calibration

10.2.1. A CCV and CCB are run at the beginning of each analytical batch (if not calibrating), every 10 sample injections and at the end of each run. The concentration of the CCV must be varied throughout the course of each analytical batch.

10.3. Batch QC Requirements

10.3.1. For aqueous and soil samples a MB and LCS must be analyzed every 20 new client samples. A MS and MSD must be analyzed every 10 new client samples (Batch QC is not required for dilutions of previously analyzed samples).

11. Sample Preparation and Analysis

11.1. Aqueous Sample Preparation

11.1.1. Using 5mL Dionex auto sampler vials fill to the 5ml mark on the vial.

11.1.2. Using 2mL Autosampler Vials, fill to at least the 1mL mark on vial.

11.1.3. Dilute samples into DI water.

11.2. Extraction of soils

11.2.1. Use 50mL centrifuge tubes for sample preparation.

11.2.2. Set up Prep in LIMS.

11.2.3. An MB and LCS must be prepared for every 20 samples.

11.2.4. An MS and MSD must be prepared for every 10 samples.

11.2.5. Label centrifuge tubes with appropriate HEAL number.

11.2.6. Using tabletop balance, weigh 10.00g (+/- 0.05g) of sample into each centrifuge tube.

11.2.7. To the MB and LCS do not add anything. To the MS and MSD add 10.00g of the sample.

11.2.8. Spike the appropriate amount of spike into the LCS, MS, and MSD.

11.2.9. Prepare three centrifuge tubes with 28.5mL DI.

11.2.10. Pour 28.5mL DI water on top of the LCS, MS, and MSD samples.

11.2.11. Using the auto-pipettor add 30mL DI water to the remaining centrifuge tubes. Be sure to add 30mL DI to the MB tube.

11.2.12. Agitate the samples on a horizontal shaker for 15 minutes. Remove from shaker and centrifuge for 15 minutes at approximately1500rpm.

11.2.13. For analysis, pipette or pour the aqueous extract portion into a 5mL-sample vial. Dilute as needed into DI depending on the amount of soil and debris suspended in solution.

11.3. Analysis

11.3.1. Open Chromeleon and press the "Start-Up" button on the instrument control screen. This begins the IC pumping. Allow approximately 10-20 minutes for the conductivity to stabilize.

11.3.2. Every analytical sequence must include the IC Analysis Summary sheet. While the instrument is stabilizing, begin to compile the necessary information as outlined on the IC Summary. See Appendix H.

11.3.2.1. Be sure to include all relevant calibration information from the associated curve, eluent IDs, pipette calibration checks, and dispenser IDs.

11.3.3. Prepare standards and QC samples in the 5mL Dionex or 2mL auto sampler vials.

11.3.4. Select previous day's sequence from the list of sequences on the left of the screen, or press both Control and Tab keys together to switch to the sequence screen.

11.3.5. In the File menu click "Save as" to save the sequence file as a new name. (Label as the date, i.e.: 06-26-06). Make sure "save raw data" box is checked to carry the calibration data over to the new sequence.

11.3.6. Delete all the sample files except the calibration files at the top of the sequence.

11.3.7. Before preparing the samples for analysis, key the standards and samples to be analyzed into Chromeleon. This will serve as an instrument work list. Prepare the samples from this list.

11.3.8. Add the samples in the sequence. Starting with a conditioning blank, CCV, CCB, MB, and LCS. Save sequence by clicking the disk icon in the upper left corner of the screen. Samples with short hold times may be analyzed before the MB and LCS.

11.3.9. For the AS-50 autosampler the location of the sample, QC, etc. must be specified in the analytical sequence.

11.3.10. For the AS-40 autosampler, place the conditioning blank, CCV, CCB, MB, and LCS in the auto sampler rack. Holding the rack with the open side toward you add the samples in the rack from left to right.

11.3.11. Place in the AS-40 auto sampler with the belt cogs (just flip the rack around) facing you. Once the rack is seated properly, the belt and the rack are lined up, the spring loaded rack holder will be securely pushing on the rack.

11.3.12. Press Hold/Run button on the front panel of the AS-40 auto sampler. The first sample should move into injection position.

11.3.13. Check the conductivity on the instrument control window to make sure it has stabilized. Record conductivity and pressure from the instrument panel screen into the maintenance logbook.

11.3.14. At the top of the screen click "Batch" and "Select Sequence" to get to the sequence selection box.

11.3.15. Next click the "Add" button on the right, and select the sequence(s) from the correct instrument to be analyze. Add the Shutdown method at the end of the list.

11.3.16. Click the "Start" button on the bottom of the sequence selection box to trigger the auto sampler to begin with the first injection.

11.3.17. It is possible to continue to add samples once the analysis has begun. Save any changes that are made to the analytical sequence. Be sure to always key the sample ID, dilutions, CCV, etc. into Chromeleon before preparing the samples.

11.4. Data Import into LIMS

11.4.1. See **Appendix E** for a detailed outline of the data importation from Chromeleon to LIMS.

12. Calculations

12.1. Calculations for Aqueous samples:

$$Sample\ Amount,\ mg/L = A \times D$$

Where:
A = result from chromatogram, mg/L
D = dilution factor

12.2. Calculations for Non-Aqueous samples:

$$Sample\ Amount,\ mg/kg = A \times \frac{F}{I} \times \frac{1000g}{1kg} \times \frac{1L}{1000mL} \times D$$

Where:
A= result from chromatogram, mg/L
F = final volume of extracted sample, mL
I = initial volume (mass) of sample, g
D = dilution factor

12.3. All other calculations, including those for calibration can be found in the QAP.

13. Method Performance/Estimation of Uncertainty/DOCs/LCR

13.1. Method Performance

13.1.1. PT or "blind study" samples are routinely analyzed to check the accuracy of the anions analysis.

13.2. Estimation of Measurement Uncertainty

13.2.1. Estimation of uncertainty is to be conducted for all anions in each matrix on an annual basis using LCS results. Refer to current HEAL Quality Assurance plan for measurement uncertainty details.

13.3. Demonstration of Capability DOC

13.3.1. IDOCs are to be analyzed by all new analysts before handling client samples. IDOCs must be prepared from the second source stock solution. The IDOC for 300.0 must also include an MDL study. Before a new analyst can begin reporting client samples, the IDOC as outlined in the HEAL QAP and the MDL study must be completed.

13.3.2. ADOCPs (Annual Documentation of Continued Proficiency) are to be analyzed every year by all analysts working on the instrument. If four sequential LCSs are to be used they must be prepared from a second source. See HEAL QAP for more information.

13.4. Linear Calibration Range (LCR)

13.4.1. The linearity of the calibration curves (or LCR) for all analytes must be demonstrated during initial set-up of the instrument, when a calibration model is changed, or when a significant change in instrument response has occurred. Linearity must also be verified at least every six months. See **Appendix F** for LCR determination procedure.

13.4.2. For the LCR to be acceptable the calibration model employed must be linear, the ***r*** value (correlation coefficient) must be acceptable, and the recovery of each calibration point, when assessed against the curve, must be within +/-10% of the true value of that point. Once the initial LCR is established, the bi-annual LCR verification must meet the requirements outlined in the method.

14. Pollution Prevention

14.1. Small sample volumes are used (1-5mL).

14.2. Mobile phase (eluent) is sodium carbonate and sodium bicarbonate solution. It is made in the lab from bulk materials.

14.3. Sulfuric acid preserved bottles are usually sampled in 125mL plastics or in sulfuric preserved plastics that require additional tests.

14.4. Self-regenerating auto suppression does not require external (chemical) suppression of conductivity. Thus eliminating disposal of chemicals.

15. Data Assessment and Acceptance Criteria for Quality Control Measures

15.1. See **Appendix F** for a summary of the calibration, continuing calibration, and batch QC requirements.

15.2. Examination of Chromatograms

15.2.1. Identification of unknown anions in samples is based upon their retention times relative to a known standard. Peak shape and levels of interfering anions also influence correct identification.

15.2.2. Retention times, detection parameters, and integration information is detailed in **Appendix D.**

15.2.3. Retention Times

15.2.3.1. The retention times of each anion have been previously determined by comparison to a single anion standard.

15.2.3.2. The retention time windows for each anion are calculated by taking +/- a percentage of the absolute retention time of a calibration standard. For example if the absolute retention time of Chloride is 5.000 minutes the relative retention time value is 10%, the window has the range of 5.000 minutes +/- 10%. Or 4min 30sec to 5min 30sec.

15.2.3.3. Even though the instrument automatically identifies the anions of interest, it is still important to examine closely eluting unidentified peaks. This is done by overlaying the chromatogram of the unknown with that of the nearest CCV standard.

15.2.3.4. Slight variations of analyte retention time occur with mobile phase concentration differences, backpressure on the system, ambient column temperature, and high levels of anions in samples. Be sure to include these slight variations in the overall assessment of analyte identification.

15.3. Acceptance Criteria for Quality Control Measures

15.3.1. Initial Calibration

15.3.1.1. A multiple point linear average response, weighted 1/X, or weighted $1/X^2$ calibration is used. Calibration must have a correlation coefficient percent $\geq$ 99.5.

15.3.1.2. ICV must fall between 90-110% of the assigned value.

15.3.1.3. ICB must not contain analytes above the PQL.

15.3.2. Continuing Calibration

15.3.2.1. At the beginning of every new batch if not calibrating.

15.3.2.1.1. CCV recovery must fall between 90-110% of assigned value.

15.3.2.1.2. CCB must not contain analytes above the PQL

15.3.2.2. CCV recoveries must fall between 90-110% of assigned value.

15.3.2.3. CCB must not contain analytes above the PQL.

15.3.3. Batch QC

15.3.3.1. MB must not contain analytes above the PQL.

15.3.3.2. LCS spike concentrations should fall between the high and low standards of the calibration curve. LCS recoveries should fall between 90-110% of assigned value to be considered valid.

15.3.3.3. MS/MSD spike concentrations are the same as the LCS. Recoveries must fall between established control limits. See LIMS for current limits. Limits are updated at least annually. See current HEAL QAP for more information and calculations.

15.3.3.4. The RPD for the MSD must be less than 20%.

15.3.4. Peak Shape

15.3.4.1. Occasionally baseline anomalies are identified as analytes of interest. Analyte peaks in the sample should have the same shape as those in the standard. A wide peak may be identified as an anion of interest by the Chromeleon software, reanalyze for confirmation, if necessary, at dilution.

15.3.5. Levels of Interfering Anions

15.3.5.1. High levels of interfering anions (chloride and sulfate) can effect the position of the other anions on the chromatogram or completely mask/overlap a closely eluting anion peak of interest.

15.3.5.2. Common problems occur with high levels of chloride interfering with nitrite and high levels of sulfate interfering with phosphate. To accurately report a result for nitrite or phosphate when high levels of interferents are present, overlay the sample with a chromatogram of the nearest CCV standard. If the analyte is present and the peak can be integrated, then no dilution is necessary. Otherwise, dilute the sample to minimize the effect of the interferent and to allow accurate quantitation of the anion of interest.

15.3.5.3. Manual integrations are generally performed on nitrite (when high levels of chloride are present). The baseline must be drawn from valley-to-valley and reported. In the event that the analyst must perform a manual integration, they are to consult HEALs Manual Integration SOP, and utilize the experience and expertise of technical directors and senior analysts in order to properly and effectively execute the manual integration. A "before" and "after" printout of all manual integrations must be initialed and dated by the analyst who performed the integration, and included in the data packet.

15.3.5.4. Anions of interest that do not elute closely to the interfering peak may still be affected by that interferent. If a shift in retention time occurs, overlay the sample with a CCV standard to check for retention times shifts. A dilution may be necessary to minimize interference in the sample.

15.4. Data Assessment

15.4.1. Any analyte that quantitates above the highest calibration standard for a particular anion must be diluted. All dilutions are in DI water.

15.4.2. Occasionally, it is necessary to manually adjust the integration baseline to accurately quantitate a particular anion.

15.4.2.1. For example, a nitrite peak that elutes at the tail end of a large chloride peak. In this case, it is necessary to adjust the integration baseline from the beginning of the nitrite peak to the end of the nitrite peak, not including the area from the tail end of the chloride peak.

15.4.2.2. As a rule <u>never</u> adjust peak integration. Always default to how the instrument integrates a peak. Only adjust if absolutely necessary. Any chromatogram that is modified by the analyst must be stamped with the "Adjust baseline . . ." stamp and initialed by the IC analyst. See 16.2.5.3 above, for further information.

15.4.2.3. All analytes for MS/MSD samples should be reported, at all times, unless the following issues are encountered:

15.4.2.3.1. Any analytes in the MS/MSD exceed the upper calibration range.

15.4.2.3.2. Analyte concentration in a sample exceeds 4x the spike concentration.

15.4.2.3.3. Matrix or analyte concentration inhibits the response of any analyte.

15.4.2.3.4. When all analytes fail to recovered established control limits, rerun the MS/MSD samples.

15.4.2.3.5. When analyte recovery in the MS/MSD samples fail, and batch and QC have shown the analyte is in control, the analyte is reported on the MS/MSD to demonstrate the effect on matrix on the spike recovery.

15.4.2.3.6. NO_3, NO_2 and PO_4 are reported on the MS/MSD samples when they are analyzed when 48 hours of sample collection.

16. Corrective Actions for Out-of-Control Data

16.1. Initial Calibration

16.1.1. If the correlation coefficient percent is unacceptable, the failing analyte(s) must be recalibrated. If correlation coefficient continues to fail, remake standards from stocks and recalibrate. Perform instrument maintenance if necessary.

16.1.2. If the ICV sample should fail, reanalyze the standard one additional time. If ICV sample continues to fail remake ICV standard from stocks and reanalyze. Analytes that are failing will not be reported for any samples using that calibration. If it continues to fail recalibrate instrument.

16.1.3. If ICB fails, reanalyze one additional time. If ICB continues to fail, failing anions will not be reported for any samples using that calibration. Recalibrate instrument and reanalyze ICB.

16.2. Continuing Calibration

16.2.1. Failing anion(s) on the opening CCV or CCB may not be reported until the next acceptable CCV or CCB.

16.2.2. Samples prior to and following a failing CCV or CCB must be reanalyzed with acceptable CCV or CCB.

16.2.3. All of the samples before an unacceptable closing CCV or CCB must be reanalyzed with acceptable CCV or CCB.

16.2.4. When a CCB has hit above the PQL or the CCV recovery greater than 110%, all samples associated with the failures that are non-detect for analytes of interest, can be reported.

16.3. Batch QC

16.3.1. If the MB contains anions above the PQL contamination may be present, reanalyze MB one additional time. If contamination is still present, all samples associated with that MB must be reanalyzed (or re-extracted if associated with a soil extraction batch).

16.3.2. If the LCS is unacceptable, reanalyze one time. If the LCS still fails, all samples associated with that LCS must be reanalyzed (or re-extracted if associated with a soil extraction batch.)

16.3.3. If the MS or MSD recoveries are unacceptable, evaluate using the 4x rule.

16.3.3.1. The 4x rule states that if the sample concentration is greater than 4 times the spike concentration then it is not necessary to recover the spike.

16.3.3.2. If the 4x rule does not apply, examine the recoveries of the other anions (even if those anions are not being reported). If other anion recoveries are acceptable, an un-identified interference is likely.

16.3.3.3. If the recovery of any analyte falls outside the designated control limits and the laboratory performance for that analyte is shown to be in control, the recovery problem encountered with the MS/MSD is judged to be either matrix or solution related, not system related; therefore, no corrective action is necessary beyond qualifying the data for the failure.

16.3.3.4. Interferences associated with the matrix most commonly occur with the more reactive anions (NO_2, NO_3, and PO_4). If analyst error is suspected re-extract the sample, the MS, and the MSD. Otherwise flag the unacceptable recoveries and report the data.

16.3.3.5. Do not report MS/MSD results for analytes outside the range of reportability as defined by the curve.

16.3.4. If the RPD for the MSD is > 20%. Possible matrix inconsistency is suspected. A corrective action form may need to be filled out and results qualified and reported.

17. Contingencies for Handling Out-of-Control Data

17.1. All out-of-control data is flagged, a corrective action report is generated, and results reported to the client.

17.2. To notify end user of any failure not defined on the QA/QC Report. Including: analyst errors , deviations from the SOP, instrument failures, or in response to any internal or external audit findings.

18. Waste Management

18.1. All standards waste generated by the ICs will be disposed of according to the HEAL Chemical Hygiene Plan.

19. References

19.1. USEPA Method 300.0 Determination of Inorganic Anions by Ion Chromatography, US EPA Revision 2.1, (1993).

19.2. Chemical Hygiene Plan, HEAL.

19.3. Health and Safety SOP, HEAL.

19.4. Quality Assurance Plan, HEAL.

19.5. Manual Integration SOP, HEAL.

Appendix A

Method 300.0 - PQLs

	Aqueous	Soil
Anion	PQL (mg/L)	PQL (mg/kg)
Fluoride	0.1	0.3
Chloride	0.5	1.5
Nitrite as N	0.1	0.3
Bromide	0.1	0.3
Nitrate as N	0.1	0.3
Phosphate as P	0.5	1.5
Sulfate	0.5	1.5
Nitrite/Nitrate	0.2	

Appendix B

Dionex ICS-1000 Operating Conditions

ICS-1000 w/ AS-40 autosampler using AG/AS-14A Guard and Analytical Columns

Instrument Control File

```
        On
        Pressure.LowerLimit =                    200
        Pressure.UpperLimit =                    5000
        %A.Equate =                              "%A"
        LoadPosition
        Data_Collection_Rate =                   5
        CellTemperature =                        35
        ColumnTemperature =                      30
        Suppressor_Type =                        ASRS_4mm
        ; Pump_ECD.Carbonate =                  8
        ; Pump_ECD.Bicarbonate =                1
        ; Pump_ECD.Hydroxide =                  0
        ; Pump_ECD.Tetraborate =                0
        ; Pump_ECD.Other eluent =               0
        ; Pump_ECD.Recommended Current =        43
        Suppressor_Current =                     43
        ECD_Total.Step =                         0.2
        ECD_Total.Average =                      Off
        Channel_Pressure.Step =                  0.2
        Channel_Pressure.Average =               Off
        Flow =                                   1
        Pump_ECD_Relay_1.Open
 -2.300 Pump_ECD_Relay_1.Closed

 -0.000 Pump_ECD_Relay_1.Open

  0.000 Autozero
        ECD_1.AcqOn
        ECD_Total.AcqOn
        Channel_Pressure.AcqOn
        Pump_InjectValve.InjectPosition
 15.000 ECD_1.AcqOff
        ECD_Total.AcqOff
        Channel_Pressure.AcqOff

        End
```

Summary of Control File

Flow rate (mL/min):	1.00
Conductivity Cell Temperature (°C)	35
Column Temperature (°C):	30
Supressor Current (mA):	43
Pressure Upper Limit (psi):	5000
Pressure Lower Limit (psi):	200

ICS-1600 w/ AS-50 autosampler using AG/AS-22 Fast Guard and Analytical Columns

Instrument Control File

```
        Sampler.AquireExclusive Access
        Pressure.LowerLimit =                    200
        Pressure.UpperLimit =                    3000
        %A.Equate =                              "%A"
        Flush                                    Volume=1000
        Wait                                     FlushState
        Needle Height =                          3 [mm]
        CutSegmentVolume =                       10 [ul]
        SyringeSpeed =                           4
        CycleTime =                              0 [min]
        WaitForTemperature =                     FALSE
        Data_Collection_Rate =                   5.0 [Hz]
        CellTemperature.Nominal                  35.0 [oC]
        ColumnTemperature.Nominal =              30.0 [oC]
        Supressor_Type =                         ASRS_4mm
        ; Pump_ECD.Carbonate =                  4.5
        ; Pump_ECD.Bicarbonate =                1.4
        ; Pump_ECD.Hydroxide =                  0
        ; Pump_ECD.Tetraborate =                0
        ; Pump_ECD.Other eluent =               0
        ; Pump_ECD.Recommended Current =        31
        Supressor_Current =                      31 [mA]
        ECD_Total.Step =                         0.20 [s]
        ECD_Total.Average =                      Off
        Channel_Pressure.Step =                  0.20 [s]
        Channel_Pressure.Average =               Off
        Flow =                                   1.2 [ml/min]
        Wait

  0.000 Autozero
        Load
        Wait                                     CycleTimeState
        Inject
        Wait                                     InjectState
        ECD_1.AcqOn
        ECD_Total.AcqOn
        Channel_Pressure.AcqOn
        Sampler.ReleaseExclusiveAccess

 10.000 ECD_1.AcqOff
        ECD_Total.AcqOff
        Channel_Pressure.AcqOff

        End
```

Summary of Control File

Flow rate (mL/min):	1.20
Conductivity Cell Temperature (°C)	35
Column Temperature (°C):	30
Supressor Current (mA):	31
Pressure Upper Limit (psi):	3000
Pressure Lower Limit (psi):	200

Appendix C

Calibration Procedure

Stock Standards

Individual Stock Standard

	Stock conc ug/ml.
Nitrite	1000

60 ug/ml. NO2. Volume in mLs

	Stock conc ug/ml.	Std Volume	Conc. of Std	Final Volume
Nitrite	1000	18.0	90.0	200

10 ug/ml. NO2 - Low Level. Volume in mLs

	Stock conc ug/ml.	Std Volume	Conc. of Std	Final Volume
Nitrite	1000	1.0	10.0	100

Custom Mixed Stock Standard

	Stock Std Conc. ug/ml.
Fluoride	40
Chloride	200
Bromide	200
Nitrate	120
Phosphate	200
Sulfate	600

Custom Stock Standard - Low Level

	Stock Std Conc. ug/ml.
Fluoride	10
Chloride	50
Bromide	10
Nitrate	10
Phosphate	50
Sulfate	50

Calibration Standards

Volumes are in mLs. All calibration standards are made as dilution in the autosampler vials or small cups except A1 and A7.

A7 - 200 ml. total volume, Custom Stock and 60 mg/L NO_2.

	Custom Mixed Std. ug/ml	V1 Stock Volume	C2 Conc. of A7	V2 Final Volume
Fluoride	40	20.0	4	200
Chloride	200	20.0	20	200
Nitrite	60	20.0	6	200
Bromide	200	20.0	20	200
Nitrate	120	20.0	12	200
Phosphate	200	20.0	20	200
Sulfate	600	20.0	60	200

A1- 200 ml. total volume. Low level custom stock and 10 mg/L NO_2.

	Low Level Custom ug/ml	V1 Stock Volume	C2 Conc. of A1	V2 Final Volume
Fluoride	10	2.00	0.10	200
Chloride	50	2.00	0.50	200
Nitrite	10	2.00	0.10	200
Bromide	10	2.00	0.10	200
Nitrate	10	2.00	0.10	200
Phosphate	50	2.00	0.50	200
Sulfate	50	2.00	0.50	200

A6 - A7 diluted 75% conc., 4.0 ml. total volume.

	C1 A7 ug/ml	V1 A7 Volume	C2 Conc. of A6	V2 Final Volume
Fluoride	4.00	3.0	3	4.0
Chloride	20.00	3.0	15	4.0
Nitrite	6.00	3.0	4.5	4.0
Bromide	20.00	3.0	15	4.0
Nitrate	12.00	3.0	9	4.0
Phosphate	20.00	3.0	15	4.0
Sulfate	60.00	3.0	37.5	4.0

A5 - A7 diluted x2, 4.0 ml. total volume.

	C1 A7 ug/ml	V1 A7 Volume	C2 Conc. of A5	V2 Final Volume
Fluoride	4.00	2.0	2	4.0
Chloride	20.00	2.0	10	4.0
Nitrite	6.00	2.0	4	4.0
Bromide	20.00	2.0	10	4.0
Nitrate	12.00	2.0	6	4.0
Phosphate	20.00	2.0	10	4.0
Sulfate	60.00	2.0	25	4.0

→

A3 - A5 diluted x8, 4.0 ml. total volume.

	C1 A5 ug/ml	V1 A5 Volume	C2 Conc. of A3	V2 Final Volume
Fluoride	2.0	0.5	0.25	4.0
Chloride	10.0	0.5	1.25	4.0
Nitrite	4.0	0.5	0.50	4.0
Bromide	10.0	0.5	1.25	4.0
Nitrate	6.0	0.5	0.75	4.0
Phosphate	10.0	0.5	1.25	4.0
Sulfate	25.0	0.5	3.125	4.0

A4 - A7 diluted x5, 5.0 ml. total volume.

	C1 A7 ug/ml	V1 A7 Volume	C2 Conc. of A4	V2 Final Volume
Fluoride	4.00	1.0	0.8	5.0
Chloride	20.00	1.0	4.0	5.0
Nitrite	6.00	1.0	1.2	5.0
Bromide	20.00	1.0	4.0	5.0
Nitrate	12.00	1.0	2.4	5.0
Phosphate	20.00	1.0	4.0	5.0
Sulfate	60.00	1.0	10.0	5.0

Appendix C

Calibration Procedure

A2 - A7 diluted x20, 5.0 mL total volume.

	C1 A7 ug/ml	V1 A7 Volume	C2 Conc. of A2	V2 Final Volume
Fluoride	4.00	0.25	0.2	5.0
Chloride	20.00	0.25	1.0	5.0
Nitrite	8.00	0.25	0.4	5.0
Bromide	20.00	0.25	1.0	5.0
Nitrate	12.00	0.25	0.6	5.0
Phosphate	20.00	0.25	1.0	5.0
Sulfate	50.00	0.25	2.5	5.0

Continuing Calibration Standards

Custom Mixed Stock Standard

	Stock conc ug/mL
Fluoride	40
Chloride	200
Bromide	200
Nitrate	120
Phosphate	200
Sulfate	500

80 ug/mL NO2

	Stock conc ug/mL
Nitrite	80.0

A4 CCV - 200 mL total volume.

	Custom Mixed Std. ug/mL	V1 CCV Std. Volume	C2 Conc. of A4 CCV	V2 Final Volume
Fluoride	40	4	0.8	200
Chloride	200	4	4.0	200
Nitrite	80	4	1.6	200
Bromide	200	4	4.0	200
Nitrate	120	4	2.4	200
Phosphat	200	4	4.0	200
Sulfate	500	4	10.0	200

A8 CCV - 200 mL total volume.

	Custom Mixed Std. ug/mL	V1 CCV Std. Volume	C2 Conc. of A8 CCV	V2 Final Volume
Fluoride	40	10	2.0	200
Chloride	200	10	10.0	200
Nitrite	80	10	4.0	200
Bromide	200	10	10.0	200
Nitrate	120	10	6.0	200
Phosphat	200	10	10.0	200
Sulfate	500	10	25.0	200

A9 CCV - 200 mL total volume.

	Custom Mixed Std. ug/mL	V1 CCV Std. Volume	C2 Conc. of A9 CCV	V2 Final Volume
Fluoride	40	15	3.0	200
Chloride	200	15	15.0	200
Nitrite	80	15	6.0	200
Bromide	200	15	15.0	200
Nitrate	120	15	9.0	200
Phosphat	200	15	15.0	200
Sulfate	500	15	37.5	200

Second Source Standards

	C1 Stock conc. ug/mL	V1 Stock Volume	C2 Conc. of 2nd std.	V2 Final Volume
Fluoride	1000	2.0	10	200
Chloride	1000	20.0	100	200
Nitrite	1000	4.0	20	200
Bromide	1000	10.0	50	200
Nitrate	1000	10.0	50	200
Phosphate	1000	20.0	100	200
Sulfate	1000	40.0	200	200

ICV	C1 2nd Std. ug/mL	V1 2nd Std. Volume	C2 Conc. of ICV	V2 Final Volume
Fluoride	10	20.0	1	200
Chloride	100	20.0	10	200
Nitrite	20	20.0	2	200
Bromide	50	20.0	5	200
Nitrate	50	20.0	5	200
Phosphate	100	20.0	10	200
Sulfate	200	20.0	20	200

LCS/MS	C1 2nd Std. ug/mL	V1 2nd Std. Volume	C2 Conc. of LCS/MS	V2 Final Volume
Fluoride	10	0.25	0.5	5.0
Chloride	100	0.25	5.0	5.0
Nitrite	20	0.25	1.0	5.0
Bromide	50	0.25	2.5	5.0
Nitrate	50	0.25	2.5	5.0
Phosphate	100	0.25	5.0	5.0
Sulfate	200	0.25	10.0	5.0

Appendix D

Retention Times, Detection Parameters, and Intergration Information

Orion

Detection Table

No.	Retention Time (min)	Parameter	Value	Channel
1	0.000	Minimum Area	0.01 "[Signal]"min"	All Channels
2	0.000	Inhibit Integration	On	All Channels
3	2.800	Inhibit Integration	Off	All Channels
4	2.800	Fronting Sensitivity Factor	2	All Channels
5	4.110	Inhibit Integration	On	All Channels
6	4.120	Inhibit Integration	Off	All Channels
7	6.500	Minimum Area	0.001"[Signal]"min"	All Channels
8	9.000	Minimum Area	0.01 "[Signal]Min"	All Channels
9	9.001	Inhibit Integration	On	All Channels
10	9.002	Inhibit Integration	Off	All Channels

Peak Table

Use Recently Detected Retention Times: Off
Peak Retention Time Determination: Absolute
Delay Time of 2nd Detector: <None>
Delay Time of 3rd Detector: <None>

No.	Peak Name	Retention time (min)	Window	Standard	Integration Type	Calibration Type	Peak Type	Amount STD1	Amount STD2	Amount STD3	Amount STD4	Amount STD5	Amount STD6	Amount STD7
1	Fluoride	3.350	5.000 RG	External	Area	XLOff	Auto	0.10	0.20	0.25	0.80	2.00	3.00	4.00
2	Chloride	4.850	5.000 RG	External	Area	XLOff	Auto	0.50	1.00	1.25	4.00	10.00	15.00	20.00
3	Nitrite	5.700	5.000 RG	External	Area	XLOff	B-M-B	0.10	0.40	0.50	1.60	4.00	6.00	8.00
4	Bromide	7.100	5.000 RG	External	Area	XLOff	B-M-B	0.10	1.00	1.25	4.00	10.00	15.00	20.00
5	Nitrate	7.950	5.000 RG	External	Area	XXLOff	B-M-B	0.10	0.60	0.75	2.40	6.00	9.00	12.00
6	Phosphate	10.850	3.000 AF	External	Area	XLOff	Auto	0.50	1.00	1.25	4.00	10.00	15.00	20.00
7	Sulfate	12.900	5.000 RG	External	Area	XXLOff	Auto	0.50	2.50	3.13	10.00	25.00	37.50	50.00

Amount Table

Dimension of Amounts: mg/L
Reference volume for amounts: Use inject volume of first standard
Number of Amount Columns: 9
Sample column used for amount column assignment: Sample Name

No.	Peak Name	Retention time (min)	Response Factor	Amount STD1	Amount STD2	Amount STD3	Amount STD4	Amount STD5	Amount STD6	Amount STD7	Match Criteria	Threshold
1	Fluoride	3.350	1.000	0.10	0.20	0.25	0.80	2.00	3.00	4.00	Least Square	950
2	Chloride	4.850	1.000	0.50	1.00	1.25	4.00	10.00	15.00	20.00	Least Square	950
3	Nitrite	5.700	1.000	0.10	0.40	0.50	1.60	4.00	6.00	8.00	Least Square	950
4	Bromide	7.100	1.000	0.10	1.00	1.25	4.00	10.00	15.00	20.00	Least Square	950
5	Nitrate	7.950	1.000	0.10	0.60	0.75	2.40	6.00	9.00	12.00	Least Square	950
6	Phosphate	10.850	1.000	0.50	1.00	1.25	4.00	10.00	15.00	20.00	Least Square	950
7	Sulfate	12.900	1.000	0.50	2.50	3.13	10.00	25.00	37.50	50.00	Least Square	950

CAL TYPES: LOff = Linear ALOff = Linear Average Response XLOff = Linear 1/X Weighting XXLOff = Linear $1/X^2$ Weighting
Calibration Types and Retention Times will change as over time due to changes in the system.

Appendix D

Retention Times, Detection Parameters, and Intergration Information

Triton

Detection Table

No.	Retention Time (min)	Parameter	Value	Channel
1	0.000	Minimum Area	0.01 "[Signal]*min"	All Channels
2	0.000	Inhibit Integration	On	All Channels
3	2.950	Fronting Sensitivity Factor	2	All Channels
4	2.950	Inhibit Integration	Off	All Channels
5	4.110	Inhibit Integration	On	All Channels
6	4.120	Inhibit Integration	Off	All Channels
7	6.500	Minimum Area	0.001"[Signal]*min"	All Channels
8	8.500	Minimum Area	0.01 "[Signal]Min"	All Channels

Peak Table

Use Recently Detected Retention Times: Off
Peak Retention Time Determination: Absolute
Delay Time of 2'nd Detector: <None>
Delay Time of 3'rd Detector: <None>

No.	Peak Name	Retention time (min)	Window	Standard	Integration Type	Calibration Type	Peak Type	Amount STD1	Amount STD2	Amount STD3	Amount STD4	Amount STD5	Amount STD6	Amount STD7
1	Fluoride	3.370	5.000 RG	External	Area	LOff	B-M-B	0.10	0.20	0.25	0.80	2.00	3.00	4.00
2	Chloride	4.780	5.000 RG	External	Area	XLOff	B-M-B	0.50	1.00	1.25	4.00	10.00	15.00	20.00
3	Nitrite	5.600	5.000 RG	External	Area	XLOff	B-M-B	0.10	0.40	0.50	1.60	4.00	6.00	8.00
4	Bromide	6.960	5.000 RG	External	Area	XLOff	B-M-B	0.10	1.00	1.25	4.00	10.00	15.00	20.00
5	Nitrate	7.850	5.000 RG	External	Area	XXLOff	B-M-B	0.10	0.60	0.75	2.40	6.00	9.00	12.00
6	Phosphate	10.420	10.000 RG	External	Area	XLOff	B-M-B	0.50	1.00	1.25	4.00	10.00	15.00	20.00
7	Sulfate	12.510	10.000 RG	External	Area	XLOff	B-M-B	0.50	2.50	3.13	10.00	25.00	37.50	50.00

Amount Table

Dimension of Amounts: mg/L
Reference volume for amounts: Use inject volume of first standard
Number of Amount Columns: 9
Sample column used for amount column assignment: Sample Name

No.	Peak Name	Retention time (min)	Response Factor	Amount STD1	Amount STD2	Amount STD3	Amount STD4	Amount STD5	Amount STD6	Amount STD7	Match Criteria	Threshold
1	Fluoride	3.370	1.000	0.10	0.20	0.25	0.80	2.00	3.00	4.00	Least Square	950
2	Chloride	4.780	1.000	0.50	1.00	1.25	4.00	10.00	15.00	20.00	Least Square	950
3	Nitrite	5.600	1.000	0.10	0.40	0.50	1.60	4.00	6.00	8.00	Least Square	950
4	Bromide	6.960	1.000	0.10	1.00	1.25	4.00	10.00	15.00	20.00	Least Square	950
5	Nitrate	7.850	1.000	0.10	0.60	0.75	2.40	6.00	9.00	12.00	Least Square	950
6	Phosphate	10.420	1.000	0.50	1.00	1.25	4.00	10.00	15.00	20.00	Least Square	950
7	Sulfate	12.510	1.000	0.50	2.50	3.13	10.00	25.00	37.50	50.00	Least Square	950

CAL TYPES: LOff = Linear ALOff = Linear Average Response XLOff = Linear 1/X Weighting XXLOff = Linear $1/X^2$ Weighting
Calibration types and Retention Times will change as over time due to changes in the system.

Appendix D

Retention Times, Detection Parameters, and Intergration Information

Gemini

Detection Table

No.	Retention Time (min)	Parameter	Value	Channel
1	0.000	Minimum Area	0.005 "[Signal]*min"	All Channels
2	0.000	Void Volume Treatment	On	All Channels
3	0.000	Tailing Sensitivity Factor	6.0	All Channels
4	0.000	Inhibit Integration	On	All Channels
5	1.750	Inhibit Integration	Off	All Channels
6	2.480	Inhibit Integration	On	All Channels
7	2.500	Inhibit Integration	Off	All Channels
8	3.000	Void Volume Treatment	Off	All Channels
9	5.000	Inhibit Integration	On	All Channels
10	5.300	Inhibit Integration	Off	All Channels

Peak Table

Use Recently Detected Retention Times: Off
Peak Retention Time Determination: Absolute
Delay Time of 2nd Detector: <None>
Delay Time of 3rd Detector: <None>

No.	Peak Name	Retention time (min)	Window	Standard	Integration Type	Calibration Type	Peak Type	Amount STD1	Amount STD2	Amount STD3	Amount STD4	Amount STD5	Amount STD6	Amount STD7
1	Fluoride	1.957	5.000 RG	External	Area	XLOff	Auto	0.10	0.20	0.25	0.80	2.00	3.00	4.00
2	Chloride	2.809	5.000 RG	External	Area	XXLOff	Auto	0.50	1.00	1.25	4.00	10.00	15.00	20.00
3	Nitrite	3.384	5.000 RG	External	Area	XXLOff	Auto	0.10	0.40	0.50	1.60	4.00	6.00	8.00
4	Bromide	4.044	5.000 RG	External	Area	XLOff	Auto	0.10	1.00	1.25	4.00	10.00	15.00	20.00
5	Nitrate	4.602	5.000 RG	External	Area	XXLOff	Auto	0.10	0.60	0.75	2.40	6.00	9.00	12.00
6	Phosphate	5.932	5.000 RG	External	Area	XXLOff	Auto	0.50	1.00	1.25	4.00	10.00	15.00	20.00
7	Sulfate	6.770	5.000 RG	External	Area	XXLOff	Auto	0.50	2.50	3.13	10.00	25.00	37.50	50.00

Amount Table

Dimension of Amounts: mg/L
Reference volume for amounts: Use inject volume of first standard
Number of Amount Columns: 7
Sample column used for amount column assignment: Sample Name

No.	Peak Name	Retention time (min)	Response Factor	Amount STD1	Amount STD2	Amount STD3	Amount STD4	Amount STD5	Amount STD6	Amount STD7	Match Criteria	Threshold
1	Fluoride	1.957	1.000	0.10	0.20	0.25	0.80	2.00	3.00	4.00	Least Square	950
2	Chloride	2.809	1.000	0.50	1.00	1.25	4.00	10.00	15.00	20.00	Least Square	950
3	Nitrite	3.384	1.000	0.10	0.40	0.50	1.60	4.00	6.00	8.00	Least Square	950
4	Bromide	4.044	1.000	0.10	1.00	1.25	4.00	10.00	15.00	20.00	Least Square	950
5	Nitrate	4.602	1.000	0.10	0.60	0.75	2.40	6.00	9.00	12.00	Least Square	950
6	Phosphate	5.932	1.000	0.50	1.00	1.25	4.00	10.00	15.00	20.00	Least Square	950
7	Sulfate	6.770	1.000	0.50	2.50	3.13	10.00	25.00	37.50	50.00	Least Square	950

CAL TYPES: LOff = Linear ALOff = Linear Average Response XLOff = Linear 1/X Weighting XXLOff = Linear $1/X^2$ Weighting
Calibration types and Retention Times will change as over time due to changes in the system.

Appendix E

Procedure for Importing Data into LIMS

A. From previous days run, highlight samples from the ICV to the closing CCB.
B. Right click in the highlighted area, and choose batch report.
C. Check the printout box (if not already selected) and the export box.
   1. This will open a new window, select the Excel file format box, click next.
   2. In the new window, uncheck all except the summary option. Click finish, click ok to print and export.
D. Click OK when all have been printed and exported.
E. In the same days run, right click, choose print sequence.
   1. Click OK with boxes unchecked.
   2. Click OK to print.
F Open Microsoft Excel program.
   1. Go to File, Open.
   2. Find file from previous days run in G:/Omega/Data/300/Orion/MySequences or G:/Omega/Data/300/Triton/MySequences depending on which instrument is being imported.
   3. Open the run and double click on the last sample to retrieve all data from the run.
G. With the previous days run open, go to File, Open.
   1. Open the macro from G:/Omega/Data/300/19Macro3.
H. Return to the spreadsheet containing exported data .
   1. Go to Tools, Macro, Macros.
   2. Choose 'This Workbook' option from the 'Macros in:' drop-down list.
   3. Click Run.
I. From the 19Macro3 spreadsheet, copy anion headings (E1-L1). Paste to the spreadsheet that contains exported data (E1-L1). Data is now ready to be saved to the G:/Omega/Data/300 folder.
   1. Select Save As from File.
   2. Enter date and the instrument that data is from as the file name.
   3. Save, and keep open.
J. In the LIMS, open data entry.
   1. Enter Orion or Triton in Intstrument ID.
   2. Enter date that data is from in the Run Start Date box
   3. Select analyst who performed the run.
   4. Select Data Import
      a. In the Specification box, select Omega Excel File.
      b. Test Code will be 300_W, Sample type will be Samp (very important, do not leave either box empty or data will not import.)
      c. Import Data as: **Raw Result**.
      d. Click run import, Select the file that contains data of interest from the G:/Omega/Data/300 folder.
      e. A data range will need to be entered in the window that opens. The range will always begin with A1. To complete the range, refer to the excel spreadsheet that contains the imported data. The range will end with the AnalDate column and the row that contains the closing CCB. (ex. A1:L32).
      f. Click OK.

Appendix F

Procedure for Determining LCR

1. Perform instrument maintenance to ensure optimal instrument performance.

2. Calibrate instrument using all curve calibration points. Remember to analyze ICV and ICB.

3. Using the Arizona Calibration Workbook, enter all of the peak areas and standard concentrations into the "Initial Calibration" section of the spreadsheet.

4. The spreadsheet then calculates the results for linear average response, linear regression, and quadratic regression.

5. Copy and paste the concentrations and areas from the "Initial Calibration" section of the work sheet to the "Sample Calculations" section of the work sheet

6. As a final check include the ICV concentration and area in the "Sample Calculations" section.

7. In the "Sample Calculations" section, the spreadsheet details the results of four linear models (average response, equal weighing, 1/X weighing, and $1/X^2$ weighing). It also includes a non-liner quadratic calibration model.

8. For the LCR to be acceptable the calibration model employed must be linear, the *r* value (correlation coefficient) must be acceptable, and the recovery of each calibration point, when assessed against the curve, must be within +/-10% of the true value of that point.

Appendix G

Inorganic Anions Analysis Using EPA 300.0
Calibration Specification And Corrective Action Procedures

Analytical Method	Parameter	QC Element	Frequency	Acceptance Criteria	Corrective Action
EPA 300.0	Wet Chemistry	Intial Calibration (7-9 points)	Periodically; prior to sample analysis	Correlation coefficient r ≥ 99.5	1. Recalibrate 2. Remake calibration standards from stocks and recalibrate 3. Perform instrument maintenance, recalibrate
		Intial Calibration Verification (ICV)	After initial calibration, prior to sample analysis	Results ±10% of assigned value	1. Reanalyze ICV, one additional time. 2. Initial Calibration: recalibrate instrument, and reanalyze. 3. Initial Calibration: Remake ICV standards, recalibrate, and reanalyze.
		Initial Calibration Blank (ICB)	After calibration, prior to sample analysis	< Practical quantitation limit (PQL)	1. Reanalyze one additional time 2. Recalibrate instrument
		Method Blank (MB)	1 per preperation/analysis batch (≤ 20 samples)	< PQL	1. Reanalyze method blank, one additional time 2. Reprep/reanalyze analytical analytical batch as appropriate
		Laboratory Control Sample (LCS)	1 per preperation/analysis batch (≤ 20 samples)	% Recovery ± 10% of assigned value	1. Reanlayze LCS, one additional time 2. Reprep/reanalyze LCS and affected samples
		Matrix Spike (MS)	1 per preperation/analysis batch (≤ 10 samples)	% Recovery must fall between established limits	1. Asses data (4 x rule) 2. Narrate all outliers
		Matrix Spike Duplicate (MSD)	1 per preperation/analysis batch (≤ 10 samples)	% Recovery must fall between established limits Relative percent difference (RPD) <20%	1. Asses data (4 x rule) 2. Narrate all outliers 3. RPD: Narrate all outliers
		Continuing Calibration Verification (CCV)	At the beginning, every 10 samples, and at the end of the analytical sequence	Results ±10% of assigned value	1. Reanalyze affected samples back to the last acceptible CCV and to the next acceptable CCV
		Continuing Calibration Blank (CCB)	At the beginning, every 10 samples, and at the end of the analytical sequence	< Practical quantitation limit (PQL)	1. Reanalyze affected samples back to the last acceptible CCB and to the next acceptable CCB

Appendix H

IC Analysis Summary

Sequence ID: _____ Run ID: _____

ORION **TRITON** **GEMINI**

Calibration

Calibration ID: _____

	no	yes	Do not report:
Any failures in curve?	☐	☐	F Cl NO_2-N Br NO_3-N PO_4-P SO_4
Any failures in ICV?	☐	☐	F Cl NO_2-N Br NO_3-N PO_4-P SO_4
Any failures in ICB?	☐	☐	F Cl NO_2-N Br NO_3-N PO_4-P SO_4

Notes:

Calibration QA: _____

Eluent

Eluent ID: _____

Pipette Calibration Check

Pipette No: ____ Spike Volume: 0.250 mL Wt DI (g): _____ Rm Temp (°C): _____ Analyst: _____

Pipette No: ____ Spike Volume: 0.500 mL Wt DI (g): _____ Rm Temp (°C): _____ Analyst: _____

Pipette No: ____ Spike Volume: 1.00 mL Wt DI (g): _____ Rm Temp (°C): _____ Analyst: _____

Pipette No: ____ Spike Volume: 5.00 mL Wt DI (g): _____ Rm Temp (°C): _____ Analyst: _____

Dispensor

Dispensor ID: _____

Hand Calculation

Hand Calculation? yes ☐

Work List Verification

Work List Verified By: _____

Notes:

Loaded By: _____ 1st Review/Date: _____ Qa'd/Date: _____

H:\Hal\Doc\eops\SOP Revisions\S300.0 Tables for Revision10.xls\Appendix H

Hall Environmental Analysis Laboratory

Common Name: EPA Method 8270C	S.O.P. Number: S-8270C-10
Method: Semi-Volatile Organic Compounds by Gas Chromatography / Mass Spectrometry (GC/MS).	
Effective Date: 26 August 2011	Supersedes: S-8270C-9
Laboratory Manager:	QA/QC Officer:

1.0 Scope and Application

1.1. Method 8270C is used to determine the concentration of semi-volatile organic compounds in aqueous and non-aqueous matrices. The following table indicates compounds that may be determined by this method.

1.2. Use of Method 8270C can be used to quantitate most neutral, acidic and basic organic compounds that are both soluble in methylene chloride, and capable of being eluted without derivatization as sharp peaks from a gas chromatographic fused-slica capillary column coated with a slightly polar silicone. Such compounds include poly-nuclear aromatic hydrocarbons, chlorinated hydrocarbons and pesticides, phthalate esters, organophosphate esters, nitrosamines, haloethers, aldehydes, ethers, ketones, anilines, pyridines, quinolines, aromatic nitro compounds, and phenols, including nitrophenols. See table 1 for the list of compounds that can be analyzed by this method.

1.3. The current PQL and MDL for 8270 analytes at Hall Environmental Analysis Lab can be found in the LIMS.

Table 1

Compounds	CAS No[a]	Appropriate Preparation Techniques[b] 3510	3520	3540/ 3541	3550	3580
Acenaphthene	83-32-9	X	X	X	X	X
Acenaphthene-d_{10} (IS)		X	X	X	X	X
Acenaphthylene	208-96-8	X	X	X	X	X
Acetophenone	98-86-2	X	ND	ND	ND	X
2-Acetylaminofluorene	53-96-3	X	ND	ND	ND	X
1-Acetyl-2-thiourea	591-08-2	LR	ND	ND	ND	LR
Aldrin	309-00-2	X	X	X	X	X
2-Aminoanthraquinone	117-79-3	X	ND	ND	ND	X
Aminoazobenzene	60-09-3	X	ND	ND	ND	X
4-Aminobiphenyl	92-67-1	X	ND	ND	ND	X
3-Amino-9-ethylcarbazole	132-32-1	X	X	ND	ND	ND
Anilazine	101-05-3	X	ND	ND	ND	X
Aniline	62-53-3	X	X	ND	X	X
o-Anisidine	90-04-0	X	ND	ND	ND	X
Anthracene	120-12-7	X	X	X	X	X
Aramite	140-57-8	HS(43)	ND	ND	ND	X
Aroclor 1016	12674-11-2	X	X	X	X	X
Aroclor 1221	11104-28-2	X	X	X	X	X
Aroclor 1232	11141-16-5	X	X	X	X	X
Aroclor 1242	53469-21-9	X	X	X	X	X
Aroclor 1248	12672-29-6	X	X	X	X	X
Aroclor 1254	11097-69-1	X	X	X	X	X
Aroclor 1260	11096-82-5	X	X	X	X	X
Azinphos-methyl	86-50-0	HS(62)	ND	ND	ND	X
Barban	101-27-9	LR	ND	ND	ND	LR
Benzidine	92-87-5	CP	CP	CP	CP	CP
Benzoic acid	65-85-0	X	X	ND	X	X
Benz(a)anthracene	56-55-3	X	X	X	X	X
Benzo(b)fluoranthene	205-99-2	X	X	X	X	X
Benzo(k)fluoranthene	207-08-9	X	X	X	X	X
Benzo(g,h,i)perylene	191-24-2	X	X	X	X	X
Benzo(a)pyrene	50-32-8	X	X	X	X	X

Compounds	CAS No[a]	Appropriate Preparation Techniques[b] 3510	3520	3540/ 3541	3550	3580
p-Benzoquinone	106-51-4	OE	ND	ND	ND	X

Benzyl alcohol	100-51-6	X	X	ND	X	X
α-BHC	319-84-6	X	X	X	X	X
β-BHC	319-85-7	X	X	X	X	X
δ-BHC	319-86-8	X	X	X	X	X
γ-BHC (Lindane)	58-89-9	X	X	X	X	X
Bis(2-chloroethoxy)methane	111-91-1	X	X	X	X	X
Bis(2-chloroethyl) ether	111-44-4	X	X	X	X	X
Bis(2-chloroisopropyl) ether	108-60-1	X	X	X	X	X
Bis(2-ethylhexyl) phthalate	117-81-7	X	X	X	X	X
4-Bromophenyl phenyl ether	101-55-3	X	X	X	X	X
Bromoxynil	1689-84-5	X	ND	ND	ND	X
Butyl benzyl phthalate	85-68-7	X	X	X	X	X
Captafol	2425-06-1	HS(55)	ND	ND	ND	X
Captan	133-06-2	HS(40)	ND	ND	ND	X
Carbaryl	63-25-2	X	ND	ND	ND	X
Carbofuran	1563-66-2	X	ND	ND	ND	X
Carbophenothion	786-19-6	X	ND	ND	ND	X
Chlordane (NOS)	57-74-9	X	X	X	X	X
Chlorfenvinphos	470-90-6	X	ND	ND	ND	X
4-Chloroaniline	106-47-8	X	ND	ND	ND	X
Chlorobenzilate	510-15-6	X	ND	ND	ND	X
5-Chloro-2-methylaniline	95-79-4	X	ND	ND	ND	X
4-Chloro-3-methylphenol	59-50-7	X	X	X	X	X
3-(Chloromethyl)pyridine hydrochloride	6959-48-4	X	ND	ND	ND	X
1-Chloronaphthalene	90-13-1	X	X	X	X	X
2-Chloronaphthalene	91-58-7	X	X	X	X	X
2-Chlorophenol	95-57-8	X	X	X	X	X
4-Chloro-1,2-phenylenediamine	95-83-0	X	X	ND	ND	ND
4-Chloro-1,3-phenylenediamine	5131-60-2	X	X	ND	ND	ND
4-Chlorophenyl phenyl ether	7005-72-3	X	X	X	X	X
Chrysene	218-01-9	X	X	X	X	X
Chrysene-d_{12} (IS)		X	X	X	X	X
Coumaphos	56-72-4	X	ND	ND	ND	X
p-Cresidine	120-71-8	X	ND	ND	ND	X
Crotoxyphos	7700-17-6	X	ND	ND	ND	X
2-Cyclohexyl-4,6-dinitro-phenol	131-89-5	X	ND	ND	ND	LR
4,4'-DDD	72-54-8	X	X	X	X	X
4,4'-DDE	72-55-9	X	X	X	X	X
4,4'-DDT	50-29-3	X	X	X	X	X
Demeton-O	298-03-3	HS(68)	ND	ND	ND	X
Demeton-S	126-75-0	X	ND	ND	ND	X
Diallate (cis or trans)	2303-16-4	X	ND	ND	ND	X

Appropriate Preparation Techniques[b]

3540/

Compounds	CAS No[a]	3510	3520	3541	3550	3580
2,4-Diaminotoluene	95-80-7	DC,0E(42)	ND	ND	ND	X
Dibenz(a,j)acridine	224-42-0	X	ND	ND	ND	X
Dibenz(a,h)anthracene	53-70-3	X	X	X	X	X
Dibenzofuran	132-64-9	X	X	ND	X	X
Dibenzo(a,e)pyrene	192-65-4	ND	ND	ND	ND	X
1,2-Dibromo-3-chloropropane	96-12-8	X	X	ND	ND	ND
Di-n-butyl phthalate	84-74-2	X	X	X	X	X
Dichlone	117-80-6	OE	ND	ND	ND	X
1,2-Dichlorobenzene	95-50-1	X	X	X	X	X
1,3-Dichlorobenzene	541-73-1	X	X	X	X	X
1,4-Dichlorobenzene	106-46-7	X	X	X	X	X
1,4-Dichlorobenzene-d_4 (IS)		X	X	X	X	X
3,3'-Dichlorobenzidine	91-94-1	X	X	X	X	X
2,4-Dichlorophenol	120-83-2	X	X	X	X	X
2,6-Dichlorophenol	87-65-0	X	ND	ND	ND	X
Dichlorovos	62-73-7	X	ND	ND	ND	X
Dicrotophos	141-66-2	X	ND	ND	ND	X
Dieldrin	60-57-1	X	X	X	X	X
Diethyl phthalate	84-66-2	X	X	X	X	X
Diethylstilbestrol	56-53-1	AW,0S(67)	ND	ND	ND	X
Diethyl sulfate	64-67-5	LR	ND	ND	ND	LR
Dihydrosaffrole	56312-13-1	ND	ND	ND	ND	ND
Dimethoate	60-51-5	HE,HS(31)	ND	ND	ND	X
3,3'-Dimethoxybenzidine	119-90-4	X	ND	ND	ND	LR
Dimethylaminoazobenzene	60-11-7	X	ND	ND	ND	X
7,12-Dimethylbenz(a)-anthracene	57-97-6	CP(45)	ND	ND	ND	CP
3,3'-Dimethylbenzidine	119-93-7	X	ND	ND	ND	X
α,α-Dimethylphenethylamine	122-09-8	ND	ND	ND	ND	X
2,4-Dimethylphenol	105-67-9	X	X	X	X	X
Dimethyl phthalate	131-11-3	X	X	X	X	X
1,2-Dinitrobenzene	528-29-0	X	ND	ND	ND	X
1,3-Dinitrobenzene	99-65-0	X	ND	ND	ND	X
1,4-Dinitrobenzene	100-25-4	HE(14)	ND	ND	ND	X
4,6-Dinitro-2-methylphenol	534-52-1	X	X	X	X	X
2,4-Dinitrophenol	51-28-5	X	X	X	X	X
2,4-Dinitrotoluene	121-14-2	X	X	X	X	X
2,6-Dinitrotoluene	606-20-2	X	X	X	X	X
Dinocap	39300-45-3	CP,HS(28)	ND	ND	ND	CP
Dinoseb	88-85-7	X	ND	ND	ND	X
Dioxathion	78-34-2	ND	ND	ND	ND	ND
Diphenylamine	122-39-4	X	X	X	X	X
5,5-Diphenylhydantoin	57-41-0	X	ND	ND	ND	X
1,2-Diphenylhydrazine	122-66-7	X	X	X	X	X

Compounds	CAS No[a]	Appropriate Preparation Techniques[b] 3510	3520	3540/ 3541	3550	3580
Di-n-octyl phthalate	117-84-0	X	X	X	X	X
Disulfoton	298-04-4	X	ND	ND	ND	X
Endosulfan I	959-98-8	X	X	X	X	X
Endosulfan II	33213-65-9	X	X	X	X	X
Endosulfan sulfate	1031-07-8	X	X	X	X	X
Endrin	72-20-8	X	X	X	X	X
Endrin aldehyde	7421-93-4	X	X	X	X	X
Endrin ketone	53494-70-5	X	X	ND	X	X
EPN	2104-64-5	X	ND	ND	ND	X
Ethion	563-12-2	X	ND	ND	ND	X
Ethyl carbamate	51-79-6	DC(28)	ND	ND	ND	X
Ethyl methanesulfonate	62-50-0	X	ND	ND	ND	X
Famphur	52-85-7	X	ND	ND	ND	X
Fensulfothion	115-90-2	X	ND	ND	ND	X
Fenthion	55-38-9	X	ND	ND	ND	X
Fluchloralin	33245-39-5	X	ND	ND	ND	X
Fluoranthene	206-44-0	X	X	X	X	X
Fluorene	86-73-7	X	X	X	X	X
2-Fluorobiphenyl (surr)	321-60-8	X	X	X	X	X
2-Fluorophenol (surr)	367-12-4	X	X	X	X	X
Heptachlor	76-44-8	X	X	X	X	X
Heptachlor epoxide	1024-57-3	X	X	X	X	X
Hexachlorobenzene	118-74-1	X	X	X	X	X
Hexachlorobutadiene	87-68-3	X	X	X	X	X
Hexachlorocyclopentadiene	77-47-4	X	X	X	X	X
Hexachloroethane	67-72-1	X	X	X	X	X
Hexachlorophene	70-30-4	AW,CP(62)	ND	ND	ND	CP
Hexachloropropene	1888-71-7	X	ND	ND	ND	X
Hexamethylphosphoramide	680-31-9	X	ND	ND	ND	X
Hydroquinone	123-31-9	ND	ND	ND	ND	X
Indeno(1,2,3-cd)pyrene	193-39-5	X	X	X	X	X
Isodrin	465-73-6	X	ND	ND	ND	X
Isophorone	78-59-1	X	X	X	X	X
Isosafrole	120-58-1	DC(46)	ND	ND	ND	X
Kepone	143-50-0	X	ND	ND	ND	X
Leptophos	21609-90-5	X	ND	ND	ND	X
Malathion	121-75-5	HS(5)	ND	ND	ND	X
Maleic anhydride	108-31-6	HE	ND	ND	ND	X
Mestranol	72-33-3	X	ND	ND	ND	X
Methapyrilene	91-80-5	X	ND	ND	ND	X
Methoxychlor	72-43-5	X	ND	ND	ND	X
3-Methylcholanthrene	56-49-5	X	ND	ND	ND	X

4,4'-Methylenebis (2-chloroaniline)	101-14-4	OE,OS(0)	ND	ND	ND	LR

Appropriate Preparation Techniques[b]

Compounds	CAS No[a]	3510	3520	3540/ 3541	3550	3580
4,4'-Methylenebis (N,N-dimethylaniline)	101-61-1	X	X	ND	ND	ND
Methyl methanesulfonate	66-27-3	X	ND	ND	ND	X
2-Methylnaphthalene	91-57-6	X	X	ND	X	X
Methyl parathion	298-00-0	X	ND	ND	ND	X
2-Methylphenol	95-48-7	X	ND	ND	ND	X
3-Methylphenol	108-39-4	X	ND	ND	ND	X
4-Methylphenol	106-44-5	X	ND	ND	ND	X
Mevinphos	7786-34-7	X	ND	ND	ND	X
Mexacarbate	315-18-4	HE,HS(68)	ND	ND	ND	X
Mirex	2385-85-5	X	ND	ND	ND	X
Monocrotophos	6923-22-4	HE	ND	ND	ND	X
Naled	300-76-5	X	ND	ND	ND	X
Naphthalene	91-20-3	X	X	X	X	X
Naphthalene-d_8 (IS)		X	X	X	X	X
1,4-Naphthoquinone	130-15-4	X	ND	ND	ND	X
1-Naphthylamine	134-32-7	OS(44)	ND	ND	ND	X
2-Naphthylamine	91-59-8	X	ND	ND	ND	X
Nicotine	54-11-5	DE(67)	ND	ND	ND	X
5-Nitroacenaphthene	602-87-9	X	ND	ND	ND	X
2-Nitroaniline	88-74-4	X	X	ND	X	X
3-Nitroaniline	99-09-2	X	X	ND	X	X
4-Nitroaniline	100-01-6	X	X	ND	X	X
5-Nitro-o-anisidine	99-59-2	X	ND	ND	ND	X
Nitrobenzene	98-95-3	X	X	X	X	X
Nitrobenzene-d_5 (surr)		X	X	X	X	X
4-Nitrobiphenyl	92-93-3	X	ND	ND	ND	X
Nitrofen	1836-75-5	X	ND	ND	ND	X
2-Nitrophenol	88-75-5	X	X	X	X	X
4-Nitrophenol	100-02-7	X	X	X	X	X
5-Nitro-o-toluidine	99-55-8	X	X	ND	ND	X
Nitroquinoline-1-oxide	56-57-5	X	ND	ND	ND	X
N-Nitrosodi-n-butylamine	924-16-3	X	ND	ND	ND	X
N-Nitrosodiethylamine	55-18-5	X	ND	ND	ND	X
N-Nitrosodimethylamine	62-75-9	X	X	X	X	X
N-Nitrosomethylethylamine	10595-95-6	X	ND	ND	ND	X
N-Nitrosodiphenylamine	86-30-6	X	X	X	X	X
N-Nitrosodi-n-propylamine	621-64-7	X	X	X	X	X
N-Nitrosomorpholine	59-89-2	ND	ND	ND	ND	X
N-Nitrosopiperidine	100-75-4	X	ND	ND	ND	X
N-Nitrosopyrrolidine	930-55-2	X	ND	ND	ND	X

Octamethyl pyrophosphoramide	152-16-9	LR	ND	ND	ND	LR
4,4'-Oxydianiline	101-80-4	X	ND	ND	ND	X

ropriate Preparation Techniques[b]

Compounds	CAS No[a]	3510	3520	3540/ 3541	3550	3580
Parathion	56-38-2	X	X	ND	ND	X
Pentachlorobenzene	608-93-5	X	ND	ND	ND	X
Pentachloronitrobenzene	82-68-8	X	ND	ND	ND	X
Pentachlorophenol	87-86-5	X	X	X	X	X
Perylene-d_{12} (IS)		X	X	X	X	X
Phenacetin	62-44-2	X	ND	ND	ND	X
Phenanthrene	85-01-8	X	X	X	X	X
Phenanthrene-d_{10} (IS)		X	X	X	X	X
Phenobarbital	50-06-6	X	ND	ND	ND	X
Phenol	108-95-2	DC(28)	X	X	X	X
Phenol-d_6 (surr)		DC(28)	X	X	X	X
1,4-Phenylenediamine	106-50-3	X	ND	ND	ND	X
Phorate	298-02-2	X	ND	ND	ND	X
Phosalone	2310-17-0	HS(65)	ND	ND	ND	X
Phosmet	732-11-6	HS(15)	ND	ND	ND	X
Phosphamidon	13171-21-6	HE(63)	ND	ND	ND	X
Phthalic anhydride	85-44-9	CP,HE(1)	ND	ND	ND	CP
2-Picoline (2-Methylpyridine)	109-06-8	X	X	ND	ND	ND
Piperonyl sulfoxide	120-62-7	X	ND	ND	ND	X
Pronamide	23950-58-5	X	ND	ND	ND	X
Propylthiouracil	51-52-5	LR	ND	ND	ND	LR
Pyrene	129-00-0	X	X	X	X	X
Pyridine	110-86-1	ND	ND	ND	ND	ND
Resorcinol	108-46-3	DC,OE(10)	ND	ND	ND	X
Safrole	94-59-7	X	ND	ND	ND	X
Strychnine	57-24-9	AW,0S(55)	ND	ND	ND	X
Sulfallate	95-06-7	X	ND	ND	ND	X
Terbufos	13071-79-9	X	ND	ND	ND	X
Terphenyl-d_{14}(surr)	1718-51-0	X	X	ND	X	X
1,2,4,5-Tetrachlorobenzene	95-94-3	X	ND	ND	ND	X
2,3,4,6-Tetrachlorophenol	58-90-2	X	ND	ND	ND	X
Tetrachlorvinphos	961-11-5	X	ND	ND	ND	X
Tetraethyl dithiopyrophosphate	3689-24-5	X	X	ND	ND	ND
Tetraethyl pyrophosphate	107-49-3	X	ND	ND	ND	X
Thionazine	297-97-2	X	ND	ND	ND	X
Thiophenol (Benzenethiol)	108-98-5	X	ND	ND	ND	X
Toluene diisocyanate	584-84-9	HE(6)	ND	ND	ND	X
o-Toluidine	95-53-4	X	ND	ND	ND	X
Toxaphene	8001-35-2	X	X	X	X	X
2,4,6-Tribromophenol (surr)	118-79-6	X	X	X	X	X

1,2,4-Trichlorobenzene	120-82-1	X	X	X	X	X
2,4,5-Trichlorophenol	95-95-4	X	X	ND	X	X
2,4,6-Trichlorophenol	88-06-2	X	X	X	X	X
Trifluralin	1582-09-8	X	ND	ND	ND	X

		Appropriate Preparation Techniques[b]				
Compounds	CAS No[a]	3510	3520	3540/ 3541	3550	3580
2,4,5-Trimethylaniline	137-17-7	X	ND	ND	ND	X
Trimethyl phosphate	512-56-1	HE(60)	ND	ND	ND	X
1,3,5-Trinitrobenzene	99-35-4	X	ND	ND	ND	X
Tris(2,3-dibromopropyl) phosphate	126-72-7	X	ND	ND	ND	LR
Tri-p-tolyl phosphate	78-32-0	X	ND	ND	ND	X
O,O,O-Triethyl phosphorothioate	126-68-1	X	ND	ND	ND	X

[a] Chemical Abstract Service Registry Number
[b] See Sec. 1.2 for other acceptable preparation methods.

KEY TO ANALYTE LIST

IS = This compound may be used as an internal standard.
surr = This compound may be used as a surrogate.
AW = Adsorption to walls of glassware during extraction and storage.
CP = Nonreproducible chromatographic performance.
DC = Unfavorable distribution coefficient (number in parenthesis is percent recovery).
HE = Hydrolysis during extraction accelerated by acidic or basic conditions (number in parenthesis is percent recovery).
HS = Hydrolysis during storage (number in parenthesis is percent stability).
LR = Low response.
ND = Not determined.
OE = Oxidation during extraction accelerated by basic conditions (number in parenthesis is percent recovery).
OS = Oxidation during storage (number in parenthesis is percent stability).
X = Greater than 70 percent recovery by this technique.

1.4. The following compounds may require special treatment when being determined by this method. Benzidine can be subject to oxidative losses during solvent concentration. Also, the Benzidine chromatography is poor. Under the alkaline conditions of the extraction step, α-BHC, γ-BHC, Endosulfan I and II, and Endrin are subject to decomposition. Neutral extraction should be performed if these compounds are expected. Hexachlorocyclopentadiene is subject to thermal decomposition in the inlet of the gas chromatograph, chemical reaction in acetone

1.5. The practical quantitation limit (PQL) of Method 8270C for determining an individual compound is approximately .2 mg/kg (wet weight) for soil/sediment samples; 1-200 mg/kg for wastes (depending on matrix and method of preparation); and, 10 µg/L for ground water samples. PQLs will be proportionately higher for samples extracts that require dilution to avoid detector saturation.

1.6. This method is restricted to use by or under the supervision of analysts experienced in the use of gas chromatography/mass spectrometers and skilled in the interpretation of mass spectra. Each analyst must demonstrate the ability to generate acceptable results with this method by completing a IDOC that complies with the HEAL QAP.

2.0 Summary of Method

2.1. The samples are prepared for analysis by gas chromatography/mass spectrometry (GC/MS) using the appropriate sample preparation (refer to Method 3500) and, if necessary, sample cleanup procedures (refer to Method 3600).

2.2. The semi-volatile compounds are introduced into the GC/MS by injecting the sample extract into a gas chromatograph (GC) with a narrow-bore fused-silica capillary column. The GC column is temperature-programmed to separate the analytes, which are then detected with a mass spectrometer (MS) connected to the gas chromatograph.

2.3. Analytes eluted from the capillary column are introduced into the mass spectrometer via a direct connection. Identification of target analytes is accomplished by comparing their mass spectra with the electron impact (or electron impact-like) spectra of authentic standards. Quantitation is accomplished by comparing the response of a major (quantitation) ion relative to an internal standard using a five-point calibration curve at a minimum.

3.0 Safety

3.1. Some of the compounds that are used are known carcinogens, mutagens, and or teratogens. So the utmost care must be used in handling the neat compounds and the mixes made from them. At a minimum, latex gloves, a lab coat, and safety glasses must be worn to avoid any unnecessary contact. Plus any handling of the open containers of neat compounds and/or mixes must be done in a ventilated hood to avoid inhalation.

3.1.1. Working standards also need to be handled with care. When handling them you need at a minimum to wear latex gloves, safety glasses and a lab coat.

3.2. Latex gloves, lab coat, and safety glasses must be worn when handling solvents. Extractions can only be done if more than one person is in the same area as the person doing the extract.

3.3. When the analysts are done with standard vials they are disposed of in the appropriate manner. All standard vials, samples, and other laboratory-generated waste will be disposed in accordance to the HEAL Chemical Hygiene Plan.

4.0 Definitions

Continuing Calibration Verification (CCV) – standard from the same source as the curve used to check the stability of the instrument.
Calibration check compounds (CCCs) – used to evaluate the integrity of the chromatographic system
System performance check compounds (SPCCs) – used to check the minimum average RFs of the chromatographic system.
RRT – Relative Retention Time
Laboratory Control Spike (LCS) – laboratory fortified blank
Laboratory Control Spike Duplicate (LCSD) – laboratory fortified blank duplicate
Initial Calibration Verificatoin (ICV) – a second source standard that is used to determine the state of the calibration of an instrument after initial calibration.
Matrix Spike (MS) – laboratory fortified sample
Matrix Spike Duplicate (MSD) – laboratory fortified sample duplicate
Matrix Blank (MBLK) – laboratory blank
Method Detection Limit (MDL) – the constituent concentration that when processed through the complete method, produces a signal with a 99% probability that it is different than the blank.
Practical Quantitation Limit (PQL) – The lowest calibration point used in the initial calibration.
Sample Duplicate (DUP) – duplicate of sample.

Surrogate – a particular compound that is used to determine the overall efficiency of the extraction process.
Internal Standards – used for calibrating the instrument.
GC – Gas Chromatograph.
MS – Mass Spectrometer.

5.0 Interferences

5.1. Refer to Methods 3500, 3600, and 8000.

5.2. Raw GC/MS data from all blank, spike and client samples must be evaluated for interferences to determine if the source of the interferences are in the preparation and/or cleanup.

5.3. Contamination from carry over can occur when high-concentration samples are analyzed. To reduce carryover, the sample syringe must be rinsed between samples with solvent. Whenever an unusually concentrated sample is encountered, it should be followed by the analysis of a solvent blank to check for cross contamination.

5.4. Loss of surrogates may be due to formation of emulsions with the extraction solvent in some aqueous samples. This could also be due to excessive Sodium Sulfate added as a drying agent for the solvent.

6.0 Apparatus and Reagents

6.1. Gas chromatograph/mass spectrometer system

6.1.1. **Instrument name: ELMO.**

6.1.2. Varian 3800 gas chromatograph equipped with a Varian 1177 split/split-less injection port.

6.1.3. Column: Varian VF-5MS measuring 30m x 0.25mm ID with a 0.25μm film thickness (or equivalent).

6.1.4. Autosampler : Varian 8200 CX

6.1.5. Mass spectrometer: Varian SATURN 2000.

6.1.6. Data system: Saturn GC/MS Workstation Ver. 6.9.

6.2. Volumetric flask: Class A 1-100 ml, with a ground-glass stopper.

6.3. Graduated cylinder: 500 ml.

6.4. Micro-syringes: 10-1000 μL Hamilton and SGE.

6.5. Dionex: Accelerated Solvent Extractor (see SOP S-3545 for more detail).

6.6. Kuderna-Danish (K-D) apparatus (see SOP 3510C-5 for more detail).

6.7. Boiling chips: Chemware PTFE Boiling stones.

6.8. Heated Water bath: Capable of temperature control (±5°C). The bath should be used in an extraction hood.

6.9. Sample vials: 1.8 ml Amber Glass with Teflon-lined screw caps.

6.10. Disposable pipettes: Pasteur.

6.11. Spatula: Stainless steel

6.12. Organomation Associates: N-EVAP 112 Nitrogen Evaporator.

6.13. 1000ml Teflon Separatory funnels.

6.14. 250ml round bottom flasks.

6.15. Soxhlet extraction apparatus with water-cooled condensers.

6.16. Whatman cellulose extraction thimbles, cat.no 2800338

6.17. Extraction cells that will accommodate 20g or 30g of sample

6.18. Vials for collection of extracts 40ml or 60ml, precleaned, open top with screw cap with PTFE-lined silicone septa. (Dionex 49459, 49460, 49461 or 49462 or equivalent)

6.19. Filter disk- 1.91cm type D28 (Whatman 10289356 or equivalent)

6.20. Cell cap sealing disk (Dionex 49454, 49455 or equivalent)

6.21. 250ml Erlenmeyer flasks.

6.22. 500ml Erlenmeyer flasks.

6.23. Wide range pH test strips.

6.24. Reagent grade organic chemicals shall be used in all tests. Unless otherwise indicated, it is intended that all inorganic reagents shall conform to the specifications of the Committee on Analytical Reagents of the American Chemical Society, where such specifications are available. Other grades may be used, provided it is first ascertained that the reagent is of sufficiently high purity to permit its use without lessening the accuracy of the determination

6.25. 1:1 Sulfuric acid, H_2SO_4

6.26. Solvents: Organic free reagent water (Deionized Water) , Acetone, Methylene Chloride: pesticide quality or equivalent.

6.27. Anhydrous Sodium sulfate: EM Science Tracepur ACS

6.28. 10N NaOH.

6.29. Stock standard solutions:

6.30. A 200 PPM Ready Stock standard is used. Supplied by NSI, part number WL-507. If necessary, refer to Method 8270C for instructions on how to make Standards from neat compounds.

6.30.1. Transfer the stock standard solutions into Teflon-sealed screw-cap amber vials. Store at <6° C and protect from light. Stock standards should be frequently checked for signs of degradation or evaporation

6.30.2. Stock standard solutions must be replaced after one year or sooner if comparison with check standards indicates a problem. Stock standards are disposed of if they reach their expiration date

6.31. A separate spike mix is also required to spike LCS/LCSD or MS/MSD. This is purchased from NSI, part number WL-365 for 8270 full list compounds.

6.32. Spike NSI cat # C-447 is used for TCLP extracts.

6.33. Internal standard solutions: The internal standards used are 1,4-dichlorobenzene-d4, naphthalene-d8, Acenaphthene-d10, Phenanthrene-d10, Chrysene-d12, and perylene-d12. These are purchased ready to use from NSI at a concentration of 2000 ppm, NSI part number C-394-33. Refer to Method 8270C for detailed instructions on how to make standards from neat compounds.

6.34. GC/MS tuning standard: A methylene chloride solution containing 50 ppm of decafluorotriphenylphosphine (DFTPP) should be prepared from Stock tune mix at 500 ppm. This is obtained from NSI, part number C-491-10. The standard should also contain 50 ppm each of 4,4'-DDT, Pentachlorophenol, and Benzidine to verify injection port inertness and GC column performance. Store standard solution at -10°C or less when not being used.

6.35. Surrogate standards: The surrogate standards used are phenol-d5, 2-Fluorophenol-2,4,6-Tribromophenol, Nitrobenzene-d5, 2-Fluorobiphenyl, and p-Terphenyl-d14. These are purchased from NSI part number WL-371 in a mix at a concentration of 200ppm for phenols and 100ppm for the other compounds. See the extraction method for the instructions on preparing the surrogate standards. Determine what concentration should be in the blank extracts after all extraction, cleanup, and concentration steps. Inject this concentration into the GC/MS to determine recovery of surrogate standards in all blanks, spikes, and sample extracts. Take into account all dilutions of sample extracts.

6.36. Matrix spike standards: The matrix spike standards contain the both Base/Neutral and Acid compounds. The acid fraction of the mix is Pentachlorophenol, Phenol, 2-Chlorophenol, 4-Chloro-2-methylphenol and 4-Nitrophenol. The Base/Neutral fraction of the mix contains 1,2,4 Trichlorobenzene, Acenapthene, 2,4-Dinitrotoluene, Pyrene, N-Nitroso-di-n-propylamine and 1,4-Dichlorobenzene. This mix is purchased from NSI part number WL-365. The concentration of the acids in the mixture is 200µg/mL and the Base/Neutral compounds are 100µg/mL. See the extraction method 3500B, section 5.5.1 for the instructions on preparing the matrix spike standards.

7.0 Sample Collection Preservation and Handling

7.1. Refer to section 5.0 of the HEAL QAP for proper preservation and handling.

7.2. Water samples are collected in glass amber jars (500mL) or one liter and are unpreserved.

7.3. Soil samples are collected in glass jars 2oz or 4oz.

7.4. All samples should be kept above freezing and $<6^0$C from time of collection to just prior to analysis.

7.5. Once extracted, store the sample extracts less than -10°C, protected from light, in sealed vials (e.g., screw-cap vials or crimp-capped vials) equipped with unpierced PTFE-lined septa.

8.0 Operating conditions

8.1 Typical GC/MS **ELMO** operating conditions: To view current operating conditions refer to the newest calibration curve on the GCMS software. Operating conditions may vary depending on client needs.

Mass range:	45-650 m/z
Scan time:	0.61 sec/scan
Initial temperature:	40°C, no hold
Ramp:	40°C-310°C at 15°C/min.
Final temperature:	310°C, hold 4.5 min
Injector temperature:	275°C
Transfer line temp:	280°C
Trap temperature:	220°C
Manifold Temperature:	60°C
Injector:	Split/split with 20:1 split injection.
Injection volume:	1µL
Carrier gas:	1.1 mL/min.

9.0 Initial Calibration Information

9.1. Calibration Standards: Calibration standards at a minimum of five concentration levels are prepared through dilution of the stock standards with methylene chloride (see table below). The standards are made in a strict order. The methylene chloride is added to the vial, then the correct amount of Ready Stock and finally the 20ul of Internal Standard is added. One of the concentration levels should be a concentration at the practical quantitation limit (PQL). The remaining concentration levels should correspond to the expected range of concentrations found in real samples or should define the working range of the GC/MS system. Each standard should contain each analyte for detection by this method (e.g. some or all of the compounds listed in table 1 may be included). Each 1mL aliquot of calibration standard should be spiked with 20µL of the internal standard solution prior to analysis. Store at -10°C or less when not being used. Calibration solutions must be replaced after one year (or sooner if comparison with a check standard indicates a problem). The daily calibration standard should be prepared weekly and stored at <6° C. A new curve should be prepared and run after the response in any of the CCC's or SPCC's has changed making them fail.

Standard Conc. PPM	Vol. Standard* added (µL)	Vol. DCM added (µL)	Vol. Internal Std (µL)
2.5 PPM	12.5	967.5	20
5.0 PPM	25	955	20
10 PPM	50	930	20
20 PPM	100	880	20
30 PPM	150	830	20
40 PPM	200	780	20
50 PPM	250	730	20
80 PPM	400	580	20
100 PPM	500	480	20
120 PPM	600	380	20

- 200 PPM Ready stock standard from NSI.

9.2. An ICV is prepared from a second source from NSI purchased at a concentration of 50ppm. The working ICV standard is prepared at 30ppm by spiking 600uL of the 50ppm stock into 380uL of DCM. Then 20uL of Internal standard is added. The criteria used to determine if the ICV is acceptable can be found in section 12.2.2.

9.3. The GC/MS system must be hardware-tuned to meet the criteria for method 525 in Table 2 for a 50ng injection of DFTPP in order to analyze an initial calibration. Refer to section 11.1.1. for more tune criteria

9.4. For the curve to be in compliance the minimum average RF for all SPCCs must be 0.050. Refer to section 11.1.3.2. for the list of SPCCs.

9.5. When a curve is analyzed, each individual compound may be analyzed using linear regression (with a minimum of 5 calibration points) or quadratic regression (with a minimum of 6 calibration points. The R^2 for each compound must be >/=0.990.

9.6. The RSD for any CCCs must be less than 30% in order for the curve to be compliant. If the CCC RSD is greater than 15% but less than 30% a linear or quadratic curve may be used. Refer to section 11.1.3.3. for the list of CCCs.

9.7. If the RSD for any CCC is greater than 30% or the RF for any SPCC is less than 0.050 the system is too reactive to begin analysis. Clean up the system and recalibrate.

10.0 Extraction and final volume

10.1. Extraction:

10.1.1. Water samples are extracted first at a pH <2 with methylene chloride, and then at pH >11 using methylene chloride using Method 3510, refer to the current SOP S-3510C.

10.1.2. Solid samples are extracted using Method 3540, refer to the current SOP S-3540C or Method 3545, refer to the current SOP S-3545.

10.1.3. Chemical wastes are diluted prior to analysis by weighing out 0.2g into a 10ml volumetric. Surrogate is added after, usually adding 500ul to 500ul of sample and then adding in 20ul of internal standard.

10.1.4. Laboratory control spike (LCS), Laboratory control spike duplicate (LCSD), and Matrix blank (MBLK) are to be extracted with aqueous samples, unless otherwise indicated by a client. Soil Samples call for Matrix spikes (MS) and Matrix (MSD) spike duplicates as well as LCS and MBLK.

10.1.4.1. Add 500µl WL-365 spike mix to LCS, LCSD MS and MSD.

10.1.4.2. Add 500 µl of NSI BNA Surrogates mix 200/100 µg/ml to all samples.

10.2. Sample concentration and final volume

10.2.1. To achieve maximum sensitivity with this method, the extract must be concentrated to 1.0 ml. Then 20ul of 2000ppm Internal standard is added to bring sample to final volume of 1ml. Refer to Method 3510 for Concentration steps. Final volume is achieved by gently blowing the sample down in the concentrator nipple after it has been boiled down according to the Method 3510. Nitrogen gas is blown across the top of the nipple, being careful not to blow directly into the nipple causing loss of volume by splashing. This is continued until there is approximately 800 ul of sample in the nipple (taking into account boiling chips increase volume). The sample is then transferred to a clean, labeled amber vial. The nipple is then rinsed with approximately 300 ul of clean Methylene Chloride and also transferred to the vial. The sample is then measured with a 1ml syringe. 20ul of internal standard is then added to the sample. If the volume of the sample exceeds 1ml it may be blown down further until 1ml final volume is achieved. Some samples will require larger final volumes due to gross contamination. The final volume is left to the discretion of the analyst.

11.0 Analysis: Verification, Procedure and Interpretation

11.1. GC/MS Calibration verification.

11.1.1. The GC/MS system must be hardware-tuned to meet the criteria for method 525 in Table 2 for a 50ng injection of DFTPP. The parameters are located in the Saturn Software. Analysis does not begin until all these criteria are met. Pentachlorophenol, 4,4'-DDT and Benzidine are also included at 50ng in the tune mix to verify injector port inertness and GC column performance by being present at their normal response with no visible signs of tailing. Pentachlorophenol and Benzidine should be present at 25% to 50% the height of DFTPP. DDT should be present at 80-120% the height of the DFTPP peak. Degradation of DDT to DDD and DDE must not exceed 20% (see sec.8.0 of method 8081 for % breakdown calculation). The production of DDD and DDE can be monitored by plotting their primary ion DDD (235) and DDE (246) against the DDT (235). Areas for these peaks can then be used in the calculation from Method 8081 to determine whether break down exceeds 20%. Tailing factor for pentachlorophenol and benzidine must be examined from the same 50ng injection of DFTPP. See figure 13 from method 625 for the calculations of the tailing factor. The tailing factor for pentachlorophenol must be less than 5 and for benzidine less than 3.

11.1.1.1. The frequency of the DFTPP check, Degradation of DDT and the tailing factors for pentachlorophenol and benzinde must be performed every 12 hours of analysis.

11.1.2. The initial calibration (Sec. 9.0) for each compound of interest must be checked and verified once every 12 hours during analysis. This is accomplished by analyzing a calibration standard (CCV) that is a concentration near the midpoint concentration for the working range of the GC/MS and then checking the SPCC's and CCC's. The concentration of the CCV must vary.

11.1.3. System Performance Check Compounds (SPCCs): A system performance check must be performed every 12 hour shift. If the SPCC criteria are met, a comparison of relative response factors is made for all compounds. This is the same check that is applied during the initial calibration. If the minimum relative response factors are not met, the system must be evaluated, and corrective action must be taken before sample analysis begins. Some possible problems are standard mixture degradation, injection port inlet contamination, contamination at the front end of the analytical column, and active sites in the column or chromatographic system. This check must be met before analysis begins.

11.1.3.1. The minimum relative response factor for all SPCC's should be: ≥0.05

11.1.3.2. The SPCC's are N-nitroso-di-n-propylamine, Hexachlorocyclopentadiene, 2,4-Dinitrophenol and 4-Nitrophenol.

11.1.3.3. The CCC's are Acenapthene, Hexachlorobutadiene, Di-n-octyl phthalate, Benzo(a)pyrene, 1,4-Dichlorobenzene, Diphenylamine and Flouranthene. These represent the Base/Neutral fraction. 4-Chloro-3-methylphenol, 2-Nitrophenol, Pentachlorophenol, 2,4-Dichlorophenol, Phenol and 2,4,6-Trichlorophenol represent the acid fraction.

11.1.3.4. Response factors can be manually calculated by:

$$RF = (A_x * C_{is}) / (A_{is} * C_x)$$

Where:

A_x = Area of the characteristic ion for the compound being measured.
A_{is} = Area of the characteristic ion for the specific internal standard.
C_{is} = Concentration of the specific internal standard (ng/µL).
C_x = Concentration of the compound being measured (ng/µL).

11.1.4. Calibration Check Compounds (CCCs) - After the system performance check is met, CCCs listed in (sec. 11.1.3.3.) are used to check the validity of the initial calibration.

Calculate the percent drift using:

$$\% \text{ Drift} = \{(C_1 - C_c) / C_1\} * 100$$

where:

C_1 = Calibration Check Compound standard concentration.
C_c = Measured concentration used in selected quantitation method.

If the percent drift for each CCC is less than or equal to 20%, the initial calibration is assumed to be valid. If the criterion is not met (>20% drift) for any one CCC, corrective action must be taken. Problems similar to those listed under SPCCs could affect this criterion. If no source of the problem can be determined after corrective action has been taken, a five-point curve calibration at a minimum should be generated. This criterion must be met before sample analysis begins. If the CCCs are not analytes required by the permit, then all required analytes must meet the 20% drift criterion.

11.1.5. Evaluating the CCV the target compounds (non-CCC) must have a recovery from 60% to 140% of the expected concentration in order for samples to be reported.

11.1.6. Internal standard retention time: The retention times of the internal standards in the CCV must be evaluated immediately after the data acquisition. If the retention time for any internal standard changes by more than 30 seconds from that in the mid-point standard level of the most recent initial calibration sequence, then the chromatographic system must be inspected for malfunctions and corrections must be made, as required. When corrections are made, reanalysis of samples analyzed while the system was malfunctioning is required.

11.1.7. Internal standard response: If the area for any of the internal standards in the CCV changes by a factor of two (-50% to +100%) from that in the mid-point standard level of the most recent initial calibration sequence, the mass spectrometer must be inspected for malfunctions and corrections must be made, as appropriate. When corrections are made, reanalysis of samples analyzed while the system was malfunctioning is required.

11.2. GC/MS analysis

11.2.1. All samples and standard solutions must be allowed to warm to ambient temperature before analysis.

11.2.2. DFTPP tuning criteria and GC/MS calibration verification criteria must be met before analyzing samples.

11.2.3. The auto-sampler injects 1µL of sample directly on to the column, using the same operating conditions as were used in calibration see section 9.0. The volume to be injected should contain 50ng of base/neutral and 100ng of acid surrogates (assuming 100% recovery). The injection volume must be the same volume used for the calibration standards.

11.2.4. Any necessary sample dilutions are performed manually by the analyst. This may be prior to initial analysis i.e. sample can not be brought to 1mL final volume. The other instance will be when a sample is analyzed and certain components may exceed the working range of the curve.

11.3. Data interpretation

11.3.1. Qualitative analysis

11.3.1.1. The qualitative identification of compounds determined by this SOP is based on retention time, and on comparison of the sample mass

spectrum, after background correction, with characteristic ions in a reference mass spectrum. The reference mass spectrum is generated by using the conditions described in this SOP. The characteristic ions from the reference mass spectrum are defined to be the three ions of greatest relative intensity, or any ions over 30% relative intensity if less than three such ions occur in the reference spectrum. Compounds are identified as present when the criteria below are met.

11.3.1.1.1. The intensities of the characteristic ions of a compound maximize in the same scan or within one scan of each other. Selection of a peak by a data system target compound search routine where the search is based on the presence of a target chromatographic peak containing ions specific for the target compound at a compound-specific retention time will be accepted as meeting this criteria.

11.3.1.1.2. The relative intensities of the characteristic ions should agree within 30% of the relative intensities of these ions in the reference spectrum. (Example: For an ion with an abundance of 50% in the reference spectrum, the corresponding abundance in the sample spectrum can range between 20% and 80%). While checking the relative intensities is a good indicator of qualifying a compound it is left to a qualified mass spec interpreter to determine if a compound can be identified as a positive result.

11.3.1.1.3. Some analytes may contain qual ions which are not 30% or greater in intensity. In such instances, only qual ions that are at a minimum of 10% intensity should be used.

11.3.1.1.4. Some analytes may only have one qual ion which is also the quan ion. In such a case, selection of the quan ion as the qual ion is the only solution.

11.3.1.1.5. Should a sample contain an analyte that has one or more qual ions that does not meet the +/-30% requirement, a more thorough visual inspection and a library search should be completed. If the library search returns a match for the analyte, a printout showing the library search results should be included. The original chromatograph report should be notated in such a manner that is clear that the analyst is accepting the quantitative result based on the library search return. The Technical Director, or his designee, can also

sign off on acceptance of spectra, when multiple compounds are returned in a library search.

11.3.1.1.6. The RRT of the sample component is within +/- 0.06 RRT units of the standard component.

11.3.1.1.7. Structural isomers that produce very similar mass spectra should be identified as individual isomers if the have sufficiently different GC retention times. Sufficient GC resolution is achieved if the height of the valley between the two isomer peaks is less than 25% of the sum of the two peak heights. Otherwise, structural isomers are identified as isomeric pairs.

11.3.1.1.8. Identification is hampered when sample components are not resolved chromatographically and produce mass spectra containing ions contributed by more than one analyte. When gas chromatographic peaks obviously represent more than one sample component (i.e. , a broadened peak with shoulder(s) or a valley between two or more maxima), appropriate selection of analyte spectra and background spectra is important. Examination of extracted ion current profiles of appropriate ions can aid in the selection of spectra and in qualitative identification of compounds. When analytes co-elute (i.e., only one chromatographic peak is apparent), the identification criteria can be met, but each analyte spectrum will contain extraneous ions contributed by the co-eluting compound.

11.3.1.1.9. If any of the above mentioned scenarios are unable to be met for a specific analyte, guidance from the Technical Director should be sought. The Technical Director can determine if the quantified spectra should be reported.

11.3.1.2. For samples containing components not associated with the calibration standards, a library search may be made for the purpose of tentative identification. The necessity to perform this type of identification will be determined by the purpose of the analyses being conducted. Computer generated library search routines should not use normalization routines that would misrepresent the library or unknown spectra when compared to each other. For example, RCRA permit or waste delisting requirements may require the reporting of non-target analytes. Only after visual comparison of sample spectra with the nearest library searches will the mass spectral interpretation specialist

assign a tentative identification. Guidelines for making tentative identification are:

(1)Relative intensities of major ions in the reference spectrum (ions> 10% of the most abundant ion) should be present in the sample spectrum.

(2)The relative intensities of the major ions should agree within ±20%. (Example: For ions with an abundance of 50% in the standard spectrum, the corresponding sample ion abundance must be between 30 and 70%)

(3)Molecular ions present in the reference spectrum should be present in the sample spectrum.

(4)Ions present in the sample spectrum but not in the reference spectrum should be reviewed for possible background contamination or presence of co-eluting compounds.

(5)Ions present in the reference spectrum but not in the sample spectrum should be reviewed for possible subtraction, because of background contamination or co-eluting peaks. Data system library reduction programs can sometimes create these discrepancies.

11.3.2. Quantitative analysis

11.3.2.1. When a compound has been identified, the quantitation of that compound will be based on the integrated abundance from the EICP of the primary characteristic ion. Quantitation will take place using the internal standard technique. The internal standard used shall be the one nearest the retention time of that of a given analyte, see table 5 in method 8270C. See the HEAL QAP section 9.0 for necessary calculations.

11.3.2.2. When MS response is linear and passes through the origin, calculate the concentration of each identified analyte in the sample as follows:

Water

Concentration (µg/L) = $(A_x)(I_s) / (A_{is})(RF)(V_o)$

where:

A_x =Area of characteristic ion for compound being measured.
I_s = Amount of internal standard injected (ng).

A_{is} = Area of characteristic ion for the internal standard.
RF =Mean relative response factor for compound being measured.
V_o = Volume of extracted sample (mL), taking into consideration any dilutions made.

Sediment/Soil sludge (normally on a wet-weight basis) and Waste (normally on a wet weight basis)

Concentration in solid (µg/kg) = $(A_x)(I_s)(V_t) / (A_{is})(RF)(V_i)(W_s)(D)$

where:

A_x, I_s, A_{is}, RF = Same as for water.
V_{st} = Volume of total extract (µL) (use 10,000 µL or a factor of this when dilutions are made).
V_i = Volume of extracted add (µL) for purging.
W_s = Weight of sample extracted or purged (g).
D = % dry weight of sample /100, or 1 for a wet-weight basis.

11.3.2.3. Where applicable, an estimate of concentration for non-calibrated components in the sample can be made. The formulas given above should be used with the following modifications: the areas A_x and A_{is} should be from the total ion chromatograms and the RF for the compound should be assumed to be 1. The concentration obtained should be reported indicating (1) that the value is an estimate and (2) which internal standard was used to determine concentration. Use the nearest internal standard free of interferences.

11.3.2.4. Alternatively, the regression line fitted to the initial calibration may be used for determination of the analyte concentration.

12.0 Quality Control and Method Performance

12.1. Refer to Method 8000C and the HEAL Quality Assurance Plan for quality control procedures.

12.2. The following are general guidelines for data assessment and acceptance criteria for quality control measures.

12.2.1. The GC/MS system must be tuned to meet the DFTPP specifications every 12hr shift.

12.2.2. There must be an initial calibration of the GC/MS system. The ICV must be analyzed following a calibration curve and prior to any sample analysis. All failures or adjustments to the initial calibration curve shall be documented on the form located in the Appendix. This form shall be included with each data folder associated with the current calibration. The acceptance criterion for the ICV is the same as the criteria for the CCV as described in section 11.0. Then every 12 hours a CCV will be analyzed as a calibration check for the Instrument after a DFTPP tune.

12.2.3. The concentrations of the CCV should vary every 12 hour shift.

12.2.4. The absolute retention time should not exceed 0.8min of the initial calibration. If it does shift more than 0.8 min. a new curve should be analyzed.

12.2.5. The GC/MS system must meet the SPCC criteria and the CCC criteria, every 12 hours.

12.2.6. For every 20 Aqueous samples extracted, there must be an MBLK, LCS and LCSD (when there is not enough sample to do an MS/MSD or Dup). When clients provide enough sample for an MS and MSD or a sample DUP then the LCSD does not need to be extracted. With every 20 soil/waste samples extracted there must be a MBLK, LCS and MS/MSD extracted as QC.

12.2.6.1. Method 8270 stipulates that 11 compounds are to be used for the MS and the MSD. The LCS and LCSD will also use these 11 compounds. They are 4-chloro-3-methylphenol, 2-Chlorophenol, 4-Nitrophenol, Pentachlorophenol, Phenol, Acenapthene, 1,4-Dichlorobenzene, 2,4-Dinitrotoluene, N-Nitrosodi-n-propylamine, Pyrene and 1,2,4-Trichlorobenzene.

12.2.6.2. The selected samples used for MS, MSD and DUP shall be rotated among client samples at random so that various matrix problems may be noted and/or addressed.

12.2.6.3. Control limits for MBLK, MS, MSD, DUP and LCS are established at least once every year using the average limits of previous MBLK, MS, MSD, DUP and LCS from all instruments. These can be updated in the LIMS. Only the Quality Control Officer or the Semi-Volatiles Manager can update these limits.

12.2.6.4. The LCS and LCSD also have criteria for % recovery. Standard HEAL criteria for passing are +/- 30 % of expected recovery. Again, this is based on an average of the 11 spiking compounds listed in 8270C. They are typically between 70-130 percent of the amount shot

on the instrument. The QA Officer or the Semi-VOA Manager can update these in the LIMS.

12.2.6.5. For tests that have not been control charted the RPD limits have been defaulted to 20 for the LCSD and MSD.

12.2.6.6. The current control charted limits are listed in the LIMS under test information and can be reviewed at any time by trained analysts and data reviewers. The formulas and procedure used to generate control chart limits are available in the QAP.

12.3. To establish the ability to generate acceptable accuracy and precision, the analyst must perform the following operations.

12.3.1. Four Blank Spike/Blank Spike Duplicates are analyzed according to the method.

12.3.2. Calculate the average recovery (x) in µg/L, and the standard deviation of the recovery in µg/L, for each analyte using the results.

12.3.3. DOCs are being evaluated against control limits that are generated by the LIMS.

12.3.4. When one or more of the analytes tested are not within these limits, the analyst must proceed according to Sec. 12.5

12.3.5. Beginning with Sec. 12.5.1, repeat the test only for those analytes that are not comparable. Repeated failure, however, will confirm a general problem with the measurement system. If this occurs, locate and correct the source of the problem and repeat the test for all compounds of interest beginning with Sec. 12.5.1.

12.3.6. All analysts must meet an annual demonstration of continued proficiency: See the QAP for complete details.

12.4. For aqueous and soil matrices, laboratory established surrogate control limits should be compared with the control limits which are located in the LIMS.

12.5. If recovery is not within limits, the following procedures are required.

12.5.1. Check to be sure that there are no errors in the calculations, surrogate solutions or internal standards. If errors are found, recalculate the data accordingly.

12.5.2. Check instrument performance. If an instrument performance problem is identified, correct the problem and re-analyze the sample.

12.5.3. If no problem is found, re-extract and re-analyze.

12.5.4. If, upon re-analysis, the recovery is again not within limits, flag the data as "estimated concentration".

12.5.5. At a minimum surrogate recovery limits are updated on a matrix-by matrix basis, annually.

12.6. Detection Levels

12.6.1. Current practices at HEAL define the detection level as the smallest amount that can be detected above the noise in a procedure and within a stated confidence level. We presently utilize an instrument detection level (IDL), a method detection level (MDL) and a practical quantitation limit (PQL). The relationship between these levels is approximately IDL: MDL: PQL = 1:5:10.

The IDL is the constituent concentration that when injected produces a signal sufficiently large that 99% of the trials with that amount will produce a detectable signal. The IDL is used to determine the analyte concentration in an extract needed to produce a signal to permit calculations.

The MDL (see table 2 in Method 8270C) differs from the IDL in that samples containing the constituent of interest are processed through the complete analytical method. The MDL is different than the IDL because of extraction efficiency and extract concentration factors. The MDL is achieved by a well-trained analyst operating a well-calibrated instrument on a non-routine basis (Annually or semi-annually). First add reagent water or the matrix of interest, to make a concentration two to five times higher than the IDL. Prepare and analyze seven replicates of this solution. Include all sample-processing steps in the determination. Calculate the sample standard deviation (s) of the replicate measurements. The method detection limit (MDL) can be calculated according to the formula:

$$MDL = s * t(99\%)$$

t(99%) is the student's t value for the 99% confidence interval. It depends on the number of trials used in calculating the sample standard deviation, so choose the appropriate value according to the number of trials.

Number of Trials	t(99%)
7	3.143
8	2.998
9	2.896
10	2.821

Formulas for the MDL and standard deviation can be found in the HEAL QAP section 9.0.

The PQL is significant because different laboratories will produce different MDLs even though using the same analytical procedures, instruments and sample matrices. The PQL is about five times the MDL and represents a practical and routinely achievable detection level with a relatively good certainty that any reported value is reliable.

When a compound is above the instrument calibration range, there are two options for an analyst. First, reanalyze the sample in order to bring it within the instruments calibration range. Or if there is not enough sample to reanalyze, then the data will be flagged for that compound. An explanation should also be written into the LIMS so the Project Manager can explain the Flagged data to the client.

If mdl's are not performed "J" flags may not be used and HEAL will not report below the PQL of the instrument. Since TCLP samples are never reported below the TCLP limit and the instrument can see far below these limits, mdl's for TCLP will not be performed.

12.7. Corrective actions are to be made for out-of-control data.

12.7.1. The limits that have been defined for data acceptability also form the basis for corrective action initiation. Initiation of corrective action occurs when the data generated from spikes and sample duplicates exceeds the 99 % confidence interval. If corrective action is necessary, the analyst will take the following steps to determine and correct the measurement system deficiency:

12.7.2. Check all calculations and measurements data (standardization, calibration, concentrations, instrument stability, etc.)

12.7.3. Check equipment to ensure that components are functioning properly.

12.7.4. Check reagents (age, preparation, concentrations, etc.)

12.7.5. Assure that proper procedures were followed.

12.7.6. Reanalyze all samples run in conjunction with the duplicate and/or spike.

12.7.7. In the case that a sample is treated differently than is in the documented procedure, a corrective action report will be filed explaining the reason.

12.7.8. If deficiency continues, consult with the appropriate senior staff member.

12.7.9. A second mechanism for initiation of corrective action is that resulting from Quality Assurance activities concerned with performance audits, system audits, inter and intra-laboratory comparison studies. Corrective Actions initiated through this mechanism will be monitored and coordinated by the laboratory QA officer.

13.0 Calculations

13.1. Aqueous samples:

13.1.1. The result from the chromatogram (ug/mL) is multiplied by the final volume (1ml), and then divided by initial volume (500mL) and then multiplied by 1000mL. If a dilution was needed for the sample or specific analyte, multiply this number by the dilution factor. This result is the calculated final result. The result is in ppb (ug/L).

13.1.1.1. Example: 10ug/mL x1ml x 1000mL/ 1L/ 500mL = 20ug/L

13.2. Soil/Chemical waste samples

13.2.1. The result from the chromatogram (ug/mL), is multiplied by the final volume (1mL), and then divided by the initial weight (20g), 30g if using Soxhlet. If a dilution was needed for the sample or an individual analyte, multiply the resulting number by the dilution factor. This result is the calculated final result given in ppm (mg/kg).

13.2.2. Example: 10ug/mL x 1ml / 20g = 0.50 mg/kg

13.3. Estimation of Uncertainty

13.3.1. Estimation of measurement uncertainty is to be conducted on an annual basis.

13.3.2. Collect, at least, 20 of the most recent LCS data points. Calculated the standard deviation (s) and the average of all the readings. Calculate the 95% confidence interval (2 times the standard deviation). Report the estimated uncertainty as +/-2(s). Uncertainty data is kept on file in the method folders in the quality control office.

13.3.3. Calculate standard deviation (s) and 95% confidence interval according to the following formulae:

$$s = \sqrt{\frac{\sum (x - \bar{x})^2}{(n-1)}}$$

Where: s = standard deviation
x = number in series
$\bar{x}$ = calculated mean of series
n = number of samples taken

$$95\%\ confidence = 2 \times s$$

14.0 Pollution Prevention

14.1. The Accelerated Solvent Extractor (ASE) should be used when possible to reduce the amount of solvent used for soil extractions.

15.0 Waste Management

15.1. After analysis, all samples vials (1L Amber bottles, Soil jars etc.) standard vials, solvent waste etc. will be disposed of according to the HEAL Chemical Hygiene Plan.

15.2. As for pollution prevention, HEAL is committed to maintain the utmost care in disposing of our waste. HEAL is certified by the City of Albuquerque once a year on our procedures for waste disposal and pollution prevention. This is detailed in the HEAL Chemical Hygiene Plan.

16.0 References

16.1. U.S. EPA Method SW-846 8270C, Revision 3, December 1996.

16.2. Quality Assurance Plan, Hall Environmental Analysis Laboratory.

16.3. Chemical Hygiene Plan, Hall Environmental Analysis Laboratory.

16.4. Determinative Chromatographic Separations, US EPA 8000C, March, 2003.

16.5. HEAL SOP S-3540C-5: Soxhlet extraction for soil analysis.

16.6. HEAL SOP S-3545: Pressurized Fluid Extraction for soil analysis.

16.7. HEAL SOP S-3510C-5: Separatory funnel liquid/liquid extraction.

16.8. Method 3500B Organic extraction and sample preparation.

16.9. Method 3540C, Soxhlet Extraction.

16.10. Method 3545, Pressurized Fluid Extraction.

16.11. Method 3510C, Separatory funnel liquid/liquid extraction.

16.12. Method 3600C, Cleanup.

16.13. **Table 2** Method 525 tuning criteria

Mass	*Ion Abundance Criteria*
51	10-80% of mass 198
68	< 2% of mass 69
69	present
70	Less than 2% of m/z 69
127	10 to 80% of base peak(198)
197	< 2% of mass 198
198	Base peak or >50% of m/z 442
199	5-9% of mass 198
275	10-60% of mass 198
365	> 1% of mass 198
441	441 Present but less than mass 443
442	> 50% of mass 198
443	15-24% of mass 442

16.14. **Table 3 Estimated Retention Times and corresponding Internal Standard for each compound reported.**

Estimated RT	IS	Compound ID	IS Reference
2.56		N-nitrosodimethylamine	1,4-Dichlorobenzene-d4
2.592		Pyridine	1,4-Dichlorobenzene-d4
3.694		2-Fluorophenol	1,4-Dichlorobenzene-d4
4.747		Phenol-d5	1,4-Dichlorobenzene-d4
4.766		Phenol	1,4-Dichlorobenzene-d4
4.821		Aniline	1,4-Dichlorobenzene-d4
4.876		Bis(2-chloroethyl)ether	1,4-Dichlorobenzene-d4
4.965		2-Chlorophenol	1,4-Dichlorobenzene-d4
5.159		1,3-Dichlorobenzene	1,4-Dichlorobenzene-d4
5.235	X	1,4-Dichlorobenzene-d4	
5.256		1,4-Dichlorobenzene	1,4-Dichlorobenzene-d4
5.42		Benzyl alcohol	1,4-Dichlorobenzene-d4
5.464		1,2-Dichlorobenzene	1,4-Dichlorobenzene-d4
5.566		2-Methylphenol	1,4-Dichlorobenzene-d4
5.581		Bis(2-chloroisopropyl)ether	1,4-Dichlorobenzene-d4
5.767		N-Nitrosodi-n-propylamine	1,4-Dichlorobenzene-d4
5.785		3+4-Methylphenol	1,4-Dichlorobenzene-d4
5.919		Hexachloroethane	1,4-Dichlorobenzene-d4
6.011		Nitrobenzene-d5	1,4-Dichlorobenzene-d4
6.037		Nitrobenzene	1,4-Dichlorobenzene-d4
6.356		Isophorone	1,4-Dichlorobenzene-d4
6.495		2-Nitrophenol	1,4-Dichlorobenzene-d4
6.549		2,4-Dimethylphenol	1,4-Dichlorobenzene-d4
6.722		Benzoic acid	1,4-Dichlorobenzene-d4
6.691		Bis(2-chloroethoxy)methane	1,4-Dichlorobenzene-d4
6.861		2,4-Dichlorophenol	1,4-Dichlorobenzene-d4
6.967		1,2,4-Trichlorobenzene	Naphthalene-d8 (I.S)
7.052	X	Naphthalene-d8 (I.S)	
7.084		Naphthalene	Naphthalene-d8 (I.S)
7.191		4-Chloroaniline	Naphthalene-d8 (I.S)
7.252		Hexachlorobutadiene	Naphthalene-d8 (I.S)
7.961		4-Chloro-3-methylphenol	Naphthalene-d8 (I.S)
8.151		2-Methylnaphthalene	Naphthalene-d8 (I.S)
8.383		Hexachlorocyclopentadiene	Naphthalene-d8 (I.S)
8.605		2,4,6-Trichlorophenol	Naphthalene-d8 (I.S)
8.674		2,4,5-Trichlorophenol	Naphthalene-d8 (I.S)
8.735		2-Fluorobiphenyl	Naphthalene-d8 (I.S)
8.924		2-Chloronaphthalene	Naphthalene-d8 (I.S)
9.126		2-Nitroaniline	Naphthalene-d8 (I.S)
9.392		Dimethyl phthalate	Naphthalene-d8 (I.S)
9.503		2,6-Dinitrotoluene	Naphthalene-d8 (I.S)

9.579		Acenaphthylene	Acenaphthene-d10 (I.S)
9.787		3-Nitroaniline	Acenaphthene-d10 (I.S)
9.804	X	Acenaphthene-d10 (I.S)	
9.855		Acenaphthene	Acenaphthene-d10 (I.S)
9.983		2,4-Dinitrophenol	Acenaphthene-d10 (I.S)
10.133		Dibenzofuran	Acenaphthene-d10 (I.S)
10.14		4-Nitrophenol	Acenaphthene-d10 (I.S)
10.152		2,4-Dinitrotoluene	Acenaphthene-d10 (I.S)
10.521		Diethyl phthalate	Acenaphthene-d10 (I.S)
10.681		Fluorene	Acenaphthene-d10 (I.S)
10.686		4-Chlorophenyl phenyl ether	Acenaphthene-d10 (I.S)
10.771		4-Nitroaniline	Acenaphthene-d10 (I.S)
10.796		4,6-Dinitro-2-methylphenol	Acenaphthene-d10 (I.S)
10.892		N-Nitrosodiphenylamine	Acenaphthene-d10 (I.S)
10.941		Azobenzene	Acenaphthene-d10 (I.S)
11.076		2,4,6-Tribromophenol	Acenaphthene-d10 (I.S)
11.473		4-Bromophenyl phenyl ether	Phenanthrene-d10
11.555		Hexachlorobenzene	Phenanthrene-d10
11.903		Pentachlorophenol	Phenanthrene-d10
12.183	X	Phenanthrene-d10	
12.224		Phenanthrene	Phenanthrene-d10
12.306		Anthracene	Phenanthrene-d10
12.602		Carbazole	Phenanthrene-d10
13.149		Di-n-butyl phthalate	Phenanthrene-d10
14.177		Fluoranthene	Phenanthrene-d10
14.456		Benzidine	Phenanthrene-d10
14.556		Pyrene	Chrysene-d12 I.S
14.832		4-Terphenyl-d14	Chrysene-d12 I.S
15.669		Butyl benzyl phthalate	Chrysene-d12 I.S
16.598		3,3'-Dichlorobenzidine	Chrysene-d12 I.S
16.6		Benz[a]anthracene	Chrysene-d12 I.S
16.619	X	Chrysene-d12 I.S	Chrysene-d12 I.S
16.622		Bis(2-ethylhexyl)phthalate	Perylene-d12 I.S
16.664		Chrysene	Chrysene-d12 I.S
17.683		Di-n-octyl phthalate	Perylene-d12 I.S
18.358		Benzo(b)fluoranthene	Perylene-d12 I.S
18.406		Benzo[k]fluoranthene	Perylene-d12 I.S
18.938		Benzo[a]pyrene	Perylene-d12 I.S
19.048	X	Perylene-d12 I.S	
21.299		Indeno[1,2,3-cd]pyrene	Perylene-d12 I.S
21.341		Dibenz[a,h]anthracene	Perylene-d12 I.S
21.985		Benzo[g,h,i]perylene	Perylene-d12 I.S

Appendix A

8270C
Calibration Summary

Instrument: _____ **Date:** _____

Calibration Name: _____

Analyte	ICV Pass	ICV Fail	Calibration Curve Points Removed Yes	Calibration Curve Points Removed No	Notes
N-nitrosodimethylamine	☐	☐	☐	☐	
Pyridine	☐	☐	☐	☐	
2-Fluorophenol	N/A	N/A	☐	☐	
Phenol-d5	N/A	N/A	☐	☐	
Phenol	☐	☐	☐	☐	
Aniline	☐	☐	☐	☐	
Bis(2-chloroethyl)ether	☐	☐	☐	☐	
2-Chlorophenol	☐	☐	☐	☐	
1,3-Dichlorobenzene	☐	☐	☐	☐	
1,4-Dichlorobenzene	☐	☐	☐	☐	
Benzyl alcohol	☐	☐	☐	☐	
1,2-Dichlorobenzene	☐	☐	☐	☐	
2-Methylphenol	☐	☐	☐	☐	
Bis(2-chloroisopropyl)ether	☐	☐	☐	☐	
3+4-Methylphenol	☐	☐	☐	☐	
N-Nitrosodi-n-propylamine	☐	☐	☐	☐	
Hexachloroethane	☐	☐	☐	☐	
Nitrobenzene-d5	N/A	N/A	☐	☐	
Nitrobenzene	☐	☐	☐	☐	
Isophorone	☐	☐	☐	☐	
2-Nitrophenol	☐	☐	☐	☐	
2,4-Dimethylphenol	☐	☐	☐	☐	
Benzoic acid	☐	☐	☐	☐	
Bis(2-chloroethoxy)methane	☐	☐	☐	☐	
2,4-Dichlorophenol	☐	☐	☐	☐	
1,2,4-Trichlorobenzene	☐	☐	☐	☐	
Naphthalene	☐	☐	☐	☐	
4-Chloroaniline	☐	☐	☐	☐	
Hexachlorobutadiene	☐	☐	☐	☐	
4-Chloro-3-methylphenol	☐	☐	☐	☐	
2-Methylnaphthalene	☐	☐	☐	☐	
1-Methylnaphthalene	☐	☐	☐	☐	
Hexachlorocyclopentadiene	☐	☐	☐	☐	
2,4,6-Trichlorophenol	☐	☐	☐	☐	
2,4,5-Trichlorophenol	☐	☐	☐	☐	
2-Fluorobiphenyl	N/A	N/A	☐	☐	
2-Chloronephthalene	☐	☐	☐	☐	
2-Nitroaniline	☐	☐	☐	☐	
Dimethyl phthalate	☐	☐	☐	☐	
2,6-Dinitrotoluene	☐	☐	☐	☐	
Acenaphthylene	☐	☐	☐	☐	
3-Nitroaniline	☐	☐	☐	☐	
Acenaphthene	☐	☐	☐	☐	
2,4-Dinitrophenol	☐	☐	☐	☐	
4-Nitrophenol	☐	☐	☐	☐	
Dibenzofuran	☐	☐	☐	☐	

Page 1 of 2

8270C
Calibration Summary

Analyte	ICV Pass	ICV Fail	Calibration Curve Points Removed Yes	Calibration Curve Points Removed No	Notes
2,4-Dinitrotoluene	☐	☐	☐	☐	
Diethyl phthalate	☐	☐	☐	☐	
4-Chlorophenyl phenyl ether	☐	☐	☐	☐	
Fluorene	☐	☐	☐	☐	
4-Nitroaniline	☐	☐	☐	☐	
4,6-Dinitro-2-methylphenol	☐	☐	☐	☐	
N-Nitrosodiphenylamine	☐	☐	☐	☐	
Azobenzene	☐	☐	☐	☐	
2,4,6-Tribromophenol	N/A	N/A	☐	☐	
4-Bromophenyl phenyl ether	☐	☐	☐	☐	
Hexachlorobenzene	☐	☐	☐	☐	
Pentachlorophenol	☐	☐	☐	☐	
Anthracene	☐	☐	☐	☐	
Carbazole	☐	☐	☐	☐	
Di-n-butyl phthalate	☐	☐	☐	☐	
Fluoranthene	☐	☐	☐	☐	
Benzidine	☐	☐	☐	☐	
Pyrene	☐	☐	☐	☐	
4-Terphenyl-d14	N/A	N/A	☐	☐	
Butyl benzyl phthalate	☐	☐	☐	☐	
3,3'-Dichlorobenzidine	☐	☐	☐	☐	
Benz[a]anthracene	☐	☐	☐	☐	
Bis(2-ethylhexyl)phthalate	☐	☐	☐	☐	
Chrysene	☐	☐	☐	☐	
Di-n-octyl phthalate	☐	☐	☐	☐	
Benzo(b)fluoranthene	☐	☐	☐	☐	
Benzo[k]fluoranthene	☐	☐	☐	☐	
Benzo[a]pyrene	☐	☐	☐	☐	
Indeno[1,2,3-cd]pyrene	☐	☐	☐	☐	
Dibenz[a,h]anthracene	☐	☐	☐	☐	
Benzo[g,h,i]perylene	☐	☐	☐	☐	

Analyst: _____

QA: _____

Additional Comments: _____

Hall Environmental Analysis Laboratory

Common Name: EPA Method 8260B	S.O.P. Number: S-8260-10
Method: Standard Operating Procedure for the determination of Volatile Organic Compounds by Gas Chromatography/ Mass Spectrometry (GC.MS): Capillary Column Technique	
Effective Date: 26 August 2011	Supersedes: S-8260-9
Laboratory Manager:	Quality Assurance/Quality Control Officer:

1.0 Scope and Application

1.1 Method 8260B is used to determine the concentration of volatile organic compounds in aqueous, non-aqueous, solids, chemical waste and air/emission samples. The following compounds can be determined by this method:

Compound Name	CAS No.[a]	Appropriate Technique Purge And Trap
Acetone	67-64-1	pp
Acetonitrile	75-05-8	pp
Acrolein (Propenal)	107-02-8	pp
Acrylonitrile	107-13-1	pp
Allyl alcohol	107-18-6	ht
Allyl chloride	107-05-1	a
Benzene	71-43-2	a
Benzyl chloride	100-44-7	pp
Bromoacetone	598-31-2	pp
Bromochloromethane	74-97-5	a
Bromodichloromethane	75-27-4	a
4-Bromofluorobenzene (surr.)	460-00-4	a
Bromoform	75-25-2	a
Bromomethane	74-83-9	a
n-Butanol	71-36-3	ht
2-Butanone (MEK)	78-93-3	pp
Carbon Disulfide	75-15-0	pp
Carbon tetrachloride	56-23-5	a
Chloral hydrate	302-17-0	pp

Compound Name	CAS No.[a]	Appropriate Technique Purge And Trap
Chlorobenzene	108-90-7	a
2-Chloro-1,3-butadiene	126-99-8	a
Chlorodibromomethane	124-48-1	a
Chloroethane	75-00-3	a
2-Chloroethanol	107-07-03	pp
bis-(2-Chloroethyl)sulfide	505-60-2	pp
2-Chloroethylvinyl ether	110-75-8	a
Chloroform	67-66-3	a
Chloromethane	74-87-3	a
Chloroprene	126-99-8	a
3-Chloroproprene	107-05-1	a
3-Chloropropionitrile	542-76-7	i
1,2-Dibromo-3-chloropropane	96-12-8	pp
1,2-Dibromoethane	106-93-4	a
Dibromomethane	74-95-3	a
1,2-Dichlorobenzene	95-50-1	a
1,3-Dichlorobenzene	541-73-1	a
1,4-Dichlorobenzene	106-46-7	a
cis-1,4-Dichloro-2-butene	1476-11-5	a
trans-1,4-Dichloro-2-butene	110-57-6	pp
Dichlorodifluoromethane	75-71-8	a
1,1-Dichloroethane	75-34-3	a
1,2-Dichloroethane	107-06-2	a
1,1-Dichloroethene	75-35-4	a
cis-1,2-Dichloroethene	156-59-2	a
trans-1,2-Dichloroethene	156-60-5	a
1,2-Di chloropropane	78-87-5	a
1,3-Dichloro-2-propanol	96-23-1	pp
cis-1,3-Dichloropropene	10061-01-5	a
trans-1,3-Dichloropropene	10061-02-6	a
1,2,3,4-Diepoxybutane	1464-53-5	a
Diethyl ether	60-29-7	a
1,4-Dioxane	123-91-1	pp
Epichlorohydrin	106-89-8	i
Ethanol	64-17-5	i
Ethyl acetate	141-78-6	i
Ethylbenzene	100-41-4	a
Ethylene oxide	75-21-8	pp
Ethyl methacrylate	97-63-2	a
Hexachlorobutadiene	87-68-3	a
Hexachloroethane	67-72-1	i
2-Hexanone	591-78-6	pp
Iodomethane	74-88-4	a
Isobutyl alcohol	78-83-1	pp
Isopropylbenzene	98-82-8	a
Malononitrile	109-77-3	pp
Methacrylonitrile	126-98-7	pp
Methanol	67-56-1	i

Compound Name	CAS No.[a]	Appropriate Technique Purge And Trap

Methylene chloride	75-09-2	a
Methyl methacrylate	80-62-6	a
4-Methyl-2-pentanone (MIBK)	108-10-1	pp
Naphthalene	91-20-3	a
Nitrobenzene	98-95-3	a
2-Nitropropane	79-46-9	a
Pentachloroethane	76-01-7	i
2-Picoline	109-06-8	pp
Propionitrile (ethyl cyanide)	107-12-0	ht
n-Propylamine	107-10-8	a
Pyridine	110-86-1	i
Styrene	100-42-5	a
1,1,1,2-Tetrachloroethane	630-20-6	a
1,1,2,2-Tetrachloroethane	79-34-5	a
Tetrachloroethene	127-18-4	a
Toluene	108-88-3	a
1,2,4-Trichlorobenzene	120-82-1	a
1,1,1-Trichloroethane	71-55-6	a
1,1,2-Trichloroethane	79-00-5	a
Trichloroethene	79-01-6	a
Trichlorofluoromethane	75-69-4	a
1,2,3-Trichloropropane	96-18-4	a
Vinyl Acetate	108-05-4	a
Vinyl Chloride	75-01-4	a
m-Xylene	95-47-6	a
p-Xylene	108-38-3	a
o-Xylene	106-42-3	a
1,3,5-Trimethylbenzene	108-64-8	a
1,2,4-Trimethylbenzene	95-63-6	a

a indicates adequate response using this technique.
ht indicates method analyte only when purged at 80°C.
i indicates inappropriate technique for this analyte.
pp indicates poor purging efficiency, resulting in high EQLs.
surr surrogate
*This compound list is from the method, some analytes may still be analyzed and reported using this SOP even when the list marks them as "ht".

1.2 Method 8260B can be used to quantitate most volatile organic compounds that have boiling points below 200° C and that are insoluble or slightly soluble in water. Volatile water-soluble compounds can be included in this analytical technique. However, for the more soluble compounds, quantitation limits are approximately ten times higher because of poor purging efficiency. Such compounds include low-molecular-weight halogenated hydrocarbons, aromatics, ketones, nitriles, acetates, acrylates, ethers, and sulfides. See Table 4 for a list of compounds and their characteristic ions that have been evaluated on the purge-and-trap GC/MS system.

The following analytes are also amenable to analysis by Method 8260:

Bromobenzene	1-Chlorohexane
n-Butylbenzene	2-Chlorotoluene
sec-Butylbenzene	4-Chlorotoluene
tert-Butylbenzene	Chloroacetonitrile
1-Chlorobutane	cis-1,2-Dichloroethene
1,3-Dichloropropane	Methyl-tert-butyl ether (MTBE)
2,2-Dichloropropane	Pentafluorobenzene
1,1-Dichloropropane	n-Propylbenzene
p-Isopropyltoluene	1,2,3-Trichlorobenzene
Methyl acrylate	1,2,4-Trimethylbenzene
1,3,5-Trimethylbenzene	

1.3 The practical quantitation limit (PQL) of Method 8260 for determining an individual compound is somewhat instrument dependent. Low limits may be achieved using an ion trap mass spectrometer. No matter which instrument is used, PQLs will be proportionately higher for sample extracts and samples that require dilution or reduced sample size to avoid saturation of the detector. Current PQL for 8260 analytes reported by Hall Environmental can be found in the LIMS.

1.4 Method 8260 is based upon a purge-and-trap, gas chromatographic/mass spectrometric (GC/MS) procedure. This method is restricted to use by, or under the supervision of analysts experienced in the use of purge-and-trap systems and gas chromatography/mass spectrometers, and skilled in the interpretation of mass spectra and their use as a quantitative tool. Each analyst must demonstrate the ability to generate acceptable results with this method. Each analyst must establish a Demonstration of Capability (DOC) in accordance with the HEAL QA Plan prior to the analysis of client samples.

2.0 Summary of Method

2.1 The volatile compounds are introduced into the gas chromatograph by the current S-5030B. Purged sample components are trapped in a Vocarb 3000 trap. When purging is complete, the trap is heated and back-flushed with helium to desorb the sample components. The analytes are desorbed directly to a capillary column for analysis. The column is temperature programmed to separate the analytes which are then detected with a mass spectrometer (MS) interfaced to the gas chromatograph (GC).

2.2 For solids and/or soils, a portion of the sample is dispersed in organic free methanol to dissolve the volatile organic constituents in accordance with the current S-5035. A portion of the methanol is combined with organic-free reagent water in the autosampler vial. It is then analyzed by purge-and-trap GC/MS in accordance with the current S-5030B.

2.3 Analytes eluted from the capillary column are introduced directly into the mass spectrometer. Identification of target analytes is accomplished by comparing their mass spectra with the electron impact (or electron impact-like) spectra of authentic standards. Quantitation is done by comparing the response of a base (quantitation) ion relative to an internal standard with a minimum five-point calibration curve.

3.0 Safety

3.1 Some of the compounds that are used are known carcinogens, mutagens, and or teratogens. So the utmost care must be used in handling the neat Volatile compounds and the mixes made from them. At a minimum, latex gloves, a lab coat, and safety glasses must be worn to avoid any unnecessary contact. Any handling of the open containers of neat compounds must be done in a fume hood to avoid unnecessary exposure.

3.1.1 Working standards also need to be handled with care. When handling them you need at a minimum to wear latex gloves, safety glasses and a lab coat.

3.2 Latex gloves, lab coat, and safety glasses must be worn when handling solvents. Extractions can only be done if more than one person is in the same area as the person doing the extract.

4.0 Definitions

4.1 Continuing Calibration Verification (CCV) – standard from the same source as the curve used to check the stability of the instrument.
4.2 Laboratory Control Spike (LCS) – laboratory fortified blank at 20ppb.
4.3 Laboratory Control Spike Duplicate (LCSD) – laboratory fortified blank duplicate at 20ppb.
4.4 Initial Calibration Verification (ICV) – second source standard used to verify the calibration at 20ppb.
4.5 Matrix Spike (MS) – laboratory fortified sample at 20ppb.
4.6 Matrix Spike Duplicate (MSD) – laboratory fortified sample duplicate at 20ppb.
4.7 Method/ Reagent blank (MB or RB) – analyte-free matrix that is similar in nature to samples and analyzed in the same manner as a sample.
4.8 Method Detection Limit (MDL) – the constituent concentration that when processed through the complete method, produces a signal with a 99% probability that it is different than the blank.
4.9 Opening Standard – standard from the same source as the curve run at the beginning of a particular analysis day or batch run that is used to check the stability of the instrument. Same as the CCV.
4.10 Practical Quantitation Limit (PQL) - represents the lowest concentration to practically and routinely report.
4.11 Instrument Detection Level (IDL) - The IDL is the constituent concentration that when injected produces a signal sufficiently large that 99% of the trials with that amount will produce a detectable signal. The IDL is used to determine the analyte concentration in an extract needed to produce a signal to permit calculations.
4.12 Sample Duplicate (DUP) – duplicate of sample.

4.13 Surrogate – a particular compound that is used to determine the overall efficiency of the extraction process.
4.14 Internal Standard – a compound that is like, but different from analyte(s) that is added to all injections. Signal from analyte is compared with signal from the internal standard to find out how much analyte is present.
4.15 GC – gas chromatograph.
4.16 MS- mass spectrometer
4.17 Volatile Deli – sample storage refrigerator located in the Volatile laboratory along the west wall.
4.18 Mid-range concentration – considered to be 20ppb.
4.19 CCC – Calibration Check Compound. Compound that must be analyzed and recover within ±20% of expected concentration.
4.20 SPCC – System Performance Check Compound. Compound that must be analyzed and meet the minimum response factor.
4.21 TB - Trip/travel blank. QC sample that travels from lab to field and back to lab to help in assessment of possible sources of contamination.
4.22 r^2 – The square of the sample correlation coefficient between the outcomes and their predicted values, or in the case of simple linear regression, between the outcome and the values being used for prediction. In such cases, the values vary from 0 to 1.
4.23 Correlation Coefficient – Statistical measure of the interdependence or two or more random variables. The value indicates how much change in one variable is explained by a change in another.
4.24 VSB – Volatile Storage Blank. QC sample that is used to assess sample storage area is free from contamination.

5.0 Interferences

5.1 Major contaminant sources are volatile materials in the laboratory and impurities in the inert purging gas and in the sorbent trap. The use of non-polytetrafluoroethylene (PTFE) thread sealants, plastic tubing, or flow controllers with rubber components should be avoided since such materials out-gas organic compounds which will be concentrated onto the trap during the purge operation. Routine analyses of MB's provide valuable information about the presence of contaminants.

5.2 Interfering contamination may occur when a sample containing low concentrations of volatile organic compounds is analyzed immediately after a sample containing high concentrations of volatile organic compounds. After analysis of a sample containing high concentrations of volatile organic compounds, one or more MB should be analyzed to check for cross contamination. For samples containing large amounts of water soluble materials, suspended solids, high boiling point compounds or high concentrations of compounds being determined, it may be necessary to wash the purging device, rinse it with organic-free reagent water, and then dry the purging device in an oven at 105°C. In extreme situations, the whole purge and trap device may require dismantling and cleaning.

5.2.1 It is important to not introduce surfactants to the sparge vessel. Avoid soap solutions to clean the sparge vessel.

5.3 Samples can be contaminated by diffusion of volatile organics (particularly fluorocarbons) through the septum seal into the sample during shipment and storage. A trip blank prepared from organic-free reagent water and carried through the sampling and handling protocol serves as a check on such contamination.

5.4 When petroleum samples are analyzed, some chromatographic peaks will elute after the target analytes. The oven temperature program includes a post-target analysis bake out period to ensure that semi-volatile hydrocarbons are volatilized.

5.5 Care must be given to avoid laboratory contamination. Methylene chloride is a common laboratory solvent which can be a source of contamination for volatile organic analysis. All volatile analysis and sample storage must take place in an area completely free of any solvents or fuels which may be potential candidates for determination by using this method. Any analyst that has come in contact with any potential target solvents must make certain that they spend a minimum of 5 minutes away from the Volatile laboratory.

6.0 Apparatus and Reagents

6.1 Purge and Trap concentrator as described in current 5030 SOP.

6.2 GC/MS systems:

6.2.1 VAL

6.2.1.1 Varian CP3800 GC, equipped with a Varian 1177 injection port

6.2.1.2 Varian Saturn 2000 Mass Spectrometer

6.2.1.3 Varian Archon Autosampler

6.2.1.4 Tekmar Stratum Purge and Trap concentrator

6.2.1.5 MS Workstation Ver. 6.9 data system

6.2.1.6 DB-VRX, 30m x0.25mm ID with 1.2um film thickness (or equivalent).

6.2.2 THOR

6.2.2.1 Varian CP3800, equipped with a Varian 1079 injection port.

6.2.2.2 Varian Saturn 2000 Mass Spectrometer.

6.2.2.3 Varian Archon Autosampler.
6.2.2.4 Tekmar 3000 Purge and Trap concentrator.

6.2.2.5 MS Workstation Ver. 6.9 data system.

6.2.2.6 DB-VRX, 30m x0.25mm ID with 1.2um film thickness (or equivalent).

6.2.3 NEPTUNE

6.2.3.1 Varian CP3900, equipped with a Varian 1177 injection port.

6.2.3.2 Varian Saturn 2100T Mass Spectrometer.

6.2.3.3 Varian Archon Autosampler.

6.2.3.4 Tekmar 3100 Purge and Trap concentrator.

6.2.3.5 MS Workstation Ver. 6.9 data system.

6.2.3.6 DB-VRX, 30m x0.25mm ID with 1.2um film thickness (or equivalent).

6.3 Syringes: 5-mL gas tight Luerlok glass syringe, 50-mL gas tight Luerlok glass syringe, 100-mL gas tight Luerlok glass syringe.

6.4 Micro syringes: 1.0 - 1000 µL. (Class A)

6.5 Analytical balance – capable of weighing to within 0.0001 g.

6.6 Volumetric flask: Class A 1-100mL

6.7 Glass scintillation vials – 20mL, with Teflon lined screw-caps.

6.8 Disposable pipettes – Pasteur.

6.9 Spatula – Stainless steel.

6.10 Organic-free reagent water - All references to water in this method refer to organic-free reagent water.

6.11 Methanol (MeOH) - Purge and trap grade.

6.12 Stock standard solutions - Stock solutions may be prepared from pure standard materials or purchased as certified solutions. Stock standards are usually at a concentration of 2000ppm.

6.13 Working standards. Using stock standard solutions, prepare working standards in methanol, as needed, containing the compounds of interest. The working standards should be prepared at concentrations such that the aqueous calibration standards prepared in Sec. 9.3.1 will bracket the

concentrations found in real samples or should define the working range of the GC/MS system. Each standard contains all analytes for detection by this method that are reported by HEAL. Working standards should be stored with minimal headspace. The typical concentration of a working standard is 200ppm.

6.14 Surrogate standards. All injections done in accordance with this SOP must have the necessary surrogates included. See Sec. 9.2.1 for instruction on surrogate preparation.

6.15 Spiking standards (ICV/LCS). Spiking standards are prepared to include compounds that are representative of compounds which are being investigated. At a minimum the spiking standards must include 1,1-dichloroethene, benzene, trichloroethene, toluene, and chlorobenzene.

6.15.1 The LCS is used when spiking the minimum number of required compounds.

6.15.2 The ICV is used when spiking additional compounds which are project specific or permit mandated compounds

6.15.3 The spiking standards are prepared at a concentration of 20ppb.

6.15.4 All analytes must be included in a spiking standard at least once every two years.

6.16 Narrow range pH strips. Used to check the preservation pH of 1:1HCl preserved samples. See Section 12.8.10 for a discussion of usage.

7.0 Sample Collection, Preservation, Handling and Storage

7.1 All samples for analysis using this SOP must be collected in the appropriate sample container. Aqueous samples must be collected in a 44mL VOA vial. Non-aqueous samples should be collected in accordance with the current S-5035. Ideally, no sample containers should have headspace. If sample containers have headspace this is notated on sample chain of custody or sample checklist.

7.2 All samples should be kept above freezing and <6^{0}C from time of collection to just prior to analysis.

7.3 All aqueous samples are preserved with either $HgCl_2$ or 1:1HCL to maintain a sample pH <2. Samples preserved with 1:1HCL are checked for preservation after sample analysis. There is no preservation check for samples preserved with $HgCl_2$.

7.4 All samples requiring analysis using this SOP must be stored in the Volatile Deli immediately after sample receipt. Samples must remain in the Volatile Deli until all associated volatile analyses are complete.

7.5 EPA mandates the analysis of samples, using Method 8260B, must be within 14 days of collection.

7.6 All air samples are collected in 1L Tedlar bags. The air samples are analyzed as soon as possible.

8.0 Operating Conditions

8.1 Operating parameters for the purge and trap concentrators are included in the current S-5030B.

8.2 Gas Chromatograph.

8.2.1 General:

8.2.1.1 Injector temperature: 200^0-260^0C

8.2.1.2 GC to MS transfer line temperature: 200^0-260^0C

8.2.2 Column parameters:

8.2.2.1 Column flow: 1.0 to 1.5mL/min.

8.2.2.2 Split ratio: instrument dependent

8.2.2.3 Initial temperature: 40 to 60^0C

8.2.2.4 Temperature program: instrument and column dependent.

8.2.2.5 Final temperature: instrument and column dependent.

8.3 Mass Spectrometer

8.3.1 Mass range: 35 to 270 m/z

8.3.2 Scan time: 0.7 sec/scan

8.3.3 Ion trap temperature: 150^0C

9.0 Calibration Information

9.1 Preparation of fresh working standards may be necessary when a comparison of CCV's with the calibration curve indicates a problem. The CCV's are prepared at concentrations such that they will bracket the concentrations found in real samples, defining the working range of the GC. Each standard will contain each analyte for detection by this method that is reported by HEAL. Working standards must be stored with minimal headspace in the freezer and checked frequently for signs of degradation or evaporation.

9.2 Surrogate and internal standards - The analyst monitors the performance of the analytical system and the effectiveness of the method in dealing with each sample matrix by spiking each sample, standard, and reagent water blank with surrogate halocarbons. The combination of toluene-d8, 4-bromofluorobenzene, 1,2-dichloroethane-d4, and dibromofluoromethane is used to encompass the range of the temperature program used in this method. The internal standards are pentafluorobenzene, chlorobenzene-d5, and 1,4-dichlorobenzene-d4. Stock standard solutions are purchased, at a concentration of 2000 µg/mL for surrogates and 2000µg/mL for Internal Standards, in methanol. A mix prepared at 50µg/mL of this surrogate spiking solution is made approximately every other day in one of the standard reservoirs on the Archon. A 1 µL injection is performed by the Archon autosampler into every sample and reference standard analyzed.

9.2.1 To prepare surrogate/ internal standard mix place the 3040uL of methanol into the clean Archon standard reservoir. Add 40uL of the 2000ppm surrogate standard to the methanol below the surface. Add 80uL of the 2000ppm internal standard to the methanol below the surface. This preparation yields a surrogate concentration of 25ppm and internal standard concentration of 50ppm.

9.3 Calibration standards - Prepare calibration standards in organic-free reagent water from the secondary dilution of the stock standards, at a minimum of five concentrations. One of the concentrations will be at a concentration at or below the PQL. The concentrations define the working range of the GCMS, which is typically from 2.0ng up to 500ng on the column (0.4ppb to 100ppb). Each standard contains, at a minimum, all the analytes to be reported. The laboratory does not report a quantitative result for a target analyte that was not included in the calibration standards.

(Raw ng on column / mL purged) = ppb

9.4 Second source confirmation – A certified (true) second source from the same manufacturer or one from a different manufacturer than that used for calibration that is used to validate the initial calibration. This second source standard (ICV) is prepared the same way a CCV is and analyzed every time a new curve is analyzed and prior to sample analysis.

9.5 All analysts receive training of standard preparation prior to being able to complete work unsupervised. This training will included all steps from beginning in LIMS with a unique identifier to bench preparation, storage, and technique.

9.6 Each GC/MS system is hardware-tuned to meet the criteria in Table 3 for a 50ng injection or purging of 4-bromofluorobenzene. Analyses must not begin until these criteria are met.

9.7 A set of at least five calibration standards, at varying concentrations, containing the method analytes is needed. One calibration standard contains each analyte at a concentration at or below the laboratory practical quantitation level (PQL) for that compound; the other calibration standards contain analytes at concentrations that define the working range of the GCMS. Calibration is done using the sample introduction technique discussed in the current SOP for 5030.

9.8 Tabulate the area response of the characteristic ions (see Table 4) against concentration for each compound and each internal standard. Calculate response factors (RF) for each compound relative to the nearest internal standard. Table 1 shows each compound with the corresponding internal standard. The RF is calculated as follows:

$$RF = (A_x C_{is}) / (A_{is} C_x)$$

where:

A_x = Area of the characteristic ion for the compound being measured.
A_{is} = Area of the characteristic ion for the specific internal standard.
C_{is} = Concentration of the specific internal standard (ng/μL).
C_x = Concentration of the compound being measured (ng/μL).

9.9 The average RF is calculated and recorded for each compound using the minimum five RF values calculated for each compound from the initial minimum (5-point) calibration curve. A system performance check is performed to ensure that minimum average RFs are met before the calibration curve is used. Five compounds (the System Performance Check Compounds, or SPCCs) are checked for a minimum average relative response factor. These compounds are chloromethane; 1,1-dichloroethane; bromoform; 1,1,2,2-tetrachloroethane; and chlorobenzene. These compounds are used to check compound instability and to check for degradation caused by contaminated lines or active sites in the system. Examples of these occurrences are:

9.9.1 Chloromethane - This compound is the most likely compound to be lost if the purge flow is too fast.

9.9.2 Bromoform - This compound is one of the compounds most likely to be purged very poorly if the purge flow is too slow. Cold spots and/or active sites in the transfer lines may adversely affect response. Response of the quantitation ion (m/z 173) is directly affected by the tuning of BFB at ions m/z 174/176. Increasing the m/z 174/176 ratio relative to m/z 95 may improve bromoform response.

9.9.3 Tetrachloroethane and 1,1-dichloroethane - These compounds are degraded by contaminated transfer lines in purge-and-trap systems and or active sites in trapping materials.

9.10 Using the RFs from the initial calibration calculate and record the percent relative standard deviation (%RSD) for all compounds. The percent RSD is calculated as follows:

$$\%RSD = SD / avgRF * 100$$

where:

RSD = relative standard deviation.
avgRF = mean of the 5 initial RFs for a compound.
SD = standard deviation of average RFs for a compound.

$$SD = \sqrt{\sum_{i=1} (RF_i - RF)^2 / n-1}$$

where:

Rf_i = RF for each of the minimum of 5 calibration levels.
N = number of RF values (i.e., 5)

The percent relative standard deviation should be less than 15% for each compound when using an average response factor calibration. However, the %RSD for each individual Calibration Check Compound (CCC's) must be less than 30%. The CCC's are:

1,1-Dichloroethene,
Chloroform,
1,2-Dichloropropane,
Toluene,
Ethylbenzene, and
Vinyl chloride.

9.10.1 If the %RSD of any of CCC is 30% or greater, then corrective action to eliminate a system leak and/or column reactive sites is required before re-attempting calibration.

9.11 Linearity - If the %RSD of any compound is 15% or less, then the relative response factor is assumed to be consistent over the calibration range, and the average response factor may be used for quantitation.

9.11.1 If the %RSD of any compound is greater than 15%, construct calibration curves of area ratio (A/A_{is}) versus concentration using first order regression fit of the five calibration points.

9.11.2 To use an average response factor calibration the software is adjusted to derive correct calculations. First the %RSD for the compound must be ≤15%. Select force through zero and change the weighting option to $1/x^2$. While this calibration scheme is not currently employed, it is available for use. Prior to using this scheme for quantitation of client samples, the scheme must be shown valid by either hand or spreadsheet calculation for each instrument where it is employed. This must be done only once. The data collected from this calculation will be kept on file in the QAO office in the method file.

9.12 The use of calibration curves is a recommended alternative to average response factor calibration, and a useful diagnostic of standard preparation accuracy and absorption activity in the chromatographic system.

9.12.1 Use of a linear regression calibration is typically used. Each calibration must consist of a minimum of 5 points for each analyte. The r2 value must be >0.99. The CCC compounds must still have a %RSD ≤30%.

9.12.2 The analyst should select the regression order which introduces the least error into the quantitation. A 2nd order linear regression requires a minimum of 6 calibration levels. The use of a 3rd order linear regression is prohibited.

9.12.3 See EPA Method 8000B Section 11.5.2 for a discussion of linear regression fits and equations.

9.13 If a calibration model shows that a particular concentration level does not fit, the analyst should follow the guidelines set forth in Method 8000B Section 11.5.5.2.

9.13.1 Remove the calibration point in its entirety only if it is at an extreme end of the curve; i.e. the lowest or highest most concentration(s). The analyst must maintain the specified minimum calibration levels for the desired fit and can not report above the highest level being used.

9.13.2 Re-analyze the point in duplicate and replace. The duplicate must recover within the stipulated % drift allowances stipulated for the CCV and ICV.

9.13.3 Re-analyze the calibration in its entirety.

9.14 The calibration curve is verified each 12 hour shift by a calibration standard (CCV). Recalibration is required if calibration and on-going performance criteria cannot be met.

10.0 Extractions/Final Volumes.

10.1 There are two SOP's that contain all the information for method 8260 extractions.

10.1.1 The extraction criteria for non-aqueous samples are contained in the current S-5035.

10.1.2 The TCLP extraction criteria are contained in the current S-TCLP.

10.2 All non-aqueous samples are extracted in accordance with the current 5035SOP and analyzed in accordance with the current 5030SOP.

10.3 All aqueous samples are introduced to the GCMS according to the current 5030SOP

10.4 All air samples collected in Tedlar bags are introduced to the GCMS according to the current 5030SOP.

10.5 See Appendix 1 at the end of this document for the procedure that details the analysis of 8260 analytes in soil using the 5035B low level method.

11.0 Analysis: Verification, Procedure and Interpretation

11.1 GC/MS calibration verification.

11.1.1 Prior to analysis of samples, purge 50ng of 4-bromofluorobenzene standard following the current 5030SOP. The resultant mass spectra for the BFB must meet all the criteria given in Table 3. These criteria must be met during each 12 hour shift.

11.1.2 The initial calibration curve (Sec. 9.0) for each compound of interest must be checked and verified once every 12 hours during analysis. This is accomplished by analyzing a calibration standard (CCV) that is at concentrations near the midpoint concentration for the working range of the GC/MS and then checking the SPCCs and CCCs. The concentration of the CCV must vary with each 12 hour shift.

11.1.3 System Performance Check Compounds (SPCCs): A system performance check must be performed every 12 hour shift. If the SPCC criteria are met, a comparison of relative response factors is made for all compounds. This is the same check that is applied during the initial calibration. If the minimum relative response factors are not met, the system must be evaluated, and corrective action must be taken before sample analysis begins. Some possible problems are standard mixture degradation, injection port inlet contamination, contamination at the front end of the analytical column, and active sites in the column or chromatographic system. This check must be met before analysis begins.

11.1.3.1 The minimum relative response factor for volatile SPCCs are as follows:

Chloromethane	0.10
1,1-Dichloroethane	0.10
Bromoform	>0.10
Chlorobenzene	0.30
1,1,2,2-Tetrachloroethane	0.30

11.1.4 Calibration Check Compounds (CCC's) - After the system performance check is met, CCC's listed in (sec. 9.10) are used to check the validity of the initial calibration.

Calculate the percent drift using:

$$\% \text{ Drift} = \{(C_1 - C_c) / C_1\} * 100$$

where:

C_1 = Calibration Check Compound standard concentration.

C_c = Measured concentration used in selected quantitation method.

If the percent difference for each CCC is less than or equal to 20%, the initial calibration is assumed to be valid. If the criterion is not met (>20% drift) for any one CCC, corrective action must be taken. Problems similar to those listed under SPCCs could affect this criterion. If no source of the problem can be determined after corrective action has been taken, a new five-point calibration must be generated. This criterion must be met before sample analysis begins.

11.1.4.1 The acceptable (general) % drift criterion for all non-CCC target analytes is less than or equal to 30%. Acetone, Carbon Disulfide, 2-Butanone, 2-Hexanone, 4-Methyl-2-pentanone, Isopropyl Alcohol, and Isobutyl Alcohol acceptable drift is ±40%. Non typical analytes that are requested under LIMS test code 8260LF and 8260T2 have acceptable drift of ±40%. 2-Methylnaphthalene and 1-methylnaphthalene are not method analytes and have an acceptable drift of ±40%. Persistent, repeat failures of these analytes should be documented in a CAR.

11.1.4.2 It is suggested that a working standard be freshly prepared when target analytes exceed 20% drift. If a working standard fails any CCC compound, it must be reanalyzed. Repeat failure of the CCC indicates the need for possible maintenance and recalibration.

11.1.5 The internal standard responses and retention times in the calibration check standard must be evaluated immediately after or during data acquisition. If the retention time for any internal standard changes by more than 30 seconds from the last calibration check(12 hours), the chromatographic system must be inspected for malfunctions and corrections must be made, as required. If the area for any of the internal standards changes by a factor of two (-50% to +100%) from the last daily CCV, the mass spectrometer must be inspected for malfunctions and corrections must be made, as appropriate. When corrections are made, reanalysis of samples analyzed while the system was malfunctioning is required.

11.2 GC/MS analysis

11.2.1 Historical data should be checked when available. This will minimize contamination of the GC/MS system from unexpectedly high concentrations of organic compounds. Use of screening is particularly important since this method is used to achieve low detection levels.

11.2.1.1 Historical data should only be used as a tool, not as a means to circumvent any policy set forth in any US EPA methodology or the Hall Environmetnal Analysis Laboratory QA Plan.

11.2.2 All samples and standard solutions, prepared for the autosampler, must be allowed to warm to ambient temperature before analysis.

11.2.3 BFB tuning criteria and GC/MS calibration verification criteria must be met before analyzing samples.

11.2.4 The process of the Archon taking an aliquot destroys the validity of aqueous samples for future analysis; therefore, if there is only one VOA vial, the analyst should prepare a second aliquot for analysis, filling one 20mL VOA vial before analysis to protect against possible loss of sample integrity. This second sample is maintained only until such time when the analyst has determined that the first sample has been analyzed properly. If there is not enough sample left to fill a 20mL vial, an aliquot can be saved in another 5mL syringe, if analysis is needed from the back up syringe, it must be analyzed as soon as possible not to exceed 6 hours. Care must be taken to prevent air from leaking into the syringe.

11.2.4.1 The following procedure is appropriate for diluting aqueous purgeable samples. All steps must be performed without delays until the diluted sample is in a headspace free vial.

11.2.4.1.1 Dilutions are injected by the Archon or made by the analyst in a 44mL headspace free vial. No dilution greater than 100x should be done using the Archon.

11.2.4.1.2 Calculate the volume of sample to be added to the 44mL headspace free vial.

11.2.4.1.3 Inject the proper aliquot of sample from a selected microsyringe into the 44mL headspace free vial. Repeat above procedure for additional dilutions.

11.2.5 The Archon adds 1 µL of surrogate/internal standard spiking solution to each sample. The addition of 1 µL of the surrogate/internal standard spiking solution to 5mL of sample is equivalent to a concentration of 10 µg/L of each surrogate standard. The addition of 1µL of the surrogate/internal standard spiking solution to 100µL of sample is equivalent to a concentration of 0.5mg/kg of each surrogate standard.

11.2.6 Purge-and-trap by current S-5030. If the initial analysis of sample or a dilution of the sample has a concentration of analytes that exceeds the calibration range, the sample must be reanalyzed at a higher dilution. Secondary ion quantitation is allowed when there are sample interferences with the primary ion. When a sample is analyzed that has saturated ions from a compound, this analysis must be followed by a blank organic-free reagent water analysis. If the blank analysis is not free of interferences, the system must be decontaminated. Sample analysis may not resume until the blank analysis is demonstrated to be free of interferences.

11.2.6.1 All dilutions should keep the response of the major constituents (previously saturated peaks) in the upper half of the linear range of the curve if possible. Proceed to (Secs. 11.3.1 and 11.3.2) for qualitative and quantitative analysis.

11.2.7 Tedlar bag air analysis is done by carefully placing 50mL of the sample into a gastight Luer-Lock syringe. This sample portion is then injected into the sparge vessel on the concentrator

and analyzed in the same manner as an aqueous sample. To achieve lower detection levels, increase the sample volume being injected into the sparge vessel.

11.3 Data interpretation

11.3.1 Qualitative analysis

11.3.1.1 The qualitative identification of compounds determined by this method is based on retention time, and on comparison of the sample mass spectrum, after background correction, with characteristic ions in a reference mass spectrum. The reference mass spectrum is generated by using the conditions described in this method. The characteristic ions from the reference mass spectrum are defined to be the three ions of greatest relative intensity, or any ions over 30% relative intensity if less than three such ions occur in the reference spectrum. Compounds are identified as present when the criteria below are met.

11.3.1.1.1 The intensities of the characteristic ions of a compound maximize in the same scan or within one scan of each other. Selection of a peak by a data system target compound search routine where the search is based on the presence of a target chromatographic peak containing ions specific for the target compound at a compound-specific retention time will be accepted as meeting this criteria.

11.3.1.1.2 The relative intensities of the characteristic ions agree within 30% of the relative intensities of these ions in the reference spectrum. (Example: For an ion with an abundance of 50% in the reference spectrum, the corresponding abundance in the sample spectrum can range between 20% and 80%.)

11.3.1.1.2 Some analytes may not contain qual ions which are not 30% or greater in intensity. In such instances, only qual ions that are at a minimum of 10% intentsity should be used.

11.3.1.1.3 Some analytes may only have one qual ion which is also the quan ion. An example of this would be dichlorodifluoromethane. In such a case, selection of the quan ion as the qual ion is the only solution.

11.3.1.1.4 Should a sample contain an analyte that has one or more qual ions that does not meet the ±30% requirement, a more thorough visual inspection and a library search should be completed. If the library search returns a match for the analyte, a printout showing the library search results should be included. The original chromatograph report should be notated in such a manner that is clear that the analyst is accepting the quantitative result based on the library search return. The Technical Director, or his

designee, can also sign off on acceptance of spectra, when multiple compounds are returned in a library search.

11.3.1.1.5 The RRT of the sample component is within ±0.06 RRT units of the standard component.

11.3.1.1.6 Structural isomers that produce very similar mass spectra should be identified as individual isomers if they have sufficiently different GC retention times. Sufficient GC resolution is achieved if the height of the valley between the two isomer peaks is less than 25% of the sum of the two peak heights. Otherwise, structural isomers are identified as isomeric pairs.

11.3.1.1.7 Identification is hampered when sample components are not resolved chromatographically and produce mass spectra containing ions contributed by more than one analyte. When gas chromatographic peaks obviously represent more than one sample component (i.e., a broadened peak with shoulder(s) or a valley between two or more maxima), appropriate selection of analyte spectra and background spectra is important. Examination of extracted ion current profiles of appropriate ions can aid in the selection of spectra and in qualitative identification of compounds. When analytes coelute (i.e., only one chromatographic peak is apparent), the identification criteria can be met, but each analyte spectrum will contain extraneous ions contributed by the coeluting compound.

11.3.1.1.8 If any of the above mentioned scenarios are unable to be met for a specific analyte, guidance from the Technical Director should be sought. The Technical Director can determine if the quantified spectra should be reported.

11.3.1.2 For samples containing components not associated with the calibration standards, a library search may be made for the purpose of tentative identification. The necessity to perform this type of identification will be determined by the purpose of the analyses being conducted. Computer generated library search routines should not use normalization routines that would misrepresent the library or unknown spectra when compared to each other. For example, RCRA permit or waste delisting requirements may require the reporting of non-target analytes. Only after visual comparison of sample spectra with the nearest library searches will the mass spectral interpretation specialist assign a tentative identification. Guidelines for making tentative identification are:

(1) Relative intensities of major ions in the reference spectrum (ions> 10% of the most abundant ion) should be present in the sample spectrum.

(2) The relative intensities of the major ions should agree within ±20%. (Example: For ions with an abundance of 50% in the standard spectrum, the corresponding sample ion abundance must be between 30 and 70%)

(3) Molecular ions present in the reference spectrum should be present in the sample spectrum.

(4) Ions present in the sample spectrum but not in the reference spectrum should be reviewed for possible background contamination or presence of coeluting compounds.

(5) Ions present in the reference spectrum but not in the sample spectrum should be reviewed for possible subtraction, because of background contamination or coeluting peaks. Data system library reduction programs can sometimes create these discrepancies.

11.3.2 Quantitative analysis

11.3.2.1 When a compound has been identified, the quantitation of that compound will be based on the integrated abundance of the primary characteristic ion. Quantitation will take place using the internal standard technique. The internal standard used shall be the one nearest the retention time of that of a given analyte. See Table 1 for each analyte and its corresponding internal standard.

11.3.2.2 When MS response is linear and passes through the origin, calculate the concentration of each identified analyte in the sample as follows:

Water

Concentration (μg/L) = $(A_x)(I_s) / (A_{is})(RF)(V_o)$

where:

A_x= Area of characteristic ion for compound being measured.
I_s = Amount of internal standard injected (ng).
A_{is}= Area of characteristic ion for the internal standard.
RF= Mean relative response factor for compound being measured.
V_o= Volume of water purged (mL), taking into consideration any dilutions made.

Sediment/Soil sludge (on a dry-weight basis) and Waste (normally on a wet weight basis)

Concentration in solid (μg/kg) = (A_x)(I_s)(V_t) / (A_{is})(RF)(V_i)(W_s)(D)

where:

A_x, I_s, A_{is}, RF = Same as for water.
V_{st} = Volume of total extract (μL) (use 10,000 μL or a factor of this when dilutions are made).
V_i = Volume of extracted add (μL) for purging.
W_s = Weight of sample extracted or purged (g).
D = % dry weight of sample /100, or 1 for a wet-weight basis.

11.3.2.3 Where applicable, an estimate of concentration for non-calibrated components in the sample can be made. The formulas given above should be used with the following modifications: the areas A_x and A_{is} should be from the total ion chromatograms and the RF for the compound should be assumed to be 1. The concentration obtained should be reported indicating (1) that the value is an estimate and (2) which internal standard was used to determine concentration. Use the nearest internal standard free of interferences.

11.3.2.4 Alternatively, the regression line fitted to the initial calibration may be used for determination of the analyte concentration.

12.0 Quality Control

12.1 Refer to Method 8000B/C and the HEAL Quality Assurance Plan for quality control procedures.

12.1.1 Table 5 is a list of commonly used acronyms. These acronyms are used to abbreviate time needed for writing of explanations.

12.2 The following are general guidelines for data assessment and acceptance criteria for quality control measures.

12.2.1 The GC/MS system must be tuned to meet the BFB specifications.

12.2.2 There must be an initial calibration of the GC/MS system. After each calibration curve, an ICV will be used as a validation. The ICV must be analyzed following a calibration curve and prior to any sample analysis. All failures or adjustments to the initial calibration curve shall be documented on the form located in Appendix 2. This form shall be included with each data folder associated with the current calibration. Every 12 hours a CCV will be analyzed as a

continuing calibration check for the Instrument. The allowable % drift for the ICV is the same as that established for the CCV. These failures will be tracked using an excel spreadsheet to monitor frequency of failures.

12.2.3 The concentrations of the CCV must vary every 12 hour shift.

12.2.4 The absolute retention time should not exceed 0.8min of the initial calibration. If it does shift more than 0.8 min. a new curve should be analyzed.

12.2.4.1 The relative retention times of the sample components should be within ±0.06 RRT units of the standard component.

12.2.5 The GC/MS system must meet the SPCC criteria and the CCC criteria every 12 hours.

12.2.6 For every 20 samples, there should be a MB, LCS, MS, MSD, or Duplicate

12.2.6.1 Method 8260B stipulates that 5 compounds are to be used for the MS and the MSD. The LCS and LCSD will also use these five compounds. They are 1,1-Dichloroethene, Benzene, Trichloroethene, Toluene, and Chlorobenzene. These are to be used to assess the batch.

12.2.6.2 Also, for every 20 samples, there should be a MSD (matrix spike duplicate) or a sample DUP (duplicate). This is done in order to assess the instrument capability to reproduce data.

12.2.6.3 If the sample is highly contaminated or sample volume/ quantity is limited, the analyst should analyze a LCS/LCSD in combination with a sample duplicate.

12.2.6.4 The selected samples used for MS, MSD and DUP shall be rotated among client samples at random so that various matrix problems may be noted and/or addressed. If there is poor performance with the sample selected the client should be notified.

12.2.6.5 For every 20 samples, a RB/MB (reagent blank / method blank) or a soil extraction blank must be analyzed.

12.2.6.6 Control limits for MS, MSD, DUP and LCS are established using the average limits of previous MS, MSD, DUP and LCS from all instruments. These can be updated in the LIMS. Only the Quality Assurance Officer or a Technical Director can update these limits. These limits are updated at a minimum of twice per year. The procedure and formulas are available in the QAP

12.2.6.6.1 New control limits that have been generated can be viewed in the LIMS.

12.3 Air samples collected in Tedlar bags require the analysis of a MB, LCS, and a DUP.

12.4 Additional required instrument QC is found in Section 9.0

12.5 For aqueous and soil matrices, surrogate recoveries should be compared with the control limits listed in the LIMS.

12.5.1 If recovery is not within limits, the following procedures are required.

12.5.1.1 Check to be sure that there are no errors in the calculations, surrogate solutions or internal standards. If errors are found, recalculate the data accordingly.

12.5.1.2 Check instrument performance. If an instrument performance problem is identified, correct the problem and re-analyze the sample.

12.5.1.3 If no problem is found, re-extract and re-analyze.

12.5.1.4 If, upon re-analysis, the recovery is again not within limits, flag the data as "estimated concentration". A brief description of possible matrix interference should be stated in either a CAR or in the sample comments on the Data Entry page of the LIMS.

12.5.2 At a minimum surrogate recovery limits on a matrix by matrix basis, are updated annually.

12.6 Detection Levels

The method detection limit (MDL) is defined as the minimum concentration of a substance that can be measured and reported with 99% confidence that the value is above zero. The MDL actually achieved in a given analysis will vary depending on instrument sensitivity and matrix effects.

Current practices at HEAL define the detection level as the smallest amount that can be detected above the noise in a procedure and within a stated confidence level. We presently utilize an instrument detection level (IDL), a method detection level (MDL) and a practical quantitation limit (PQL).

The IDL is the constituent concentration that when injected produces a signal sufficiently large that 99% of the trials with that amount will produce a detectable signal. The IDL is used to determine the analyte concentration in an extract needed to produce a signal to permit calculations.

The MDL differs from the IDL in that samples containing the constituent of interest are processed through the complete analytical method. The MDL is different than the IDL because of extraction efficiency and extract concentration factors. The MDL is achieved by a well-trained analyst operating a well-calibrated instrument on a non-routine basis (Annually or semi-annually). First add reagent water or the matrix of interest, to make a concentration two to five times higher than the IDL. Prepare and analyze a minimum of seven replicates of this solution. Include all sample-processing steps in the determination. Calculate the sample standard deviation (s) of the replicate measurements. The method detection limit (MDL) can be calculated according to the formula:

MDL = s * t(99%)

t(99%) is the student's t value for the 99% confidence interval. It depends on the number of trials used in calculating the sample standard deviation, so choose the appropriate value according to the number of trials.

Number of Trials	t(99%)
7	3.143
8	2.998
9	2.896
10	2.821
11	2.764
16	2.602
21	2.528

The calculated MDL value must be no more than ten times less than the amount spiked. This is referred to as the 10x Rule.

This method has been tested in a single laboratory using spiked water. Single laboratory precision and accuracy data can be found in Table 6. HEAL calculated MDL data can be found in the LIMS.

The PQL is significant because different laboratories will produce different MDLs even though using the same analytical procedures, instruments and sample matrices. The PQL is generally five times the MDL.

The PQL of Tedlar air samples is determined by using data collected from the analysis of aqueous PQL's and dividing by 50. Lower levels can be achieved by increasing sample amount.

Annual PQL studies can be done when a compound will not be reported below the PQL. The PQL study consists of the analysis of a PQL concentration standard. The allowable % drift is ±50%. PQL studies should only be done with prior approval from Project Management or QAO.

12.7 When a compound is above the instrument calibration range, there are two options for an analyst. First, reanalyze the sample at a dilution in order to bring it within the instruments calibration range. Or if there is not enough sample to reanalyze, then the data will be flagged for that compound. An explanation should also be written into the LIMS so the Project Manager can explain the Flagged data to the client.

12.8 Corrective action reports are to be made for out-of-control data.

12.8.1 Check all calculations and measurements data (standardization, calibration, concentrations, instrument stability, etc.)

12.8.2 Check equipment to ensure that components are functioning properly.
12.8.3 Check reagents (age, preparation, concentrations, etc.)

12.8.4 Assure that proper procedures were followed.

12.8.5 Reanalyze all samples run in conjunction with the duplicate and/or spike.

12.8.6 In the case that a sample is treated differently than is in the documented procedure, a corrective action report will be filed explaining the reason.

12.8.7 If deficiency continues, consult with the appropriate senior staff member.

12.8.8 A second mechanism for initiation of corrective action is that resulting from Quality Assurance activities concerned with performance audits, system audits, inter and intra-laboratory comparison studies. Corrective Actions initiated through this mechanism will be monitored and coordinated by the laboratory QA Officer.

12.8.9 If an analysis must be reported to the client and it is out-of-control, corrective action sheets are to be filled out and the client will be notified. These are to be kept in the analysts daily run folder and the LIMS. The corrective action sheet will be signed both by the analyst and the supervisor or the QA Officer.

12.8.10 All samples which have been preserved using HCL are checked by the analyst after analysis to determine if the pH requirement for appropriate preservation is met. This is done using narrow range pH paper. The pH must be at or below 2. The sample identification and resulting pH is recorded in the pH logbook (V-42) along with the date and initials of the analyst. If the pH is higher than 2, the resulting pH value should be included in the narrative/ comments section in the LIMS.

12.8.11 The sample storage deli/refrigerator is checked weekly for possible contamination using a VSB (volatile storage blank). The data collected from the VSB are scrutinized to ensure samples are not being cross contaminated or being contaminated from possible laboratory contaminations.

12.8.12 See HEAL QAP for additional QA parameters and an in-depth description of DOC/ADOCP.

13.0 Calculations

13.1 This method has been tested in a single laboratory using spiked water. Using a mega-bore capillary column, water was spiked at concentrations between 0.5 and 10 μg/L. Single laboratory accuracy

and precision data are presented for the method analytes in Table 6 of method 8260B. Calculated MDLs are available in the method folder located in the QAO office or through the LIMS.

13.2 Calculations for Aqueous and Tedlar Air samples:

$$\left[\frac{Height\,or\,Area\,of\,sample}{Height\,or\,Area\,of\,standard}\right] X \left[Standard\,concentration\right] X \left[Dilution\,factor\right] X \left[\frac{Final\,volume}{Inital\,volume}\right]$$

13.2.1 Example calculation for an aqueous sample:

Given: Sample height = 1500, Standard height = 4158, and Standard concentration = 5.12 µg/mL

$$\left[\frac{1800}{4158}\right] X \left[\frac{5.12\,\mu g}{ml}\right] X \left[1.0x\,dilution\right] X \left[\frac{\left(2\,ml / .400\,ml\right)}{500\,ml}\right] X \left[\frac{1000\,ml}{1\,L}\right] = 22.2\,\frac{\mu g}{L}$$

report as 22µg/L

13.2.1.1 Example calculation for an aqueous sample using a HEAL generated report.

For example, the amount on the 8260B chromatogram for benzene = 100ng. Divide this number by 5 to get a reporting level of 20ug/L. If there was a dilution, multiply the ug/L result times the dilution to get the new sample concentration in ug/L. For example, benzene = 100ng. There was a 20X dilution. 100/5=20ug/L. Then take the 20ug/L and multiply by 20 to get a concentration of 400ug/L.

13.2.2 Calculations for non-aqueous samples:

$$\left[\frac{Height\,or\,Area\,of\,sample}{Height\,or\,Area\,of\,standard}\right] X \left[Standard\,concentration\right] X \left[Dilution\,factor\right] X \left[\frac{Final\,volume}{Inital\,weight}\right]$$

13.2.2.1 Example calculation for a non-aqueous sample using a HALL generated report.

For example, the amount on the 8260B chromatogram for toluene = 200ng. Divide this number by the number of uL of sample injected on the concentrator. Standard non-diluted samples contain 100uL of sample per 5ml of DI water. Therefore the 200ng amount reported on the chromatogram for toluene is divided by 100 to yield a result of 2.0 mg/Kg. If the sample was diluted by 10, only 10 uL of sample would be inject on the concentrator. Thus a 200ng result on the chromatogram would be divided by 10 to yield a result of 20 mg/Kg.

13.2.3 See Method 8000B/C, Method 8260B, and QAP for any additional calculations which may not be presented here.

14.0 Pollution Prevention

14.1 Care must be given by each analyst to minimize the number of re-analyses using this SOP. In doing so, waste generated in the use of this SOP will be kept to a minimum.

14.2 The analyst should use historical data as a guideline for possible sample dilutions which will reduce the number of re-analyses.

14.2.1 Historical data should be used only as a tool to minimize waste. The historical data should not be used to circumvent any policy set forth in US EPA methodology or the Hall Environmental Analysis Laboratory QA Plan.

15.0 Waste Management

15.1 After completion of analysis, all sample and sample containers will be disposed of in accordance with the HEAL Chemical Hygiene Plan.

16.0 Method Performance

16.1 Each analyst must establish a Demonstration of Capability (DOC) by performing a Precision and Accuracy (P&A) study before being considered proficient for this Method. The P&A will consist of generating 4 spike samples and their analysis. Acceptable criteria that is used for determining the compliance of the P&A study is defined in the current HEAL QAP. Spike data for DOC's will be included in each analysts file folder. DOC's will be updated on an annual basis, either through proficiencies or using spikes associated with extraction batches.

16.2 Analysts also analyze PT (proficiency test) samples every 6 months. The PT is a multifold test consisting of a blind spike that checks both analysts and instrumentation.

16.3 All test specific, method performance data can be viewed via the LIMS. All analysts have received training on how to access this data in the LIMS. This data includes control limits, MDL's, and PQL's.

16.4 Estimation of Measurement Uncertainty

16.4.1 Estimation of measurement uncertainty is to be conducted on an annual basis.

16.4.2 Collect, at least, 20 of the most recent LCS data points. Calculate the standard deviation (s) and the average of all the readings. Calculate the 95% confidence interval (2 times the standard deviation). Report the estimated uncertainty as ±2(s). Uncertainty data is kept on file in the method folders in the quality control office.

16.4.3 Calculate standard deviation (s) and 95% confidence interval according to the following formulae:

$$s = \sqrt{\frac{\sum (x - \bar{x})^2}{(n-1)}}$$

Where: s = standard deviation
x = number in series
$\bar{x}$ = calculated mean of series
n = number of samples taken

$$95\%\ confidence = 2 \times s$$

16.6 MDLs must be empirically established annually. Refer to the Quality Assurance Manual.

16.6 PQL studies, when used, are completed annually.

17.0 References

17.1 Test Methods for Evaluating Solid Waste: Physical/Chemical Methods, US EPA SW-846, 3rd Edition, September 1986.

17.2 Volatile Organic Compounds by Gas Chromatography/ Mass Spectrometry (GC/MS), US EPA 8260B, December, 1996.

17.3 Quality Assurance Plan, Hall Environmetnal Analysis Laboratory.

17.4 Chemical Hygiene Plan, Hall Environmetnal Analysis Laboratory.

17.5 Standard Operating Procedure for Closed System Purge and Trap Analysis of Volatile Organics in Aqueous Samples and Method 5035, Hall Environmetnal Analysis Laboratory.

17.6 Determinative Chromatographic Separations, US EPA 8000B.

17.7 Extraction Method for Volatile Organic Compounds in Soil and Waste Samples – High Concentration – for Closed System Purge and Trap Analysis, Hall Environmetnal Analysis Laboratory.

17.8 pH Logbook V-42, HEAL, February, 2006. (or most current)

Table 1.
Chromatographic Elution Order and Corresponding Internal Standard For Volatile Organic Compounds on a Capillary Column

Compound Name	Elution Order	Internal Standard
Dichlorodifluoromethane	1	Pentafluorobenzene
Chloromethane	2	Pentafluorobenzene
Vinyl chloride	3	Pentafluorobenzene
Bromomethane	4	Pentafluorobenzene
Chloroethane	5	Pentafluorobenzene
Trichlorofluoromethane	6	Pentafluorobenzene
Acetone	7	Pentafluorobenzene
1,1-Dichloroethene	8	Pentafluorobenzene
Carbon Disulfide	9	Pentafluorobenzene
Methylene chloride	10	Pentafluorobenzene
trans-1,2-Dichloroethene	11	Pentafluorobenzene
1,1-Dichloroethane	12	Pentafluorobenzene
2-Butanone	13	Pentafluorobenzene
2,2-Dichloropropane	14	Pentafluorobenzene
cis-1,2-Dichloroethene	15	Pentafluorobenzene
Bromochloromethane	16	Pentafluorobenzene
Chloroform	17	Pentafluorobenzene
1,1,1-Trichloroethane	18	Pentafluorobenzene
1,1-Dichloropropene	19	Pentafluorobenzene
Carbon tetrachloride	20	Pentafluorobenzene
Benzene	21	Pentafluorobenzene
1,2-Dichloroethane	22	Pentafluorobenzene
Trichloroethene	23	Pentafluorobenzene
1,2-dichloropropane	24	Pentafluorobenzene
4-methyl-2-pentanone	25	Benzene-d5, chloro-
Bromodichloromethane	26	Benzene-d5, chloro-
Dibromomethane	27	Benzene-d5, chloro-
cis-1,3-Dichloropropene	28	Benzene-d5, chloro-
Toluene	29	Benzene-d5, chloro-
trans-1,3-Dichloropropene	30	Benzene-d5, chloro-
1,1,2-Trichloroethane	31	Benzene-d5, chloro-
1,3-Dichloropropane	32	Benzene-d5, chloro-
2-Hexanone	33	Benzene-d5, chloro
Tetrachloroethene	34	Benzene-d5, chloro-
Dibromochloromethane	35	Benzene-d5, chloro-
Ethylene dibromide	36	Benzene-d5, chloro-
Chlorobenzene	37	Benzene-d5, chloro-
Ethylbenzene	38	Benzene-d5, chloro-
1,1,1,2-Tetrachloroethane	39	Benzene-d5, chloro-
m-Xylene	40	Benzene-d5, chloro-

Compound Name	Elution Order	Internal Standard

p-Xylene	41	Benzene-d5, chloro-
o-Xylene	42	Benzene-d5, chloro-
Styrene	43	Benzene-d5, chloro-
Isopropylbenzene	44	Benzene-d5, chloro-
Bromoform	45	Benzene-d5, chloro-
1,1,2,2-Tetrachloroethane	46	Benzene-d5, chloro-
1,2,3-Trichloropropane	47	1,4-dichlorobenzene-d4
Propylbenzene	48	1,4-dichlorobenzene-d4
Bromobenzene	49	1,4-dichlorobenzene-d4
1,3,5-Trimethylbenzene	50	1,4-dichlorobenzene-d4
2-Chlorotoluene	51	1,4-dichlorobenzene-d4
4-Chlorotoluene	52	1,4-dichlorobenzene-d4
tert-Butylbenzene	53	1,4-dichlorobenzene-d4
1,2,4-Trimethylbenzene	54	1,4-dichlorobenzene-d4
sec-Butylbenzene	55	1,4-dichlorobenzene-d4
p-Isopropyltoluene	56	1,4-dichlorobenzene-d4
1,3-Dichlorobenzene	57	1,4-dichlorobenzene-d4
1,4-Dichlorobenzene	58	1,4-dichlorobenzene-d4
n-Butylbenzene	59	1,4-dichlorobenzene-d4
1,2-Dichlorobenzene	60	1,4-dichlorobenzene-d4
1,2-Dibromo-3-chloropropane	61	1,4-dichlorobenzene-d4
1,2,4-Trichlorobenzene	62	1,4-dichlorobenzene-d4
Hexachlorobutadiene	63	1,4-dichlorobenzene-d4
Naphthalene	64	1,4-dichlorobenzene-d4
1,2,3-Trichlorobenzene	65	1,4-dichlorobenzene-d4

Internal Standards/Surrogates
Chlorobenzene-d5
1,4-Dchlorobenzene -d4
4-Bromofluorobenzene
1,2-Dichloroethane-d4
Dibromofluoromethane
Toluene-d8
Pentafluorobenzene

Elution orders are both column and instrument dependant and are subject to change. The associated Internal Standard may vary slightly between instruments due to elution order.

Table 2.
Estimated Quantitation Limits For Volatile Analytes

	Estimated Quantitation Limits (All analytes in Table 1)	
	Ground water µg/L	Low Soil/Sediment µg/kg
Purging 5mL of water	5	--
Purging 25mL of water	1	--
Soil/Sediment	-	5

Estimated Quantitation Limit (EQL) - The lowest concentration that can be reliably achieved within specified limits of precision and accuracy during routine laboratory operating conditions. The EQL is generally 5 to 10 times the MDL. However, it may be nominally chosen within these guidelines to simplify data reporting. For many analytes the EQL is selected from the lowest non-zero standard in the calibration curve. Sample EQLs are highly matrix-dependent. The EQLs listed herein are provided for guidance and may not always be achievable.

EQLs listed for soil/sediment are based on wet weight. Normally data are reported on a dry weight basis; therefore, EQLs will be higher, based on the percent dry weight in each sample.

Other Matrices	Factors
Water miscible liquid waste	50
High-concentration soil and sludge	125
Non-water miscible waste	500

EQL = [EQL follow soil/sediment (see Table 1)] x [Factor]. For non-aqueous samples, the factor is on a wet-weight basis.

Table 3.
BFB Mass - Intensity Specifications (4-Bromofluorobenzene)

Mass	Intensity Required (relative abundance)
50	15 to 40% of mass 95
75	30 to 60% of mass 95
95	base peak, 100% relative abundance
96	5 to 9% of mass 95
173	less than 2% of mass 174
174	greater than 50% of mass 95
175	5 to 9% of mass 174
176	greater than 95% but less than 101% of mass 174
177	5 to 9% of mass 176

Alternate tuning criteria may be used (e.g. CLP, Method 524.2, or manufacturers' instructions), provided that method performance is not adversely affected.

Table 4.
Characteristic Mass (M/Z) For Purgeable Organic Compounds

Compound Name	Primary Characteristic Ion	Secondary Characteristic Ion
Acetone	58	43
Acetonitrile	41	41, 40, 39
Acrolein (Propenal)	56	55, 58
Acrylonitrile	53	52,51
Allyl alcohol	57	57, 58, 39
Allyl chloride	76	76, 41, 39, 78
Benzene	78	-
Benzyl chloride	91	91, 126, 65, 128
Bromoacetone	136	43, 136, 138, 93, 95
Bromochloromethane	156	77, 158
Bromodichloromethane	128	49, 130
Bromoform	173175, 254	
Bromomethane	94	96
n-Butanol	56	41
2-Butanone (MEK)	72	43,72
n-Butylbenzene	91	92, 134
sec-Butylbenzene	105	134
tert-Butylbenzene	119	91, 134

Table 4. (continued)

Compound Name	Primary Characteristic Ion	Secondary Characteristic Ion
Carbon disulfide	76	78
Carbon tertachloride	117	119
Chloral hydrate	82	44, 84, 86, 111
Chloroacetomitrile	48	75
Chlorobenzene	112	77, 114
Chlorodibromomethane	129	208, 206
Chloroethane	64(49*)	66(51*)
2-Chloroethanol	49	49, 44, 43, 51, 80
bis-(2-Chloroethyl)sulfide	109	111, 158, 160
2-Chloroethyl vinyl ether	63	65,106
Chloroform	83	85
Chloromethane	50 (49*)	52(51*)
Chloroprene	53	53, 88, 90, 51
3-Chloropropionitrile	54	54, 49, 89, 91
2-Chlorotoluene	91	126
4-Chlorotoluene	91	126
1,2-Dibromo-3-chloropropane	75	155, 157
1,2-Dibromoethane	107	109,188
Dibromomethane	93	95,174
1,2-Dichlorobenzene	146	111,148
1,3-Dichlorobenzene	146	111, 148
1,4-Dichlorobenzene	146	111, 148
cis-1,4-Dichloro-2-butene	75	75, 53, 77, 124, 89
trans-1,4-Dichloro-2-butene	53	88, 75
Dichlorodifluoromethane	85	87
1,1-Dichloroethane	63	65, 83
1,2-Dichloroethane	62	98
1,1-Dichloroethene	96	61, 63
cis-1,2-Dichloroethene	96	61, 98
trans-1,2-Dichloroethene	96	61, 98
1,2-Dichloropropane	63	112
1,3-Dichloropropane	76	78
2,2-Dichloropropane	77	97
1,3-Dichloro-2-propanol	79	79, 43, 81, 49
1,1-Dichloropropene	75	110, 77
cis-1,3-Dichloropropene	75	77, 39
trans-1,3-Dichloropropene	75	77, 39
1,2,3,4-Diepoxybutane	55	55, 57, 56
Diethyl ether	74	45, 59
1,4-Dioxane	88	88, 58, 43, 57
Epichlorhydrin	57	57, 49, 62, 51
Ethanol	31	45, 27, 46
Ethyl acetate	88	43, 45, 61
Ethylbenzene	91	106
Ethylene oxide	44	44, 43, 42
Ethyl methacrylate	69	69, 41, 99 86, 114

Table 4. (continued)

Compound Name	Primary Characteristic Ion	Secondary Characteristic Ion
Hexachlorobutadiene	225	223, 227
Hexachloroethane	201	166, 199, 203
2-Hexanone	43	58, 57, 100
Iodomethane	142	127, 141
Isobutyl alcohol	43	43, 41, 42, 74
Isopropylbenzene	105	120
Malononitrile	66	66, 39, 65, 38
Methacrylonitrile	41	41, 67, 39, 52, 66
Methyl acrylate	55	85
Methyl-t-butyl ether	73	57
Methylene chloride	49	86, 84
Methyl iodide	142	142, 127, 141
Methyl methacrylate	69	69, 41, 100, 39
4-Methyl-2-pentanone (MIBK)	100	43, 58, 85
Naphthalene	128	-
Nitrobenzene	123	51, 77
2-Nitropropane	46	-
Pentachloraethane	167	167, 130, 132, 165, 169
2-Picoline	93	93, 66, 92, 78
Propargyl alcohol	55	55, 39, 38, 53
β-Propiolactone	42	42, 43, 44
Propionitrile (ethyl cyanide)	54	54, 52, 55,40
n-Propylamine	59	59, 41, 39
n-Propylbenzene	91	120
Pyridine	79	52
Styrene	104	78
1,2,3-Trichlorobenzene	180	182, 145
1,2,4-Trichlorobenzene	180	182, 145
1,1,2,2,-Tetrachloroethane	83	131, 85
1,1,1,2-Tetrachloroethane	131	133, 119
Tetrachloroethene	164	129, 131, 166
Toluene	92	91
1,1,1-Trichloroethane	97	99, 61
1,1,2-Trichloroethane	83	97, 85
Trichloroethene	95	97, 130, 132
Trichlorofluoromethane	151	101, 153
1,2,3-Trichloropropane	75	77
1,2,4-Trimethylbenzene	105	120
1,2,5-Trimethylbenzene	105	120
Vinyl acetate	43	86
Vinyl chloride	62	64
o-Xylene	106	91
m-Xylene	106	91
p-Xylene	106	91

Table 4. (continued)

Compound Name	Primary Characteristic Ion	Secondary Characteristic Ion
InternalStandards/Surrogates		
Chlorobenzene-d5	117	
1,4-Dichlorobenzene-d4 150	115, 152	
4-Bromofluorobenzene 95	174, 176	
Dibromofluoromethane 111		
1,2-Dichloroethane-d4 65	102	
Toluene-d8 98		
Pentafluorobenzene	99	168

* - characteristic ion for an ion trap mass spectrometer (to be used when ion-molecule reactions are observed)

1.0 Scope and Application

1.1 This procedure describes the process of preparation of instrumentation, including calibration and sample preparation and sample analysis using the low soil method outlined in EPA Method 5035.

1.2 The information contained herein is currently only pertinent to one GCMS system; GCMS THOR.

1.3 The use of this procedure is restricted to use by, or under the supervision of analysts experienced in the use of purge-and-trap systems and gas chromatograph/mass spectrometers and skilled in the interpretation of mass spectral data. The analyst must have established Demonstration of Capability (DOC) using EPA Method 5035B.

2.0 Summary

2.1 The 5 gram samples are collected in hermetically sealed containers with zero headspace. The sample(s) is immediately stored on ice at 4^0C for shipment to the laboratory. The sample(s) is the placed in a freezer able to maintain a temperature <-15^0C until transferred to a 40-mL VOA vial.

2.2 The entire sample is place into a 40-mL VOA vial containing a stir bar and 5mL of de-ionized water. The sample is the either analyzed immediately or frozen and analyzed within 14 days of collection.

2.3 The sample analysis occurs using the Archon attached to GCMS Thor. The sample vial is placed into the sample tray and allowed to warm to room temperature. The Archon is then programmed to purge the sample for approximately 9 minutes; i.e. the purging takes place directly on the Archon.

3.0 Apparatus and Reagents

3.1 Purge-and-trap device

4.1.1 Tekmar 3000 equipped with a Vocarb 3000 trap (Type K)

3.2 Archon Auto-sampler

4.2.1 Archon must be equipped with a soil sparge option. Currently the only Archon equipped with this option is connected to GCMS THOR.

3.3 Gas chromatograph/mass spectrometer (GCMS) system:

4.3.1 Instrument name: THOR

3.3.1.1 Varian CP-3800 gas chromatograph equipped with a Varian 1079 injector.

3.3.1.2 Varian Saturn 2000 Mass Spectrometer.

3.3.1.3 Data system: Varian MSWorkstation 6.9

3.3.1.4 Column: RTX-Volatiles 30m x 0.25mm ID x 1.0µm

3.4 Top-loading balance capable of weighing to 0.01g.

3.5 Gas-tight micro-syringes – 1.0µl to 1000µl.

3.6 40mL VOA vials.

3.7 Teflon coated Stir bars 12mm x 4mm.

3.8 Stainless steel spatula.

3.9 Organic-free reagent water – all references to water in this method refer to organic-free reagent water.

3.10 Methanol – used in standard preparation.

3.11 Standards – All stock standards are at a concentration of 2000 or 2500ppm.

4.0 Procedure

4.1 Samples are received by the laboratory and placed in an appropriate freezer able to maintain a temperature < -15^{0}C.

4.2 Before any sample analysis can be analyzed, a valid calibration must be current. If the calibration is not valid a calibration must be analyzed.

4.2.1 A 5ppm 8260 standard shall be prepared for calibration levels.

4.2.2 A minimum of 6 calibration levels should be analyzed.

4.2.3 A 5ppm 8260 ICV shall be prepared to validate the calibration.

4.2.4 Obtain appropriate number of VOA vials and place a clean stir bar in each vial. Place a vial on the top-loading balance and tare. Place 5g of clean sand (OTTAWA sand) into each vial. After placing sand in vial, add approximately 5mL of DI-water into each vial. Label each vial cap with a ppb concentration amount. Spike each vial with the appropriate micro-liter amount using the 5ppm 8260 calibration standard. See Table A.

4.2.5 Tuning criteria must be met prior to calibration or sample analysis. Analyze a blank vial (not spiked with standard).

4.2.6 Once confirming tune criteria has been met, analyze calibration in increasing concentrations. Analyze two blank vials after the highest concentration and before the analyses of the ICV. The ICV spiked into the VOA vial shall be at a concentration of 5ppb.

4.2.7 Pass/Fail criteria and Quality Control frequency for this addendum is no different than that required by 8260B.

4.3 After an acceptable calibration is achieved and validated, sample analysis can begin.

4.3.1 Remove sample from the storage freezer.

4.3.2 Obtain appropriate amount of VOA vials and place a stir bar in each.

4.3.3 Place each vial onto the top-loading balance and tare.

4.3.4 Carefully remove the sample for its collection vessel and place entire amount into the VOA vial. Record the sample weight on the PREP LOG.

4.3.4.1 Do not forget to transfer all weights into the LIMS PREP LOG for ease of LIMS operation.

4.3.5 After making certain each weight is recorded, add 5mL of DI water using a pipette. The amount of water does not have to be exact, merely consistent.

4.3.6 Once sample preparation is complete, place vials into the auto-sampler tray. Allow samples to thaw and reach ambient temperature.

4.3.7 Using method 30 on the Archon auto-sampler, program the appropriate vial positions for analysis.

4.3.7.1 Make certain that the purge heater temperature is at 40^{0}C and that the stir option is set to "YES".

4.3.8 Start the archon on method 30.

4.3.9 No dilutions can be done using this SOP.

4.4 After analysis, stir bars are removed from VOA vials. The stir bars are washed with de-ionized water and rinsed with methanol. The stir bars are then placed in a drying oven for a minimum of eight hours.

5.0 Quality Control and Data Results

5.1 All quality control parameters required by S-8260B-8 pertain herein. The only exception to this is the requirement of an MS/MSD. Analyses of MS/MSD can only be done if provided adequate sample volume. A LCS/LCSD and sample duplicate shall be analyzed.

5.2 To obtain the final calculated result. Divide the report value by the sample weight. The final reportable units are µg/Kg.

6.0 Reference

6.1 EPA METHOD 5035, December 1996

TABLE A.

Amount of 5ppm Standard in µL	Concentration in ppb
0.4	0.4
1	1
2	2
3	3
5	5
10	10
20	20

Appendix 2. Calibration Check List

Instrument: _____ **Date:** _____

Calibration Name: _____

Analyte	ICV Pass	ICV Fail	Calibration Curve Points Removed Yes	Calibration Curve Points Removed No	Notes

Dichlorodifluoromethane	☐	☐	☐	☐	_____
Chloromethane	☐	☐	☐	☐	_____
Vinyl chloride	☐	☐	☐	☐	_____
Vinyl Acetate	☐	☐	☐	☐	_____
Chloroethane	☐	☐	☐	☐	_____
Bromomethane	☐	☐	☐	☐	_____
Trichlorofluoromethane	☐	☐	☐	☐	_____
1,1-Dichloroethene	☐	☐	☐	☐	_____
Carbon disulfide	☐	☐	☐	☐	_____
MTBE	☐	☐	☐	☐	_____
Isopropyl ether	☐	☐	☐	☐	_____
Acetone	☐	☐	☐	☐	_____
Iodomethane	☐	☐	☐	☐	_____
t-butanol	☐	☐	☐	☐	_____
ETBE	☐	☐	☐	☐	_____
trans-1,2-DCE	☐	☐	☐	☐	_____
1,1-Dichloroethane	☐	☐	☐	☐	_____
2,2-Dichloropropane	☐	☐	☐	☐	_____
Methylene Chloride	☐	☐	☐	☐	_____
2-Butanone	☐	☐	☐	☐	_____
TAME	☐	☐	☐	☐	_____
1,1,1-Trichloroethane	☐	☐	☐	☐	_____
1,1-Dichloropropene	☐	☐	☐	☐	_____
Carbon Tetrachloride	☐	☐	☐	☐	_____
cis-1,2-DCE	☐	☐	☐	☐	_____
Benzene	☐	☐	☐	☐	_____
Chloroform	☐	☐	☐	☐	_____
Bromochloromethane	☐	☐	☐	☐	_____
Trichloroethene (TCE)	☐	☐	☐	☐	_____
Dibromofluoromethane	N/A	N/A	☐	☐	_____
1,2-Dichloroethane-d4	N/A	N/A	☐	☐	_____
1,2-Dichloroethane (EDC)	☐	☐	☐	☐	_____
1,2-Dichloropropane	☐	☐	☐	☐	_____
isobutyl alcohol	☐	☐	☐	☐	_____
4-Methyl-2-pentanone	☐	☐	☐	☐	_____
2-choroethyl-vinyl-ether	☐	☐	☐	☐	_____
Toluene-d8	N/A	N/A	☐	☐	_____
Toluene	☐	☐	☐	☐	_____
Tetrachloroethene (PCE)	☐	☐	☐	☐	_____
2-Hexanone	☐	☐	☐	☐	_____
cis-1,3-dichloropropene	☐	☐	☐	☐	_____
Bromodichloromethane	☐	☐	☐	☐	_____
Dibromomethane	☐	☐	☐	☐	_____
1,3-Dichloropropane	☐	☐	☐	☐	_____
trans-1,3-dichloropropene	☐	☐	☐	☐	_____
Ethylbenzene	☐	☐	☐	☐	_____

Analyte	ICV Pass	ICV Fail	Calibration Curve Points Removed Yes	Calibration Curve Points Removed No	Notes
mp-Xylenes	☐	☐	☐	☐	
1,1,2-Trichloroethane	☐	☐	☐	☐	
Chlorobenzene	☐	☐	☐	☐	
1,2-Dibromoethane (EDB)	☐	☐	☐	☐	
o-Xylene	☐	☐	☐	☐	
Isopropylbenzene	☐	☐	☐	☐	
1,1,1,2-Tetrachloroethane	☐	☐	☐	☐	
Dibromochloromethane	☐	☐	☐	☐	
Styrene	☐	☐	☐	☐	
n-Propylbenzene	☐	☐	☐	☐	
1,3,5-Trimethylbenzene	☐	☐	☐	☐	
tert-Butylbenzene	☐	☐	☐	☐	
2-Chlorotoluene	☐	☐	☐	☐	
sec-Butylbenzene	☐	☐	☐	☐	
4-Bromofluorobenzene	N/A	N/A	☐	☐	
4-Chlorotoluene	☐	☐	☐	☐	
1,2,4-Trimethylbenzene	☐	☐	☐	☐	
Bromobenzene	☐	☐	☐	☐	
Toluene, p-isopropyl	☐	☐	☐	☐	
Bromoform	☐	☐	☐	☐	
n-Butylbenzene	☐	☐	☐	☐	
1,2,3-Trichloropropane	☐	☐	☐	☐	
1,1,2,2-Tetrachloroethane	☐	☐	☐	☐	
1,3-Dichlorobenzene	☐	☐	☐	☐	
1,4-Dichlorobenzene	☐	☐	☐	☐	
1,2-Dichlorobenzene	☐	☐	☐	☐	
Hexachlorobutadiene	☐	☐	☐	☐	
1,2-Dibromo-3-chloropropane	☐	☐	☐	☐	
1,2,4-Trichlorobenzene	☐	☐	☐	☐	
1,2,3-Trichlorobenzene	☐	☐	☐	☐	
Naphthalene	☐	☐	☐	☐	
2-Methylnaphthalene	☐	☐	☐	☐	
1-Methylnaphthalene	☐	☐	☐	☐	

Analyst: _____

QA: _____

Additional Comments: _____

6

Hall Environmental Analysis Laboratory

SOP Acknowledgement Sheet
SOP Name-SM2320B
SOP Number- S-SM2320-5

I state that I have read the above referenced SOP. I understand all portions of the document as they apply to me and my responsibilities at Hall Environmental Analysis Laboratory. I further recognize this as the most current application of the SOP and that it supersedes any previous documents that I may have utilized.

Effective: 6/15/2011 - 8/29/2011

Name	Signature	Date	Supervisor Initials	QA Officer Initials
Ian Cameron		6/16/11		
Tiffany Shaw		6/13/11		
Lavonda Begay		6/15/11		
Carolyn Swanson		8/9/2011		

Hall Environmental Analysis Laboratory

Common Name: Standard Methods 2320B	S.O.P. Number: S-SM2320-5
Method: Automated and Manual Methods for Total Alkalinity, Low Level Alkalinity, Carbonate, Bicarbonate, and Hydroxide in Aqueous Samples	
Effective Date: June 15, 2011	Supersedes: S-SM2320-4
Laboratory Manager	Quality Assurance/Quality Control Officer:

1.0 Scope and Application

1.1. This SOP describes the procedure for determining the total alkalinity and its three common forms found in most waters (carbonate, bicarbonate, and hydroxide). This SOP is applicable to drinking, surface, and saline waters, domestic and industrial wastes.

1.2. There are two processes that can be used for alkalinity determination.

1.2.1 Titration of alkalinity by the automated system, Poseidon.

1.2.1.1 The automated titration system is the preferred procedure for the determination of alkalinity. Before use of the automated system, the analyst must have read and understood this SOP and the current pH and eC SOPs within proven demonstration of capability.

1.2.1.1.1 This SOP relies heavily on information already included in the current pH SOP. Any change in the current pH SOP must be adhered to when using this SOP.

1.2.1.2 Appendix B contains a comparison study of automated titration versus manual titration. This is included to show that there is no significant difference between the two procedures.

1.2.2 Manual titration using a 50mL and or 10mL micro-burette, and Memphis a pH meter.

1.2.2.1 The manual titration system is used as a back up system should the auto-titrator system be temporarily out of service, or if analyzing samples that have a difficult matrix with an extreme buffering capacity.

1.3. This SOP also includes the procedure for determining low level alkalinities (alkalinities that are less than 20mg/L).

2.0 Summary of method

2.1. A 50mL aliquot of sample is titrated with 0.02N H_2SO_4 to the pH 8.3 endpoint (if possible) and the pH 4.5 endpoint. Carbonates, bicarbonates, and hydroxides are calculated from the titration volumes to both the pH 8.3 and 4.5 endpoints.

2.1.1 The automated system (Poseidon) titrates to all pH endpoints and calculates all results.

2.1.1.1 All method parameters and calculations used by the Tiamo software are included in Appendix C.

2.2. For low level alkalinities 50mL of sample is titrated with 0.02N H_2SO_4 using a 10mL micro burette to pH 4.5 ±0.20 endpoint. Additional titrant is added to lower the pH an additional 0.3 pH units. Low level alkalinity is then calculated from the titration volumes to pH 4.5 and 0.3 pH unit's lower endpoints.

2.2.1 The automated system titrates each sample to a final endpoint of 4.2pH units.

2.2.2 The automated system calculates the low level alkalinity result.

3.0 Detection Limits

3.1. The PQL for total alkalinity and bicarbonate is 20 mg/L as $CaCO_3$. The PQL for carbonate and hydroxide is 2.0 mg/L as $CaCO_3$.

3.2. The PQL for low level alkalinity is 2.0 mg/L as $CaCO_3$.

3.3. No values for alkalinity will be reported below the appropriate PQL. A PQL study shall be done, at a minimum, of yearly.

3.3.1. The PQL study for Poseidon shall consist of a blank spike at a concentration of 20mg/L and 2.0mg/L. The calculated recovery after titration must be ±50% of each. Each value should also be blank corrected.

3.3.2. The PQL study for the manual titration procedure shall consist of a blank spike of 20mg/L using the 50mL burette and 2.0mg/L for the 10mL micro-burette. The calculated recovery after titration of each blank spike must be ±50% of the theoretical concentration spiked. Each value should be blank corrected.

4.0 Definitions

4.1. Analytical batch – up to 20 samples and including a MB, LCS, MS, and MSD; i.e. 24 individual determinations.

4.2. Blank spike – A blank spike is a know amount of DI H_2O fortified with a known concentration of desired spike. Different than the LCS due to concentration(s).

4.3. DI H_2O (DI) – De-ionized water/reagent free water. Water that is passed through a purification system and is free from interferences.

4.4. Laboratory Control Spike (LCS) – Laboratory fortified blank that is spiked at a concentration of 80mg/L.

4.5. Matrix Spike (MS) – A sample fortified, at 80mg/L, in the laboratory.

4.6. Matrix Spike Duplicate (MSD) – A duplicate sample fortified, at 80mg/L, in the laboratory.

4.7. Method Blank (MB) – A blank analysis of DI H_2O analyzed prior to LCS and samples. The calculated value must be less than that of the necessary PQL for samples being determined. Used to show that the automated system is clean prior to sample analysis.

5.0 Interferences

5.1. Salts of weak organic acids and inorganic acids, present in large amounts, may cause interference in electrometric pH measurements.

5.2. Oil and grease present in samples may coat the electrode and hinder the electrometric response.

5.3. Temperature of the sample(s) being analyzed can cause interference with the pH electrode. Samples should be allowed to equilibrate to room temperature prior to analysis. The pH electrode used must correct for temperature differences.

6.0 Safety

6.1. For handling procedures and exposure information always refer to the MSDS. To keep exposure to reagents and standards to a minimum, analysts must wear appropriate laboratory safety attire when working in the laboratory.

6.2. Refer to the current HEAL Health and Safety Manual.

7.0 Apparatus and Materials

7.1. Poseidon – 855 Robotic Titrosampler equipped with: 1-50mL 807 Dosino dosing unit, 1-20mL 807 Dosino dosing unit with 2L bottle, 772 peristaltic pump unit, pH combination electrode, Conductivity combination electrode, and Tiamo software for operation and data collection.

7.1.1 50mL graduated cylinder.

7.1.2 50mL disposable digestion vessels from Environmental Express.

7.1.3 60mL disposable plastic cups.

7.1.4 DI H_2O.

7.1.5 Sulfuric Acid, Standardized, 0.02000N.

7.1.5.1 The concentration of the sulfuric acid may change.

7.1.5.2 The Technical Director must be notified of change so that software can be configured to calculate appropriately due to new normality of sulfuric acid.

7.1.6 Sulfuric Acid, Standardized, 0.2N.

7.1.7 Spike solution

7.1.7.1 Stock Alkalinity solution: Dissolve 0.848g of Na_2CO_3 (dried at 250°C for two hours and cooled in a desiccator) into 200mL DI water. This is a 4000 mg/L as $CaCO_3$ stock solution.

7.1.7.2 Spike 1mL into LCS, MS, and MSD samples. Report spike results as total alkalinity only. In LCS, MS, and MSD the spike concentration is 80 mg/L as $CaCO_3$.

7.1.8 pH Buffers 1.68, 4.00, 7.00, 10.00, 12.45 for calibration of pH meter.

7.1.8.1 See current pH SOP for further detailed discussion of pH calibration and requirements.

7.2. Memphis - pH meter. See current HEAL SOP for determination of pH for detailed calibration procedures and QC requirements.

7.2.1 50mL graduated cylinder.

7.2.2 Magnet stirrer and Teflon stir-bars.

7.2.3 Class A 50mL burette – Cartman.

7.2.4 Class A 10mL micro-burette – Kyle.

7.2.5 Class A 200mL volumetric flask

7.2.6 Disposable plastic 4oz Solo cups.

7.2.7 Glass 150mL beakers.

7.2.8 DI H_2O.

7.2.9 Sulfuric Acid, Standardized, 0.02000N.

7.2.10 Spike solution

7.2.10.1 Stock Alkalinity solution: Dissolve 0.848g of Na_2CO_3 (dried at 250°C for two hours and cooled in a desiccator) into 200mL DI water. This is a 4000 mg/L as $CaCO_3$ stock solution.

7.2.10.2 Spike 1mL into LCS, MS, and MSD samples. Report spike results as total alkalinity only. In LCS, MS, and MSD the spike concentration is 80 mg/L as $CaCO_3$.

8.0 Sample collection, Preservation, and Storage

8.1. Samples are to be collected in unpreserved plastic or glass containers. Samples are stored at 0-6°C until analysis.

8.2. Holding time is 14 days.

8.3. Samples should be removed from the storage refrigerator at least 2 hours prior to analysis to allow samples to equilibrate to room temperature.

9.0 Quality Control

9.1. An analytical batch consists of a MB, LCS, MS, MSD, and up to 20 samples.

9.2. See the current HEAL pH SOP for calibration procedure for pH electrode and acceptance criteria.

10.0 Procedure for 20mg/L and 2mg/L PQL alkalinities - automated

10.1. Poseidon – automated titration

10.1.1 Remove samples from refrigerator and set them out on the counter for at least two hours before analysis.

10.1.2 Follow procedure outlined in current HEAL pH SOP for instrument preparation and calibration of the pH electrode.

10.1.2.1 Make certain that the pH calibration is verified to be acceptable prior to continuing.

10.1.3 If more than 5 minutes has passed since the ICV buffer has been analyzed, i.e. the instrument has been idle due to review of pH calibration and pH ICV results, prepare, load, and analyze a pH buffer according to the current HEAL pH SOP.

10.1.3.1 The acceptance criteria for this pH buffer is listed in the current HEAL pH SOP.

10.1.4 Obtain clean 60mL auto-sampler cups.

10.1.5 Login to the *Tiamo* software using the secure login.

10.1.6 Select the Workplace icon. Click on the *Determination Series* tab.

10.1.6.1 Double click on the first line and choose the *Cond.-pH-alk 50mL* method from the scroll down menu.

10.1.6.2 Enter the HEAL sample ID on ID1 line.

10.1.6.3 Enter additional notes for sample on ID2 line.

10.1.6.4 Enter the sample position number.

10.1.6.5 Click the right scroll arrow on the bottom left of the window to advance to the next sample position.

10.1.6.5.1 Repeat 10.1.5.2 to load additional samples into list.

10.1.6.6 Click *Apply*, and then *Close* when finished entering IDs for all desired analyses.

10.1.7 After sample list has been filled appropriately, and prior to analysis, the list should be printed. The list shall be used to ensure that samples are located in appropriate positions on the auto-sampler rack.

10.1.7.1 Click on *Sample table* and select *Print (PDF)*. This will generate a temporary PDF file.

10.1.7.2 Print the PDF file. Save the PDF file to the Sample list directory located on the desktop with the file name corresponding to the analysis date.

10.1.8 Once sample list is complete and has been printed, prepare and load samples according to the sample list. Using this sequence should reduce analyst error of loading the auto-sampler incorrectly.

10.1.9 Measure 50mL of sample in a graduated cylinder. Pour this into a 60mL disposable plastic beaker. Place samples in the auto-sampler tray in the appropriate sample positions.

10.1.9.1 Always load a MB and LCS prior to sample analysis.

10.1.9.1.1 Use 50mL DI H2O for the MB.

10.1.9.1.2 Follow Section 7.1.7.2 for the spiking procedure of the LCS.

10.1.9.2 Avoid floating particulates as they can cause interferences with the electrode and causes blockages in the instrumentation plumbing.

10.1.10 Remember to load and analyze a MS and MSD for up to every twenty samples.

10.1.11 Click *Start* to begin analysis.

10.1.12 The auto-sampler rinses between each sample.

10.1.13 The aspiration line and conductivity probe on the robotic arm are rinsed three times between sample analyses.

10.1.14 Samples may require a "dilution" to prevent over flow from the titration beaker.

10.1.14.1 Use 20mL of sample instead of the routine 50mL of sample.

10.1.14.2 If over flow is still a concern, discuss with Technical Director the changing of the 0.02N H_2SO_4 to a higher normality, 0.2N H_2SO_4.

10.1.14.3 When changing to a different normality, the system must be prepped again to make certain lines are flushed. The analysis of an additional LCS is also required.

11.0 Procedure for 20mg/L and 2mg/L PQL alkalinities - manual

11.1. Memphis – manual titration with 50mL and 10mL burettes

11.1.1 Calibrate the pH meter according to current HEAL pH SOP.

11.1.1.1 All acceptance criteria listed in the current HEAL pH SOP must be meet.

11.1.2 20mg/L PQL alkalinity

11.1.2.1 Pour out old 0.02N H_2SO_4 titrant from 50mL burette and refill with fresh solution. Remove bubbles from burette tip and record volume in logbook.

11.1.2.2 For MB measure 50mL DI H_2O using graduated cylinder.

11.1.2.2.1 Add magnetic stir bar and gently stir sample.

11.1.2.2.2 Immerse pH probe in solution. Be sure the bulb (and thermocouple if present) is fully immersed in solution. Do not allow the stir bar to hit probe tip.

11.1.2.2.3 Allow pH to stabilize, record starting pH, burette volume, and titrate to pH 4.5 ±0.20 pH units. (For MB proceed very slowly, this will be a very small volume).

11.1.2.2.4 Record ending pH and burette volume.

11.1.2.3 For LCS measure 50mL DI H_2O using a graduated cylinder and pour into a clean plastic Solo cup.

11.1.2.3.1 Using the 1mL pipette, remove 1mL of DI H_2O from Solo cup, get a new tip for the pipette, then spike 1mL of Stock Alkalinity solution (this will be the LCS).

11.1.2.3.2 Add magnetic stir bar and gently stir sample.

11.1.2.3.3 Immerse pH probe in solution. Be sure the bulb (and thermocouple if present) is fully immersed in solution. Do not allow the stir bar to hit probe tip.

11.1.2.3.4 Allow pH to stabilize, record starting pH, burette volume, and titrate to pH 4.5 ±0.20 pH units.

11.1.2.3.5 Record ending pH and burette volume.

11.1.2.4 For samples, measure 50mL of un-mixed sample in graduated cylinder, and pour into plastic 4oz Solo cup.

11.1.2.4.1 Samples with very high alkalinities may be diluted prior to titration. DO NOT dilute the sample in DI H_2O. Simply titrate less sample. Be sure there is enough sample to allow pH probe tip to be completely submerged. 20mL of sample is appropriate.

11.1.2.4.2 If a sample with a very high alkalinity has not reached the pH 4.5 endpoint but the Solo cup is full, transfer Stir bar and sample to a clean 150mL beaker and continue titration to the endpoint.

11.1.2.4.3 Add stir bar, begin mixing, and immerse probe.

11.1.2.4.4 Allow pH to stabilize. Record pH.

11.1.2.4.5 If pH is above 8.30, titrate to pH 8.30 ±0.05 pH units. Record pH and burette volume.

11.1.2.4.6 Continue titration of sample until pH 4.5 end-point is reached (±0.20 pH units). Record pH and burette volume.

11.1.2.4.7 Dump sample down the drain, dispose of plastic solo cup and rinse Teflon stir-bar with DI. Continue with the next sample.

11.1.2.5 For MS and MSD samples, measure 50mL of an un-mixed sample into a 50mL graduated cylinder, and pour into 4oz solo cup.

11.1.2.5.1 Using the 1mL pipette, remove 1mL of MS sample from Solo cup, get a new pipette tip, and then spike 1mL of Stock Alkalinity solution. Begin titration immediately.

11.1.2.5.2 Follow procedure above for the titration of a LCS. Repeat with MSD sample.

11.1.3 2.0mg/L PQL (low level alkalinity)

11.1.3.1 Low level alkalinity procedure is only used when requested by client. If low level alkalinity is requested the MB and any sample with a total alkalinity <20mg/L must be titrated using low level alkalinity procedure.

11.1.3.2 Pour out old 0.02N H_2SO_4 titrant from 10mL micro burette and refill with fresh solution. Remove bubbles from burette tip and record volume in logbook.

11.1.3.3 Measure 50mL DI H_2O in graduated cylinder and pour into a Solo cup (this will be the MB).

11.1.3.4 Add magnetic stir bar and gently stir sample.

11.1.3.5 Immerse pH probe in solution. Be sure the bulb (and thermocouple if present) is fully immersed in solution. Do not allow the stir bar to hit probe tip.

11.1.3.6 Allow pH to stabilize, record starting pH, and titrate to pH 4.5 ±0.20 pH units.

11.1.3.7 Allow pH to stabilize and record ending pH and burette volume. (Be very careful, this is a very small volume).

11.1.3.8 Continue titration to reduce the pH exactly 0.30 pH units. Record final pH and burette volume.

11.1.3.9 Repeat procedure using 50 mL of sample for any client samples that have <20 mg/L CaCO3 (require less than 1 mL of titrant to reach endpoint pH 4.5)

12.0 Calculations

12.1. Blank Correction for LCS and PQL studies

12.1.1 Obtain the calculated value for the total alkalinity of the MB and subtract this from the calculated value of the LCS and the 20mg/L PQL. This is the true value.

12.1.2 Use the Low Level calculated value for the alkalinity of the MB and subtract from the 2.0mg/L PQL low level calculated value. This is the true value.

12.2. Total Alkalinity

$$Total\ Alkalinity,\ mg/L\ CaCO_3 = \frac{T \times N \times 50{,}000}{mL\ of\ sample}$$

Where:

T=Total mL of standard acid used for titration to pH 4.5 endpoint
N=Normality of standard acid

12.3. 2mg/L PQL - Low Level Alkalinity

$$Total\ Alkalinity,\ mg/L\ CaCO_3 = \frac{((2B) - C) \times N \times 50{,}000}{mL\ of\ sample}$$

Where:

B = mL titrant to first recorded pH (± 0.2 pH units from pH 4.5)
C = total mL titrant to reach 0.30 pH units lower (mL to approx. pH 4.5 plus 0.30 ph units lower)

12.4. Calculation of Alkalinity Relationships

12.4.1 Alkalinity relationships are calculated with the assumption that the total alkalinity only consists of the sum of bicarbonate, carbonate, and hydroxide alkalinities. Other weak acids that may contribute to the total alkalinity are not included in the calculation of the alkalinity relationships.

12.4.2 These relationships are determined by examining the relationship between the volume of titrant used to reach the pH 8.3 endpoint (also known as the phenolphthalein endpoint or P) and the <u>total</u> volume of titrant (including the volume used to reach pH 8.3 endpoint) used to reach the pH 4.5 endpoint (also known as T).
P = mL titrant to reach pH 8.3.
T = total volume of titrant used to reach pH 4.5.

12.4.3 In this SOP the alkalinity relationships shall be called: Bicarbonate/Carbonate, Carbonate, and Carbonate/Hydroxide.

12.4.4 The sum of bicarbonate, carbonate, and hydroxide alkalinities must always equal that of the total alkalinity calculated in 12.1.

12.4.5 Calculation of Bicarbonate/Carbonate Alkalinity

Where P = 0 or P < 1/2 T.

Hydroxide: $OH^{-}\left(mg\ CaCO_3 / L\right) = 0$

Carbonate: $CO_3^{2-}\left(mg\ CaCO_3 / L\right) = \frac{\left(2 \times P \times 50{,}000 \times N\right)}{V}$

Bicarbonate: $HCO_3^{-}\left(mg / L\ CaCO_3 / L\right) = \frac{\left(T - 2 \times P\right) \times 50{,}000 \times N}{V}$

Where:

P = mL of titrant to pH 8.3
T = Total mL of titrant to pH 4.5
N = normality of titrant
V = volume of sample in mL

12.4.6 Calculation of Carbonate Alkalinity

Where P = ½ T.

Hydroxide: $OH^{-}\left(mg\ CaCO_3 / L\right) = 0$

Carbonate: $CO_3^{2-}\left(mg\ CaCO_3 / L\right) = \frac{2 \times P \times 50{,}000 \times N}{V}$

Bicarbonate: $HCO_3^{-}\left(mg\ CaCO_3 / L\right) = 0$

Where:
P = mL of titrant to pH 8.3
N = normality of titrant
V = volume of sample in mL

12.4.7 Calculation of Carbonate/Hydroxide Alkalinity

Where P > ½ T or P = T

Hydroxide: $OH^{-}\left(mg\ CaCO_3 / L\right) = \frac{\left(2 \times P - T\right) \times 50{,}000 \times N}{V}$

Carbonate: $CO_3^{2-}(mg\ CaCO_3 / L) = \frac{2(T-P) \times 50{,}000 \times N}{V}$

Bicarbonate: $HCO_3^{-}(mg\ CaCO_3 / L) = 0$

Where:
P = mL of titrant to pH 8.3
T = Total mL of titrant to pH 4.5
N = normality of titrant
V = volume of sample in mL

13.0 Method Performance

13.1. PT or "blind study" samples are routinely analyzed to check the accuracy of the total alkalinity analysis.

13.2. PQL studies are done at a minimum of yearly.

13.3. Control Charting of LCS recoveries shall be done for all LCS analyses.

14.0 Estimation of Uncertainty

14.1. Estimation of uncertainty is to be done on an annual basis.

14.1.1 Collect, at least, 20 of the most recent LCS data points. Calculate the standard deviation (s) and the average of all the readings. Calculate the 95% confidence interval (2 times the standard deviation). Report the estimated uncertainty as ±2(s). Uncertainty data is kept on file in the method folders in the quality control office. See current revision of QAP for the formulas required to calculate uncertainty.

15.0 Demonstration of Capability (DOC)

15.1. Demonstration of Capability (DOC) samples are analyzed before an analyst is able to independently generate data.

15.2. DOCs for total alkalinity consist of four replicate spikes.

15.3. A detailed description of DOCs can be found in the HEAL QA Plan.

15.4. See the current HEAL QAP for necessary calculation and statistical requirements for DOC studies.

15.5. Annual Documentation of Continued Proficiency (ADOCP) is performed annually after acceptable DOC has been established. See HEAL Quality Assurance Plan.

16.0 Pollution Prevention

16.1. Standard 0.02N H_2SO_4 titrant comes pre-made and standardized. No preparation or standardization is necessary.

16.2. The automated titration system uses more pH buffer volume. However, using this system currently allows for at least three determinations to be done on a single aliquot of sample.

17.0 Data Assessment and Acceptance Criteria for Quality Control Measures

17.1. See **Appendix A** for a summary of batch QC requirements.

17.2. Total alkalinity value can be found on the generated report. The software calculates all necessary values. These values are based on a 50mL sample size. If a different sample volume was used, the value must be corrected.

17.3. pH Endpoints

17.3.1 The acceptable range for pH 8.3 endpoint is ±0.05 pH units.

17.3.2 The acceptable range for pH 4.5 endpoint is ±0.20 pH units

17.3.3 The acceptable range for additional 0.30 pH units for low level alkalinities is ±0.05 pH units.

17.4. Batch QC

17.4.1 The MB must calculate to ≤20mg/L when reporting to the 20mg/L PQL.

17.4.1.1Use the *Total Alk* value from the report.

17.4.2 The MB must calculate to ≤2mg/L when reporting to the 2.0mg/L PQL.

17.4.2.1Use the *Low Level Titration* value from the report.

17.4.3 The LCS must be recovered to within ±20% of the assigned value. The LCS must be blank corrected to represent the true value.

17.4.4 The MS and MSD must be recovered to within ±20% of the assigned value.

17.4.5 The RPD for the MSD must be ≤20%.

18.0 Corrective Actions for Out-of-Control Data

18.1. pH Endpoint

18.1.1 If sample is titrated to a pH greater than the required endpoint, add additional titrant to bring pH into the acceptable range. If the sample has been titrated to less than the required endpoint (pH overshot), the sample must be discarded and re-titrated.

18.1.2 If the MB calculates >2 mg $CaCO_3$, reanalyze using fresh DI from a different location (i.e. any DI pump dispenser or wash bottle containing DI) when the 2.0mg/L PQL is required. This may be an indication that the DI water system may not be producing ultra-pure water. Notify the Inorganic manager immediately. If MB continues to fail discontinue analysis until to source of the problem is fixed.

18.2. Batch QC

18.2.1 If the LCS fails, re-spike and reanalyze. If the spike continues to fail, remake stock spiking solution, spike and reanalyze. If it continues to fail discontinue analysis until the source of the problem can be fixed. The LCS value must be blank corrected.

18.2.2 If MS or MSD fails, re-spike and reanalyze. If spike continues to fail, flag data and report.

18.2.3 If RPD for MSD fails. Re-spike and reanalyze both MS and MSD, report reanalysis. If RPD is still out flag data and report.

19.0 Contingencies for Handling Out of Control Data

19.1. All out of control data is flagged and reported to the client.

19.2. Corrective actions shall be filled out when there are deviations from the method or this SOP.

20.0 Waste Management

20.1. Analytical results are used to characterize their respective sample contamination level(s) so that the proper disposal can be performed. These wastes will be disposed of according to their hazard as well as their type and level of contamination. Refer to the Hall Environmental Analysis Laboratory Chemical Hygiene Plan for details regarding waste disposal.

21.0 References

21.1. Standard Methods 20th Edition, Method 2320B.

21.2. Quality Assurance Plan, HEAL.

21.3. Chemical Hygiene Plan, HEAL.

21.4. Health and Safety SOP, HEAL.

21.5. Tiamo Software, Online Help, Metrohm.

Appendix A

Standard Methods 2320B: Total Alkalinity, Carbonate, Bicarbonate, and Hydroxide in Aqueous Samples
Corrective Action Procedures

Analytical Method	Parameter	QC Element	Frequency	Acceptance Criteria	Corrective Action
SM2320B	Wet Chemistry	pH Endpoint (8.30)	every sample where starting pH is >8.3 with exception of the LCS, MS, MSD	±0.05 pH units	1. If pH is "overshot" sample must be re-titrated
		pH Endpoint (4.50)	every sample, including QC	±0.20 pH units	1. If pH is "overshot" sample must be re-titrated
		Reduce 0.30 pH units lower	low level alkalinity MB and low level alkalinity samples only	±0.05 pH units	1. Record pH, retitrate sample if possible
		Method Blank	1 per preparation/analysis batch (≤ 20 samples)	≤ PQL	1. Reanalyze using fresh DI from a different source. 2. Discontinue analysis.
		Laboratory Control Sample (LCS)	1 per preparation/analysis batch (≤ 20 samples)	% Recovery ±20% of actual value	1. Re-spike and reanalyze. 2. Remake spike solution. Respike and reanalyze. 3. Discontinue analysis.
		Matrix Spike (MS)	1 per preparation/analysis batch (≤ 20 samples)	% Recovery ±20% of actual value	1. Re-spike and reanalyze. 2. Flag data and report.
		Matrix Spike Duplicate (MSD)	1 per preparation/analysis batch (≤ 20 samples)	% Recovery ±20% of actual value Relative percent difference (RPD) ≤20%	1. MSD recovery: Re-spike and reanalyze. 2. MSD recovery: Flag data and report. 3. RPD: Re-spike and reanalyze both MS and MSD, report reanalysis. 4. RPD: Flag data and report.

Appendix B.
Comparison Data. Automated titration using Poseidon versus Manual titration.

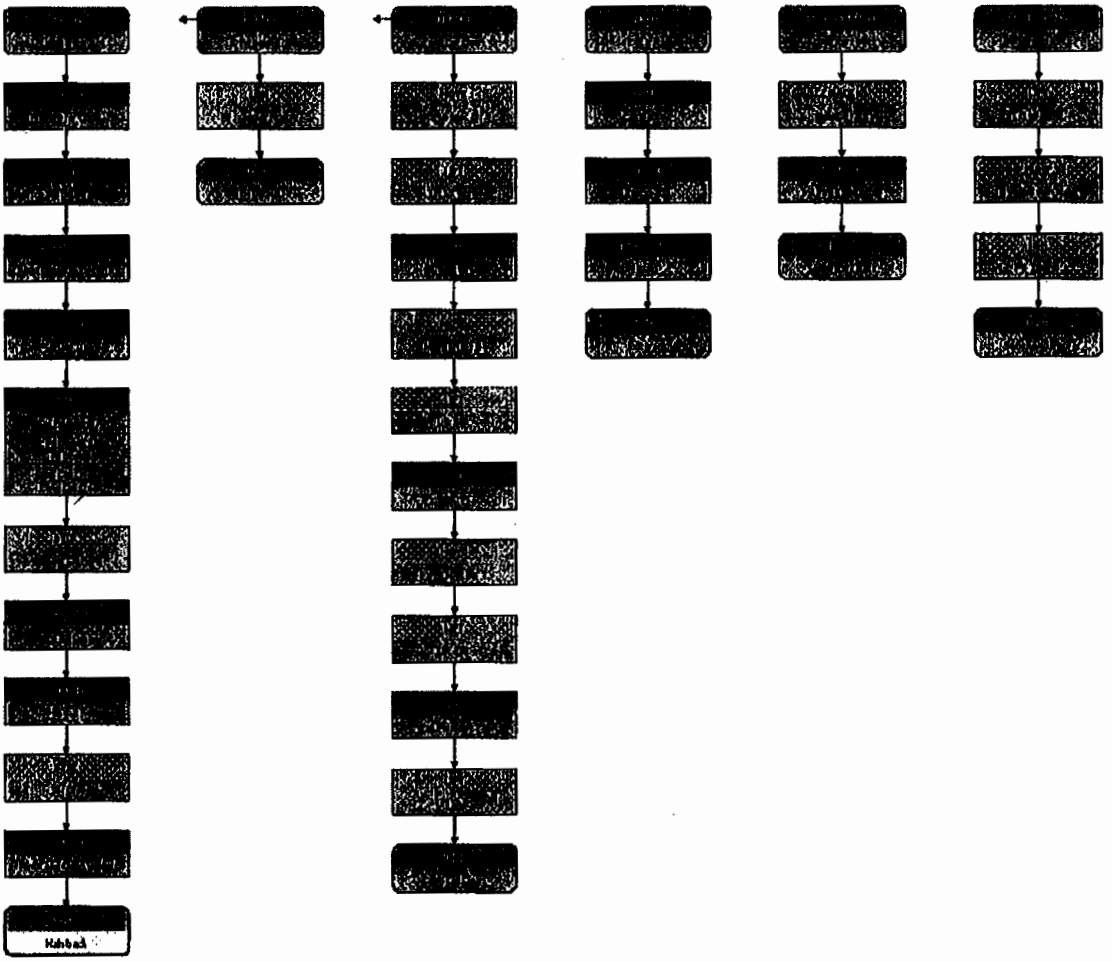
		Poseidon		Burette		%RPD
Sample 1		64.17		63		1.84%
Sample 2		424.12		433		2.07%
Sample 3		419.32		433		3.21%
Sample 4		346.12		374		7.74%
Sample 5		536.48		554		3.21%
Sample 6		521.73		542		3.81%
Sample 7		132.02		136		2.97%
Sample 8		148.47		153		3.01%
Sample 9		29.63		32		7.69%
Sample 10		14.82		15		1.21%
Sample 11		119.83		128		6.59%
Sample 12		120.41		127		5.33%
Sample 13		699.30		742		5.93%
Sample 14		246.61		258		4.51%
Sample 15		170.92		180		5.17%
Sample 16		10.12		12		17.00%
Sample 17		64.20		56		13.64%
Sample 18		91.00		95		4.30%
Sample 19		185.53		190		2.38%
Sample 20		493.20		544		9.80%

Appendix C.

Software Method Flow Chart
Software Method Parameters
Software Method Calculations

Method run

Method name: Cond.-pH-Alk- 50 mL updated
Method version: 5
Method group: Main group



Method parameters

Method ..... Cond.-pH-Alk- 50 mL updated
Method saving date 2009-02-03 14:48:03 UTC-7
Method version 5
Method group ..... Main group
Method status ..... original
Method saved by (full name) ..... Benjamin Derek Harmon
Method saved by (short name) ..... bdh

START Main track
General
Workplace view
Current view ..... on
Track view for live window
Live display 1 ..... Main track
Live display 2 ..... Main track
Statistics ..... off
Conditioning
Automatic conditioning ..... off
Application note

Method variables

Name	Type	Assignment	Value	Comment	Monitoring
Sample size	Number	Sample size		Sample size	off
Sample size unit	Text	Sample size unit		Sample size unit	off
Sample position	Number	Sample position		Sample position number	off
ID1	Text	ID1		Sample identification 1	off
ID2	Text	ID2		Sample identification 2	off
ID3	Text	ID3		Sample identification 3	off
cond	Text				off
temp	Text				off

Name ..... Sample size
Type ..... Number
Assignment ..... on ..... Sample size
Value ..... off .....
Check at start ..... off
Comment ..... Sample size
Variable monitoring ..... off
Lower limit .....
Upper limit .....
Message .....
Display message ..... on
Record message ..... on

Message by e-mail ..... off
Mail to .....
Subject ..... Message from tiamo - Method 'New method 2' - Command 'Main track'
User .....
Mail from .....
SMTP Server .....
POP3 Server .....
Acoustic signal ..... off
Action ..... off
Stop determination ..... on
Stop determination and series ..... off

Name ..... Sample position
Type ..... Number
Assignment ..... on ..... Sample position
Value ..... off .....
Check at start ..... on
Comment ..... Sample position number
Variable monitoring ..... off
Lower limit .....
Upper limit .....
Message .....
Display message ..... on
Record message ..... on
Message by e-mail ..... off
Mail to .....
Subject ..... Message from tiamo - Method 'New method 2' - Command 'Main track'
User .....
Mail from .....
SMTP Server .....
POP3 Server .....
Acoustic signal ..... off
Action ..... off
Stop determination ..... on
Stop determination and series ..... off

Name ..... Sample size unit
Type ..... Text
Assignment ..... on ..... Sample size unit
Value ..... off .....
Check at start ..... off
Comment ..... Sample size unit

Name ID1
Type ..... Text

Assignment ..... on ID1
Value ..... off .....
Check at start ..... on
Comment ..... Sample identification 1

Name ID2
Type ..... Text
Assignment ..... on ID2
Value ..... off .....
Check at start ..... off
Comment ..... Sample identification 2

Name ID3
Type ..... Text
Assignment ..... on ID3
Value ..... off .....
Check at start ..... off
Comment ..... Sample identification 3

Name ..... cond
Type ..... Text
Assignment ..... off .....
Value ..... on .....
Check at start ..... off
Comment .....

Name ..... temp
Type ..... Text
Assignment ..... off .....
Value ..... on .....
Check at start ..... off
Comment .....

SEQUENCE Prepare for Cond

MOVE Move to Sample Pos

Device

Device name ..... 855_1

Target

Tower 1

Move ..... Sample position

Beaker test

Display message ..... on

Stop determination ..... off

Stop determination and series ..... off

Parameters

Shift rate ..... 20 °/s

Shift direction ..... auto

Swing rate ..... 55 °/s

LIFT **Lift Down**

Device
- Device name ..... 855_1

Target
- Tower 1
- Lift position ..... Work position mm

Parameters
- Lift rate ..... 25 mm/s

LIFT **Lift Up**

Device
- Device name ..... 855_1

Target
- Tower 1
- Lift position ..... Rinse position mm

Parameters
- Lift rate ..... 25 mm/s

LIFT **Lift Work**

Device
- Device name ..... 855_1

Target
- Tower 1
- Lift position ..... Work position mm

Parameters
- Lift rate ..... 25 mm/s

WAIT **30s pause**

Wait
- Stop track and waiting for [Continue] ..... off
- Stop all tracks and waiting for [Continue] ..... off
- Waiting time ..... on
  - Time 30
  - Unit ..... s

Message
- Record message ..... off
- Message by e-mail ..... off
- Acoustic signal ..... off

TRANSFER **Read conductivity-temp**

Device
- Device name 712

Transfer commands

Name	Action	Command	Variable	Comparison
conductivity	Read	&l.a.m.o$q	MV.cond	(\d+\.\d+E.\d+)

Name	Action	Command	Variable	Comparison
temp	Read	&l.a.m.t$q	MV.temp	(\d+\.\d+)

Timeout ..... off

SEQUENCE 50mL Sample Transfer

LQH Fill Dosino with Sample

General/Hardware

Device

Device name ..... 855_1

Dosing device

Dosing device 1

Solution ..... Transfer Pipette

Parameters

Function ..... Aspirate

Port 1

Volume ..... 50.000 mL

Rate ..... 100.0 mL/min

WAIT Wait 3

Wait

Stop track and waiting for [Continue] ..... off

Stop all tracks and waiting for [Continue] ..... off

Waiting time ..... on

Time 3

Unit ..... s

Message

Record message ..... off

Message by e-mail ..... off

Acoustic signal ..... off

LIFT Lift to Shift

Device

Device name ..... 855_1

Target

Tower 1

Lift position ..... Shift position mm

Parameters

Lift rate ..... 25 mm/s

LQH Dispense Sample

General/Hardware

Device

Device name ..... 855_1

Dosing device

Dosing device 1

Solution ..... Transfer Pipette

Parameters

Function ..... Eject to end volume
Port 3
Rate ..... maximum mL/min

LQH **Get remaining sample**

General/Hardware

Device

Device name ..... 855_1

Dosing device

Dosing device 1
Solution ..... Transfer Pipette

Parameters

Function ..... Aspirate
Port 1
Volume ..... 15.00 mL
Rate ..... 100.0 mL/min

LQH **Dispense remaining sample**

General/Hardware

Device

Device name ..... 855_1

Dosing device

Dosing device 1
Solution ..... Transfer Pipette

Parameters

Function ..... Eject to end volume
Port 3
Rate ..... maximum mL/min

CALL **Call Fill Dosino**

Call text	Track name	Condition	Condition
Fill Dosino	Fill Dosino	off	
Cond Cell Clean	Cond Cell Clean	off	

STIR **Stir On**

Device

Device name ..... 855_1

Stirrer

Stirrer 3
Stirrer type ..... 801 Magnetic stirrer
Stirring rate 15

Action

Switch on ..... on
Switch off ..... off
Duration ..... off

MEAS pH pH

General/Hardware

Device

Device name ..... 855_1

Sensor

Measuring input 1

Sensor ..... pH electrode

Temperature measurement ..... automatic

Stirrer

Stirrer ..... off

Measuring parameters

Measurement

Measurement with drift control ..... on

Signal drift ..... 2.0 mV/min

Min. waiting time ..... 20 s

Max. waiting time ..... 60 s

Measuring interval ..... 1 s

Stop measured value pH ..... off

Measurement without drift control ..... off

Temperature

Temperature ..... 25.0 °C

Evaluations

Fix end point evaluation ..... off

Minimum evaluation ..... off

Maximum evaluation ..... off

Break point evaluation ..... off

Additional measured values

Additional calculated measured values ..... off

Additional external measured values ..... off

SET pH to 8-3

General/Hardware

Device

Device name ..... 855_1

Dosing device

Dosing device 2

Solution H2SO4

Sensor

Measuring input 1

Sensor ..... pH electrode

Temperature measurement ..... automatic

Stirrer

Stirrer ..... off

Start conditions

Initial measured value

Signal drift ..... off mV/min

Min. waiting time ..... 0 s
Max. waiting time ..... 1 s
Pause 1
Pause 1 ..... 0 s
Start volume
Start volume ..... 0 mL
Dosing rate ..... maximum mL/min
Pause 2
Pause 2 ..... 0 s
Control parameters
End point 1
EP1 at pH 8.3
Titration rate
Titration rate ..... user
Control
Dynamics pH 4.000
Max. rate ..... 8.0 mL/min
Min. rate ..... 5.0 µL/min
Stop criterion
Stop criterion ..... drift
Stop drift ..... 20 µL/min
End point 2 ..... on
EP2 at pH 4.2
Titration rate
Titration rate ..... user
Control
Dynamics pH 4.0
Max. rate ..... 10.00 mL/min
Min. rate ..... 25.0 µL/min
Stop criterion
Stop criterion ..... drift
Stop drift ..... 20 µL/min
Titration parameters
Titration direction ..... -
Extraction time ..... 0 s
Temperature ..... 25.0 °C
Time interval measuring point ..... 1 s
Stop conditions
Stop volume ..... 60.00 mL
Stop time ..... off s
Filling rate ..... maximum mL/min
Conditioning
Conditioning ..... off
Additional evaluations
Fix end point evaluation ..... on
Fixed quantity ..... Measured value

Fix EP1 at pH 4.5
Fix EP2 at pH ..... off
Fix EP3 at pH ..... off
Minimum evaluation ..... off
Maximum evaluation ..... off
Additional measured values
Additional calculated measured values ..... off
Additional external measured values ..... off

STIR Stir Off
Device
Device name ..... 855_1
Stirrer
Stirrer 3
Stirrer type ..... 801 Magnetic stirrer
Stirring rate 8
Action
Switch on ..... off
Switch off ..... on
Duration ..... off

SEQUENCE Rinse and Clean Vessel

PUMP Asp Pump On
Device
Device name ..... 855_1
Pumps
Tower 1
Pump(s) 2
Action
Switch on ..... on
Switch off ..... off
Duration ..... off

WAIT Wait 12s
Wait
Stop track and waiting for [Continue] ..... off
Stop all tracks and waiting for [Continue] ..... off
Waiting time ..... on
Time 12
Unit ..... s
Message
Record message ..... off
Message by e-mail ..... off
Acoustic signal ..... off

MOVE Move Spec 5
Device
Device name ..... 855_1

Target
Tower 1
Move ..... Special beaker
Number 5
Beaker test
Display message ..... on
Stop determination ..... off
Stop determination and series ..... off
Parameters
Shift rate ..... 20 °/s
Shift direction ..... auto
Swing rate ..... 55 °/s

LQH Clean Port 1 Tubing
General/Hardware
Device
Device name ..... 855_1
Dosing device
Dosing device 1
Solution ..... Transfer Pipette
Parameters
Function ..... Dosing
Port 1
Volume ..... 3.25 mL
Rate ..... maximum mL/min

LQH Clean port 3 tubing
General/Hardware
Device
Device name ..... 855_1
Dosing device
Dosing device 1
Solution ..... Transfer Pipette
Parameters
Function ..... Eject to end volume
Port 3
Rate ..... maximum mL/min

LIFT Shift Pos
Device
Device name ..... 855_1
Target
Tower 1
Lift position ..... Shift position mm
Parameters
Lift rate ..... 25 mm/s

LQH Fill with air

General/Hardware
Device
Device name ..... 855_1
Dosing device
Dosing device 1
Solution ..... Transfer Pipette
Parameters
Function ..... Aspirate
Port 1
Volume ..... 15.00 mL
Rate ..... maximum mL/min

LQH Dispense Air
General/Hardware
Device
Device name ..... 855_1
Dosing device
Dosing device 1
Solution ..... Transfer Pipette
Parameters
Function ..... Eject to end volume
Port 3
Rate ..... maximum mL/min

PUMP Rinse Pump-On
Device
Device name ..... 855_1
Pumps
Tower 1
Pump(s) 1
Action
Switch on ..... off
Switch off ..... off
Duration ..... on
Time ..... 4.0 s

WAIT Wait 7s
Wait
Stop track and waiting for [Continue] ..... off
Stop all tracks and waiting for [Continue] ..... off
Waiting time ..... on
Time 7
Unit ..... s
Message
Record message ..... off
Message by e-mail ..... off
Acoustic signal ..... off

PUMP Pump on
Device
Device name ..... 855_1
Pumps
Tower 1
Pump(s) 1
Action
Switch on ..... off
Switch off ..... off
Duration ..... on
Time ..... 4.0 s

WAIT Wait 10s
Wait
Stop track and waiting for [Continue] ..... off
Stop all tracks and waiting for [Continue] ..... off
Waiting time ..... on
Time 10
Unit ..... s
Message
Record message ..... off
Message by e-mail ..... off
Acoustic signal ..... off

PUMP Pump Off
Device
Device name ..... 855_1
Pumps
Tower 1
Pump(s) 2
Action
Switch on ..... off
Switch off ..... on
Duration ..... off

TRACK Fill Dosino
Return immediately ..... on
Delete all data ..... off

LQH Fill Transfer Pipette
General/Hardware
Device
Device name ..... 855_1
Dosing device
Dosing device 1
Solution ..... Transfer Pipette
Parameters
Function ..... Fill

Port 2
Rate . . . . . maximum mL/min

TRACK **Cond Cell Clean**
Return immediately . . . . . on
Delete all data . . . . . off

MOVE **Move to Spec Bkr 9**
Device
Device name . . . . . 855_1
Target
Tower 1
Move . . . . . Special beaker
Number 9
Beaker test
Display message . . . . . on
Stop determination . . . . . off
Stop determination and series . . . . . off
Parameters
Shift rate . . . . . 20 °/s
Shift direction . . . . . auto
Swing rate . . . . . 55 °/s

LIFT **Lift to work**
Device
Device name . . . . . 855_1
Target
Tower 1
Lift position . . . . . 218 mm
Parameters
Lift rate . . . . . 25 mm/s

WAIT **Wait 5s**
Wait
Stop track and waiting for [Continue] . . . . . off
Stop all tracks and waiting for [Continue] . . . . . off
Waiting time . . . . . on
Time 5
Unit . . . . . s
Message
Record message . . . . . off
Message by e-mail . . . . . off
Acoustic signal . . . . . off

MOVE **Move SP Bkr 8**
Device
Device name . . . . . 855_1
Target
Tower 1

Move ..... Special beaker
Number 8
Beaker test
Display message ..... on
Stop determination ..... off
Stop determination and series ..... off
Parameters
Shift rate ..... 20 °/s
Shift direction ..... auto
Swing rate ..... 55 °/s

LIFT Lift 226
Device
Device name ..... 855_1
Target
Tower 1
Lift position ..... 218 mm
Parameters
Lift rate ..... 25 mm/s

WAIT Wait 5s
Wait
Stop track and waiting for [Continue] ..... off
Stop all tracks and waiting for [Continue] ..... off
Waiting time ..... on
Time 5
Unit ..... s
Message
Record message ..... off
Message by e-mail ..... off
Acoustic signal ..... off

MOVE Move Spec Bkr 7
Device
Device name ..... 855_1
Target
Tower 1
Move ..... Special beaker
Number 7
Beaker test
Display message ..... on
Stop determination ..... off
Stop determination and series ..... off
Parameters
Shift rate ..... 20 °/s
Shift direction ..... auto
Swing rate ..... 55 °/s

Result name	Formula	Unit	Decimal places	Assignment	Statistics
CO3	= (2 * Case('to 8-3.EP{1}.VOL' ; 'to 8-3.EP{1}.VOL' ;0;0) * 50000 * 0.02) / 50	mg/L	2	RS09	off
HCO3 raw	= (('to 8-3.FP{1}.VOL' - (2 * Case('to 8-3.EP{1}.VOL' ; 'to 8-3.EP{1}.VOL' ;0;0))) * 50000 * 0.02) / 50	mg/L	2	none	off
HCO3	=Case('RS.HCO3 raw'<0;0;'RS.HCO3 raw')	mg/L	2	RS10	off
Low Level Titration	= (((2 * 'to 8-3.FP{1}.VOL') - 'to 8-3.EP{2}.VOL') * 0.02 * 50000) / 50	mg/L	2	RS12	off
est TDS	=0.66 * 'RS.Cond'	mg/L	0	RS13	off
Free CO2/L	= 2.0 * 'RS.HCO3' * (10 ^ (6- 'RS.pH'))	mg/L	2	RS19	off
Total CO2/L	= 'RS.Free CO2/L' + (.44 * ((2 * 'RS.HCO3') + 'RS.CO3'))	mg/L	2	RS20	off
mLs 8-3 to 4-5 (mLs 4-5 excel)	= 'RS.mLs to 4-5' - 'RS.mLs to 8-3'	mL	2	RS21	off

Result name ..... Cond
Formula ..... =1000000 * TextToNumber('MV.cond')
Unit ..... µS/cm
Decimal places 2
Assignment RS01
Statistics ..... off
Description .....
Result monitoring ..... off
Save result as common variable ..... off
Name .....
Save result as titer ..... off
Solution name .....

Result name ..... Cond Temp
Formula ..... =TextToNumber('MV.temp')
Unit ..... °C
Decimal places 1
Assignment RS02
Statistics ..... off
Description .....
Result monitoring ..... off
Save result as common variable ..... off
Name .....
Save result as titer ..... off
Solution name .....

Result name ..... pH
Formula ..... = 'pH.EME'
Unit ..... pH units
Decimal places 2

Assignment RS03
Statistics . . . . . off
Description . . . . . RS.'Result name'[.VAL] Result value.
Result monitoring . . . . . off
Save result as common variable . . . . . off
Name . . . . . .
Save result as titer . . . . . off
Solution name . . . . . .

Result name . . . . . pH Temp
Formula . . . . . = 'pH.ETE'
Unit . . . . . °C
Decimal places 1
Assignment RS04
Statistics . . . . . off
Description . . . . . .
Result monitoring . . . . . off
Save result as common variable . . . . . off
Name . . . . . .
Save result as titer . . . . . off
Solution name . . . . . .

Result name . . . . . mLs to 8-3
Formula . . . . . = Case('to 8-3.EP(1).VOL' ; 'to 8-3.EP(1).VOL' ;0;0)
Unit . . . . . mL
Decimal places 2
Assignment RS05
Statistics . . . . . off
Description . . . . . .
Result monitoring . . . . . off
Save result as common variable . . . . . off
Name . . . . . .
Save result as titer . . . . . off
Solution name . . . . . .

Result name . . . . . mLs to 4-5
Formula . . . . . = 'to 8-3.FP(1).VOL'
Unit . . . . . mL
Decimal places 2
Assignment RS06
Statistics . . . . . off
Description . . . . . .
Result monitoring . . . . . off
Save result as common variable . . . . . off
Name . . . . . .
Save result as titer . . . . . off

Solution name . . . . . .

Result name . . . . . mLs to 4-2
Formula . . . . . = 'to 8-3.EP(2).VOL'
Unit . . . . . mL
Decimal places 2
Assignment RS11
Statistics . . . . . off
Description . . . . . .
Result monitoring . . . . . off
Save result as common variable . . . . . off
Name . . . . . .
Save result as titer . . . . . off
Solution name . . . . . .

Result name . . . . . Total Alk
Formula . . . . . = ('to 8-3.FP(1).VOL' * 0.02 * 50000) / 50
Unit . . . . . mg/L
Decimal places 2
Assignment RS07
Statistics . . . . . off
Description . . . . . RS.'Result name'[.VAL] Result value.
Result monitoring . . . . . off
Save result as common variable . . . . . off
Name . . . . . .
Save result as titer . . . . . off
Solution name . . . . . .

Result name . . . . . OH raw
Formula . . . . . = (((2 * Case('to 8-3.EP(1).VOL' ; 'to 8-3.EP(1).VOL' ;0;0)) - 'to 8-3.FP(1).VOL') * 50000 * .02) / 50
Unit . . . . . mg/L
Decimal places 2
Assignment . . . . . none
Statistics . . . . . off
Description . . . . . Calculates OH value from (2P-t)*50000*N/V for any P not equal to zero.
Result monitoring . . . . . off
Save result as common variable . . . . . off
Name . . . . . .
Save result as titer . . . . . off
Solution name . . . . . .

Result name . . . . . OH
Formula . . . . . =Case('RS.OH raw'< 0;0;'RS.OH raw')
Unit . . . . . mg/L

Decimal places 2
Assignment RS09
Statistics . . . . . off
Description . . . . . If 'OH raw' is less than zero (negative number), then 'OH' equals 0. If 'OH raw' is zero or positive, then 'OH' result is equal to "OH raw' value.
Result monitoring . . . . . off
Save result as common variable . . . . . off
Name . . . . . .
Save result as titer . . . . . off
Solution name . . . . . .

Result name CO3
Formula . . . . . = (2 * Case('to 8-3.EP{1}.VOL' ; 'to 8-3.EP{1}.VOL' ;0;0) * 50000 * 0.02) / 50
Unit . . . . . mg/L
Decimal places 2
Assignment RS09
Statistics . . . . . off
Description . . . . . .
Result monitoring . . . . . off
Save result as common variable . . . . . off
Name . . . . . .
Save result as titer . . . . . off
Solution name . . . . . .

Result name . . . . . HCO3 raw
Formula . . . . . = (('to 8-3.FP{1}.VOL' - (2 * Case('to 8-3.EP{1}.VOL' ; 'to 8-3.EP{1}.VOL' ;0;0))) * 50000 * 0.02) / 50
Unit . . . . . mg/L
Decimal places 2
Assignment . . . . . none
Statistics . . . . . off
Description . . . . . Calculates HCO3 value from (T-2P)*50000*N/V for any P not equal to zero.
Result monitoring . . . . . off
Save result as common variable . . . . . off
Name . . . . . .
Save result as titer . . . . . off
Solution name . . . . . .

Result name HCO3
Formula . . . . . =Case('RS.HCO3 raw'<0;0;'RS.HCO3 raw')
Unit . . . . . mg/L
Decimal places 2
Assignment RS10
Statistics . . . . . off

Description . . . . . If 'HCO3 raw' is less than zero (negative number), then 'HCO3' equals 0. If 'HCO3 raw' is zero or positive, then 'HCO3' result is equal to 'HCO3 raw'.

Result monitoring . . . . . off
Save result as common variable . . . . . off
Name . . . . . .
Save result as titer . . . . . off
Solution name . . . . . .

Result name . . . . . Low Level Titration
Formula . . . . . = (((2 * 'to 8-3.FP(1).VOL') - 'to 8-3.EP(2).VOL') * 0.02 * 50000) / 50
Unit . . . . . mg/L
Decimal places 2
Assignment RS12
Statistics . . . . . off
Description . . . . . .
Result monitoring . . . . . off
Save result as common variable . . . . . off
Name . . . . . .
Save result as titer . . . . . off
Solution name . . . . . .

Result name . . . . . est TDS
Formula . . . . . =0.66 * 'RS.Cond'
Unit . . . . . mg/L
Decimal places 0
Assignment RS13
Statistics . . . . . off
Description . . . . . estimated Total Dissolved Solids (TDS) from Electical Conductivity (EC)
Result monitoring . . . . . off
Save result as common variable . . . . . off
Name . . . . . .
Save result as titer . . . . . off
Solution name . . . . . .

Result name . . . . . Free CO2/L
Formula . . . . . = 2.0 * 'RS.HCO3' * (10 ^ (6- 'RS.pH'))
Unit . . . . . mg/L
Decimal places 2
Assignment RS19
Statistics . . . . . off
Description . . . . . .
Result monitoring . . . . . off
Save result as common variable . . . . . off
Name . . . . . .

Save result as titer ..... off
Solution name .....

Result name ..... Total CO2/L
Formula ..... = 'RS.Free CO2/L' + (.44 * ((2 * 'RS.HCO3') + 'RS.CO3'))
Unit ..... mg/L
Decimal places 2
Assignment RS20
Statistics ..... off
Description .....
Result monitoring ..... off
Save result as common variable ..... off
Name .....
Save result as titer ..... off
Solution name .....

Result name ..... mLs 8-3 to 4-5 (mLs 4-5 excel)
Formula ..... = 'RS.mLs to 4-5' - 'RS.mLs to 8-3'
Unit ..... mL
Decimal places 2
Assignment RS21
Statistics ..... off
Description ..... total mL to 4.5 - mL to 8.3. This gives you the amt. of titrant used to get from 8.3 to 4.5 pH units. This value can be plugged into the excel sheet as a double check.
Result monitoring ..... off
Save result as common variable ..... off
Name .....
Save result as titer ..... off
Solution name .....

REPORT Report 233
Report template
Report template ..... New Results Report
Report output
Printer ..... on
Printer ..... Default printer
PDF file ..... off

DATABASE tiamo

Database

Results

SERIES START Series start track

RACK Initialize rack

Device
Device name ..... 855_1
Rack test ..... off

SEQUENCE Init Rinse Sequence

PUMP Pump On
Device
Device name ..... 855_1
Pumps
Tower 1
Pump(s) 2
Action
Switch on ..... on
Switch off ..... off
Duration ..... off

WAIT Wait 8s
Wait
Stop track and waiting for [Continue] ..... off
Stop all tracks and waiting for [Continue] ..... off
Waiting time ..... on
Time 8
Unit ..... s
Message
Record message ..... off
Message by e-mail ..... off
Acoustic signal ..... off

LQH Make room in Dosino
General/Hardware
Device
Device name ..... 855_1
Dosing device
Dosing device 1
Solution ..... Transfer Pipette
Parameters
Function ..... Eject to end volume
Port 3
Rate ..... maximum mL/min

LQH Fill w- air
General/Hardware
Device
Device name ..... 855_1
Dosing device
Dosing device 1
Solution ..... Transfer Pipette
Parameters

Function ..... Aspirate
Port 1
Volume ..... 15.00 mL
Rate ..... maximum mL/min

LQH Eject to EV

General/Hardware

Device

Device name ..... 855_1

Dosing device

Dosing device 1
Solution ..... Transfer Pipette

Parameters

Function ..... Eject to end volume
Port 3
Rate ..... maximum mL/min

WAIT Wait 3s

Wait

Stop track and waiting for [Continue] ..... off
Stop all tracks and waiting for [Continue] ..... off
Waiting time ..... on
Time 3
Unit ..... s

Message

Record message ..... off
Message by e-mail ..... off
Acoustic signal ..... off

PUMP Rinse 3s

Device

Device name ..... 855_1

Pumps

Tower 1
Pump(s) 1

Action

Switch on ..... off
Switch off ..... off
Duration ..... on
Time ..... 3.0 s

WAIT Wait 8 s

Wait

Stop track and waiting for [Continue] ..... off
Stop all tracks and waiting for [Continue] ..... off
Waiting time ..... on
Time 8
Unit ..... s

Message
Record message ..... off
Message by e-mail ..... off
Acoustic signal ..... off

PUMP Asp Pump Off
Device
Device name ..... 855_1
Pumps
Tower 1
Pump(s) 2
Action
Switch on ..... off
Switch off ..... on
Duration ..... off

SERIES END Series end track

LIFT Lift to Home
Device
Device name ..... 855_1
Target
Tower 1
Lift position ..... Home position mm
Parameters
Lift rate ..... 25 mm/s

PUMP Fill Vessel with water
Device
Device name ..... 855_1
Pumps
Tower 1
Pump(s) 1
Action
Switch on ..... off
Switch off ..... off
Duration ..... on
Time ..... 4.0 s

LQH Fill Dosino
General/Hardware
Device
Device name ..... 855_1
Dosing device
Dosing device 1
Solution ..... Transfer Pipette
Parameters
Function ..... Fill

Port 2
Rate. . . . . maximum mL/min

Hall Environmental Analysis Laboratory

Common Name: Standard Methods 2320B	S.O.P. Number: S-SM2320-6
Method: Automated and Manual Methods for Total Alkalinity, Low Level Alkalinity, Carbonate, Bicarbonate, and Hydroxide in Aqueous Samples	
Effective Date: August 29, 2011	Supersedes: S-SM2320-5
Laboratory Manager	Quality Assurance/Quality Control Officer:

1.0 Scope and Application

1.1. This SOP describes the procedure for determining the total alkalinity and its three common forms found in most waters (carbonate, bicarbonate, and hydroxide). This SOP is applicable to drinking, surface, and saline waters, domestic and industrial wastes.

1.2. There are two processes that can be used for alkalinity determination.

1.2.1 Titration of alkalinity by the automated system, Poseidon.

1.2.1.1 The automated titration system is the preferred procedure for the determination of alkalinity. Before use of the automated system, the analyst must have read and understood this SOP and the current pH and eC SOPs within proven demonstration of capability.

1.2.1.1.1 This SOP relies heavily on information already included in the current pH SOP. Any change in the current pH SOP must be adhered to when using this SOP.

1.2.1.2 Appendix B contains a comparison study of automated titration versus manual titration. This is included to show that there is no significant difference between the two procedures.

1.2.2 Manual titration using a 50mL and or 10mL micro-burette, and Memphis a pH meter.

1.2.2.1 The manual titration system is used as a back up system should the auto-titrator system be temporarily out of service, or if analyzing samples that have a difficult matrix with an extreme buffering capacity.

1.3. This SOP also includes the procedure for determining low level alkalinities (alkalinities that are less than 20mg/L).

2.1. A 50mL aliquot of sample is titrated with 0.02N H_2SO_4 to the pH 8.3 endpoint (if possible) and the pH 4.5 endpoint. Carbonates, bicarbonates, and hydroxides are calculated from the titration volumes to both the pH 8.3 and 4.5 endpoints.

2.1.1 The automated system (Poseidon) titrates to all pH endpoints and calculates all results.

2.1.1.1 All method parameters and calculations used by the Tiamo software are included in Appendix C.

2.2. For low level alkalinities 50mL of sample is titrated with 0.02N H_2SO_4 using a 10mL micro burette to pH 4.5 ±0.20 endpoint. Additional titrant is added to lower the pH an additional 0.3 pH units. Low level alkalinity is then calculated from the titration volumes to pH 4.5 and 0.3 pH unit's lower endpoints.

2.2.1 The automated system titrates each sample to a final endpoint of 4.2pH units.

2.2.2 The automated system calculates the low level alkalinity result.

3.0 Detection Limits

3.1. The PQL for total alkalinity and bicarbonate is 20 mg/L as $CaCO_3$. The PQL for carbonate and hydroxide is 2.0 mg/L as $CaCO_3$.

3.2. The PQL for low level alkalinity is 2.0 mg/L as $CaCO_3$.

3.3. MDLs shall be empirically established annually and are subject to change. The current MDL can be located in the LIMS. See HEAL QAP for more information on the appropriate determination of the MDL.

4.0 Definitions

4.1. Analytical batch – up to 20 samples and including a MB, LCS, MS, and MSD; i.e. 24 individual determinations.

4.2. Blank spike – A blank spike is a know amount of DI H_2O fortified with a known concentration of desired spike. Different than the LCS due to concentration(s).

4.3. DI H_2O (DI) – De-ionized water/reagent free water. Water that is passed through a purification system and is free from interferences.

4.4. Laboratory Control Spike (LCS) – Laboratory fortified blank that is spiked at a concentration of 80mg/L.

4.5. Matrix Spike (MS) – A sample fortified, at 80mg/L, in the laboratory.

4.6. Matrix Spike Duplicate (MSD) – A duplicate sample fortified, at 80mg/L, in the laboratory.

4.7. Method Blank (MB) – A blank analysis of DI H_2O analyzed prior to LCS and samples. The calculated value must be less than that of the necessary PQL for samples being determined. Used to show that the automated system is clean prior to sample analysis.

5.0 Interferences

5.1. Salts of weak organic acids and inorganic acids, present in large amounts, may cause interference in electrometric pH measurements.

5.2. Oil and grease present in samples may coat the electrode and hinder the electrometric response.

5.3. Temperature of the sample(s) being analyzed can cause interference with the pH electrode. Samples should be allowed to equilibrate to room temperature prior to analysis. The pH electrode used must correct for temperature differences.

6.0 Safety

6.1. For handling procedures and exposure information always refer to the MSDS. To keep exposure to reagents and standards to a minimum, analysts must wear appropriate laboratory safety attire when working in the laboratory.

6.2. Refer to the current HEAL Health and Safety Manual.

7.0 Apparatus and Materials

7.1. Poseidon – 855 Robotic Titrosampler equipped with: 1-50mL 807 Dosino dosing unit, 1-20mL 807 Dosino dosing unit with 2L bottle, 772 peristaltic pump unit, pH combination electrode, Conductivity combination electrode, and Tiamo software for operation and data collection.

7.1.1 50mL graduated cylinder.

7.1.2 50mL disposable digestion vessels from Environmental Express.

7.1.3 60mL disposable plastic cups.

7.1.4 DI H_2O.

7.1.5 Sulfuric Acid, Standardized, 0.02000N.

7.1.5.1 The concentration of the sulfuric acid may change.

7.1.5.2 The Technical Director must be notified of change so that software can be configured to calculate appropriately due to new normality of sulfuric acid.

7.1.6 Sulfuric Acid, Standardized, 0.2N.

7.1.7 Spike solution

7.1.7.1 Stock Alkalinity solution: Dissolve 0.848g of Na_2CO_3 (dried at 250°C for two hours and cooled in a desiccator) into 200mL DI water. This is a 4000 mg/L as $CaCO_3$ stock solution.

7.1.7.2 Spike 1mL into LCS, MS, and MSD samples. Report spike results as total alkalinity only. In LCS, MS, and MSD the spike concentration is 80 mg/L as $CaCO_3$.

7.1.8 pH Buffers 1.68, 4.00, 7.00, 10.00, 12.45 for calibration of pH meter.

7.1.8.1 See current pH SOP for further detailed discussion of pH calibration and requirements.

7.2. Memphis - pH meter. See current HEAL SOP for determination of pH for detailed calibration procedures and QC requirements.

7.2.1 50mL graduated cylinder.

7.2.2 Magnet stirrer and Teflon stir-bars.

7.2.3 Class A 50mL burette – Cartman.

7.2.4 Class A 10mL micro-burette – Kyle.

7.2.5 Class A 200mL volumetric flask

7.2.6 Disposable plastic 4oz Solo cups.

7.2.7 Glass 150mL beakers.

7.2.8 DI H_2O.

7.2.9 Sulfuric Acid, Standardized, 0.02000N.

7.2.10 Spike solution

7.2.10.1 Stock Alkalinity solution: Dissolve 0.848g of Na_2CO_3 (dried at 250°C for two hours and cooled in a desiccator) into 200mL DI water. This is a 4000 mg/L as $CaCO_3$ stock solution.

7.2.10.2 Spike 1mL into LCS, MS, and MSD samples. Report spike results as total alkalinity only. In LCS, MS, and MSD the spike concentration is 80 mg/L as $CaCO_3$.

8.0 Sample collection, Preservation, and Storage

8.1. Samples are to be collected in unpreserved plastic or glass containers. Samples are stored at above freezing to 6°C until analysis.

8.2. Holding time is 14 days.

8.3. Samples should be removed from the storage refrigerator at least 2 hours prior to analysis to allow samples to equilibrate to room temperature.

9.0 Quality Control

9.1. An analytical batch consists of a MB, LCS, MS, MSD, and up to 20 samples.

9.2. See the current HEAL pH SOP for calibration procedure for pH electrode and acceptance criteria.

9.3. A bracketing pH CCV must be analyzed at the end of each batch and in between every 20 new samples. Not including MB, LCS, DUP, and MS/MSD samples.

10.0 Procedure for 20mg/L and 2mg/L PQL alkalinities - automated

10.1. Poseidon – automated titration

10.1.1 Remove samples from refrigerator and set them out on the counter for at least two hours before analysis.

10.1.2 Follow procedure outlined in current HEAL pH SOP for instrument preparation and calibration of the pH electrode.

10.1.2.1 Make certain that the pH calibration is verified to be acceptable prior to continuing.

10.1.3 If more than 5 minutes has passed since the ICV buffer has been analyzed, i.e. the instrument has been idle due to review of pH calibration and pH ICV results, prepare, load, and analyze a pH buffer according to the current HEAL pH SOP.

10.1.3.1 The acceptance criteria for this pH buffer is listed in the current HEAL pH SOP.

10.1.4 Obtain clean 60mL auto-sampler cups.

10.1.5 Login to the *Tiamo* software using the secure login.

10.1.6 Select the Workplace icon. Click on the *Determination Series* tab.

10.1.6.1 Double click on the first line and choose the *Cond.-pH-alk 50mL* method from the scroll down menu.

10.1.6.2 Enter the HEAL sample ID on ID1 line.

10.1.6.3 Enter additional notes for sample on ID2 line.

10.1.6.4 Enter the sample position number.

10.1.6.5 Click the right scroll arrow on the bottom left of the window to advance to the next sample position.

10.1.6.5.1 Repeat 10.1.5.2 to load additional samples into list.

10.1.6.6 Click *Apply*, and then *Close* when finished entering IDs for all desired analyses.

10.1.7 After sample list has been filled appropriately, and prior to analysis, the list should be printed. The list shall be used to ensure that samples are located in appropriate positions on the auto-sampler rack.

10.1.7.1 Click on *Sample table* and select *Print (PDF)*. This will generate a temporary PDF file.

10.1.7.2 Print the PDF file. Save the PDF file to the Sample list directory located on the desktop with the file name corresponding to the analysis date.

10.1.8 Once sample list is complete and has been printed, prepare and load samples according to the sample list. Using this sequence should reduce analyst error of loading the auto-sampler incorrectly.

10.1.9 Measure 50mL of sample in a graduated cylinder. Pour this into a 60mL disposable plastic beaker. Place samples in the auto-sampler tray in the appropriate sample positions.

10.1.9.1 Always load a MB and LCS prior to sample analysis.

10.1.9.1.1 Use 50mL DI H2O for the MB.

10.1.9.1.2 Follow Section 7.1.7.2 for the spiking procedure of the LCS.

10.1.9.2 Avoid floating particulates as they can cause interferences with the electrode and causes blockages in the instrumentation plumbing.

10.1.10 Remember to load and analyze a MS and MSD for up to every twenty samples.

10.1.11 Click *Start* to begin analysis.

10.1.12 The auto-sampler rinses between each sample.

10.1.13 The aspiration line and conductivity probe on the robotic arm are rinsed three times between sample analyses.

10.1.14 Samples may require a "dilution" to prevent over flow from the titration beaker.

10.1.14.1 Use 20mL of sample instead of the routine 50mL of sample.

10.1.14.2 If over flow is still a concern, discuss with Technical Director the changing of the 0.02N H_2SO_4 to a higher normality, 0.2N H_2SO_4.

10.1.14.3 When changing to a different normality, the system must be prepped again to make certain lines are flushed. The analysis of an additional LCS is also required.

11.0 Procedure for 20mg/L and 2mg/L PQL alkalinities - manual

11.1. Memphis – manual titration with 50mL and 10mL burettes

11.1.1 Calibrate the pH meter according to current HEAL pH SOP.

11.1.1.1 All acceptance criteria listed in the current HEAL pH SOP must be meet.

11.1.2 20mg/L PQL alkalinity

11.1.2.1 Pour out old 0.02N H_2SO_4 titrant from 50mL burette and refill with fresh solution. Remove bubbles from burette tip and record volume in logbook.

11.1.2.2 For MB measure 50mL DI H_2O using graduated cylinder.

11.1.2.2.1 Add magnetic stir bar and gently stir sample.

11.1.2.2.2 Immerse pH probe in solution. Be sure the bulb (and thermocouple if present) is fully immersed in solution. Do not allow the stir bar to hit probe tip.

11.1.2.2.3 Allow pH to stabilize, record starting pH, burette volume, and titrate to pH 4.5 ±0.20 pH units. (For MB proceed very slowly, this will be a very small volume).

11.1.2.2.4 Record ending pH and burette volume.

11.1.2.3 For LCS measure 50mL DI H_2O using a graduated cylinder and pour into a clean plastic Solo cup.

11.1.2.3.1 Using the 1mL pipette, remove 1mL of DI H_2O from Solo cup, get a new tip for the pipette, then spike 1mL of Stock Alkalinity solution (this will be the LCS).

11.1.2.3.2 Add magnetic stir bar and gently stir sample.

11.1.2.3.3 Immerse pH probe in solution. Be sure the bulb (and thermocouple if present) is fully immersed in solution. Do not allow the stir bar to hit probe tip.

11.1.2.3.4 Allow pH to stabilize, record starting pH, burette volume, and titrate to pH 4.5 ±0.20 pH units.

11.1.2.3.5 Record ending pH and burette volume.

11.1.2.4 For samples, measure 50mL of un-mixed sample in graduated cylinder, and pour into plastic 4oz Solo cup.

11.1.2.4.1 Samples with very high alkalinities may be diluted prior to titration. DO NOT dilute the sample in DI H_2O. Simply titrate less sample. Be sure there is enough sample to allow pH probe tip to be completely submerged. 20mL of sample is appropriate.

11.1.2.4.2 If a sample with a very high alkalinity has not reached the pH 4.5 endpoint but the Solo cup is full, transfer Stir bar and sample to a clean 150mL beaker and continue titration to the endpoint.

11.1.2.4.3 Add stir bar, begin mixing, and immerse probe.

11.1.2.4.4 Allow pH to stabilize. Record pH.

11.1.2.4.5 If pH is above 8.30, titrate to pH 8.30 ±0.05 pH units. Record pH and burette volume.

11.1.2.4.6 Continue titration of sample until pH 4.5 end-point is reached (±0.20 pH units). Record pH and burette volume.

11.1.2.4.7 Dump sample down the drain, dispose of plastic solo cup and rinse Teflon stir-bar with DI. Continue with the next sample.

11.1.2.5 For MS and MSD samples, measure 50mL of an un-mixed sample into a 50mL graduated cylinder, and pour into 4oz solo cup.

11.1.2.5.1 Using the 1mL pipette, remove 1mL of MS sample from Solo cup, get a new pipette tip, and then spike 1mL of Stock Alkalinity solution. Begin titration immediately.

11.1.2.5.2 Follow procedure above for the titration of a LCS. Repeat with MSD sample.

11.1.3 2.0mg/L PQL (low level alkalinity)

11.1.3.1 Low level alkalinity procedure is only used when requested by client. If low level alkalinity is requested the MB and any sample with a total alkalinity <20mg/L must be titrated using low level alkalinity procedure.

11.1.3.2 Pour out old 0.02N H_2SO_4 titrant from 10mL micro burette and refill with fresh solution. Remove bubbles from burette tip and record volume in logbook.

11.1.3.3 Measure 50mL DI H_2O in graduated cylinder and pour into a Solo cup (this will be the MB).

11.1.3.4 Add magnetic stir bar and gently stir sample.

11.1.3.5 Immerse pH probe in solution. Be sure the bulb (and thermocouple if present) is fully immersed in solution. Do not allow the stir bar to hit probe tip.

11.1.3.6 Allow pH to stabilize, record starting pH, and titrate to pH 4.5 ±0.20 pH units.

11.1.3.7 Allow pH to stabilize and record ending pH and burette volume. (Be very careful, this is a very small volume).

11.1.3.8 Continue titration to reduce the pH exactly 0.30 pH units. Record final pH and burette volume.

11.1.3.9 Repeat procedure using 50 mL of sample for any client samples that have <20 mg/L CaCO3 (require less than 1 mL of titrant to reach endpoint pH 4.5)

12.0 Calculations

12.1. Total Alkalinity

$$Total\ Alkalinity, mg/L\ CaCO_3 = \frac{T \times N \times 50{,}000}{mL\ of\ sample}$$

Where:

T=Total mL of standard acid used for titration to pH 4.5 endpoint
N=Normality of standard acid

12.2. 2mg/L PQL - Low Level Alkalinity

$$Total\ Alkalinity, mg/L\ CaCO_3 = \frac{((2B) - C) \times N \times 50{,}000}{mL\ of\ sample}$$

Where:

B = mL titrant to first recorded pH (± 0.2 pH units from pH 4.5)
C = total mL titrant to reach 0.30 pH units lower (mL to approx. pH 4.5 plus 0.30 ph units lower)

12.3. Calculation of Alkalinity Relationships

12.3.1 Alkalinity relationships are calculated with the assumption that the total alkalinity only consists of the sum of bicarbonate, carbonate, and hydroxide alkalinities. Other weak acids that may contribute to the total alkalinity are not included in the calculation of the alkalinity relationships.

12.3.2 These relationships are determined by examining the relationship between the volume of titrant used to reach the pH 8.3 endpoint (also known as the phenolphthalein endpoint or P) and the <u>total</u> volume of titrant (including the volume used to reach pH 8.3 endpoint) used to reach the pH 4.5 endpoint (also known as T).
P = mL titrant to reach pH 8.3.
T = total volume of titrant used to reach pH 4.5.

12.3.3 In this SOP the alkalinity relationships shall be called: Bicarbonate/Carbonate, Carbonate, and Carbonate/Hydroxide.

12.3.4 The sum of bicarbonate, carbonate, and hydroxide alkalinities must always equal that of the total alkalinity calculated in 12.1.

12.3.5 Calculation of Bicarbonate/Carbonate Alkalinity

Where P = 0 or P < 1/2 T.

Hydroxide: $OH^{-}\left(mg\ CaCO_3 / L\right) = 0$

Carbonate: $CO_3^{2-}\left(mg\ CaCO_3 / L\right) = \dfrac{\left(2 \times P \times 50{,}000 \times N\right)}{V}$

Bicarbonate: $HCO_3^{-}\left(mg / L\ CaCO_3 / L\right) = \dfrac{\left(T - 2 \times P\right) \times 50{,}000 \times N}{V}$

Where:

P = mL of titrant to pH 8.3
T = Total mL of titrant to pH 4.5
N = normality of titrant
V = volume of sample in mL

12.3.6 Calculation of Carbonate Alkalinity

Where P = ½ T.

Hydroxide: $OH^{-}(mg\ CaCO_3 / L) = 0$

Carbonate: $CO_3^{2-}(mg\ CaCO_3 / L) = \frac{2 \times P \times 50{,}000 \times N}{V}$

Bicarbonate: $HCO_3^{-}(mg\ CaCO_3 / L) = 0$

Where:
P = mL of titrant to pH 8.3
N = normality of titrant
V = volume of sample in mL

12.3.7 Calculation of Carbonate/Hydroxide Alkalinity

Where P > ½ T or P = T

Hydroxide: $OH^{-}(mg\ CaCO_3 / L) = \frac{(2 \times P - T) \times 50{,}000 \times N}{V}$

Carbonate: $CO_3^{2-}(mg\ CaCO_3 / L) = \frac{2(T - P) \times 50{,}000 \times N}{V}$

Bicarbonate: $HCO_3^{-}(mg\ CaCO_3 / L) = 0$

Where:
P = mL of titrant to pH 8.3
T = Total mL of titrant to pH 4.5
N = normality of titrant
V = volume of sample in mL

13.0 Method Performance

13.1. PT or "blind study" samples are routinely analyzed to check the accuracy of the total alkalinity analysis.

13.2. MDL studies are done at a minimum of yearly. MDLs are completed for both the manual and automated procedures, and must be prepared and analyzed over a minimum of three days. Additional procedural requirements and the associated formulas can be found in the QAP.

13.3. Control Charting of LCS recoveries shall be done for all LCS analyses.

14.0 Estimation of Uncertainty

14.1. Estimation of uncertainty is to be done on an annual basis.

14.1.1 Collect, at least, 20 of the most recent LCS data points. Calculate the standard deviation (s) and the average of all the readings. Calculate the 95% confidence interval (2 times the standard deviation). Report the estimated uncertainty as ±2(s). Uncertainty data is kept on file in the method folders in the quality control office. See current revision of QAP for the formulas required to calculate uncertainty.

15.0 Demonstration of Capability (DOC)

15.1. Demonstration of Capability (DOC) samples are analyzed before an analyst is able to independently generate data.

15.2. DOCs for total alkalinity consist of four replicate spikes.

15.3. A detailed description of DOCs can be found in the HEAL QA Plan.

15.4. See the current HEAL QAP for necessary calculation and statistical requirements for DOC studies.

15.5. Annual Documentation of Continued Proficiency (ADOCP) is performed annually after acceptable DOC has been established. See HEAL Quality Assurance Plan.

16.0 Pollution Prevention

16.1. Standard 0.02N H_2SO_4 titrant comes pre-made and standardized. No preparation or standardization is necessary.

16.2. The automated titration system uses more pH buffer volume. However, using this system currently allows for at least three determinations to be done on a single aliquot of sample.

17.0 Data Assessment and Acceptance Criteria for Quality Control Measures

17.1. See **Appendix A** for a summary of batch QC requirements.

17.2. Total alkalinity value can be found on the generated report. The software calculates all necessary values. These values are based on a 50mL sample size. If a different sample volume was used, the value must be corrected.

17.3. pH Endpoints

17.3.1 The acceptable range for pH 8.3 endpoint is ±0.05 pH units.

17.3.2 The acceptable range for pH 4.5 endpoint is ±0.20 pH units

17.3.3 The acceptable range for additional 0.30 pH units for low level alkalinities is ±0.05 pH units.

17.4. Batch QC

17.4.1 The MB must calculate to ≤20mg/L when reporting to the 20mg/L PQL.

17.4.1.1 Use the *Total Alk* value from the report.

17.4.2 The MB must calculate to ≤2mg/L when reporting to the 2.0mg/L PQL.

17.4.2.1 Use the *Low Level Titration* value from the report.

17.4.3 The LCS must be recovered to within ±20% of the assigned value, until control chart limits are established.

17.4.4 The MS and MSD must be recovered to within ±20% of the assigned value.

17.4.5 The RPD for the MSD must be ≤20%.

17.5. Continuing Calibration Verification

17.5.1 pH CCV must pass within ±0.1 pH units.

18.0 Corrective Actions for Out-of-Control Data

18.1. pH Endpoint

18.1.1 If sample is titrated to a pH greater than the required endpoint, add additional titrant to bring pH into the acceptable range. If the sample has been titrated to less than the required endpoint (pH overshot), the sample must be discarded and re-titrated.

18.1.2 If the MB calculates >2 mg $CaCO_3$, reanalyze using fresh DI from a different location (i.e. any DI pump dispenser or wash bottle containing DI) when the 2.0mg/L PQL is required. This may be an indication that the DI water system may not be producing ultra-pure water. Notify the Inorganic manager immediately. If MB continues to fail discontinue analysis until to source of the problem is fixed.

18.2. Batch QC

18.2.1 If the LCS fails, re-spike and reanalyze. If the spike continues to fail, remake stock spiking solution, spike and reanalyze. If it continues to fail discontinue analysis until the source of the problem can be fixed. The LCS value must be blank corrected.

18.2.2 If MS or MSD fails, re-spike and reanalyze. If spike continues to fail, flag data and report.

18.2.3 If RPD for MSD fails. Re-spike and reanalyze both MS and MSD, report reanalysis. If RPD is still out flag data and report.

18.3. Continuing Calibration Verification

18.3.1. If pH CCV fails, all alkalinity samples to the surrounding the failing CCV must be reanalyzed to the next acceptable CCV.

19.0 Contingencies for Handling Out of Control Data

19.1. All out of control data is flagged and reported to the client.

19.2. Corrective actions shall be filled out when there are deviations from the method or this SOP.

20.0 Waste Management

20.1. Analytical results are used to characterize their respective sample contamination level(s) so that the proper disposal can be performed. These wastes will be disposed of according to their hazard as well as their type and level of contamination. Refer to the Hall Environmental Analysis Laboratory Chemical Hygiene Plan for details regarding waste disposal.

21.0 References

21.1. Standard Methods 20th Edition, Method 2320B.

21.2. Quality Assurance Plan, HEAL.

21.3. Chemical Hygiene Plan, HEAL.

21.4. Health and Safety SOP, HEAL.

21.5. Tiamo Software, Online Help, Metrohm.

Appendix A

Standard Methods 2320B: Total Alkalinity, Carbonate, Bicarbonate, and Hydroxide in Aqueous Samples
Corrective Action Procedures

Analytical Method	Parameter	QC Element	Frequency	Acceptance Criteria	Corrective Action
SM2320B	Wet Chemistry	pH Endpoint (8.30)	every sample, including QC	±0.05 pH units	1. If pH is "overshot" sample must be re-titrated
		pH Endpoint (4.50)	every sample, including QC	±0.20 pH units	1. If pH is "overshot" sample must be re-titrated
		Method Blank	1 per preparation/analysis batch (≤ 20 samples)	≤ PQL	1. Reanalyze using fresh DI from a different source. 2. Discontinue analysis.
		Laboratory Control Sample (LCS)	1 per preparation/analysis batch (≤ 20 samples)	% Recovery +/- 20% of actual value	1. Re-spike and reanalyze. 2. Remake spike solution. Respike and reanalyze. 3. Discontinue analysis.
		Matrix Spike (MS)	1 per preparation/analysis batch (≤ 20 samples)	% Recovery +/- 20% of actual value	1. Re-spike and reanalyze. 2. Flag data and report.
		Matrix Spike Duplicate (MSD)	1 per preparation/analysis batch (≤ 20 samples)	% Recovery +/- 20% of actual value Relative percent difference (RPD) ≤20%	1. MSD recovery: Re-spike and reanalyze. 2. MSD recovery: Flag data and report. 3. RPD: Re-spike and reanalyze both MS and MSD, report reanalysis. 4. RPD: Flag data and report.
		Continuing Calibration Verification (CCV)	1 per every 20 new samples	±0.1 pH units of actual value	1. All previous 20 new samples, and following 20 new samples must be reanalyzed to the next passing CCV

Appendix B.
Comparison Data. Automated titration using Poseidon versus Manual titration.

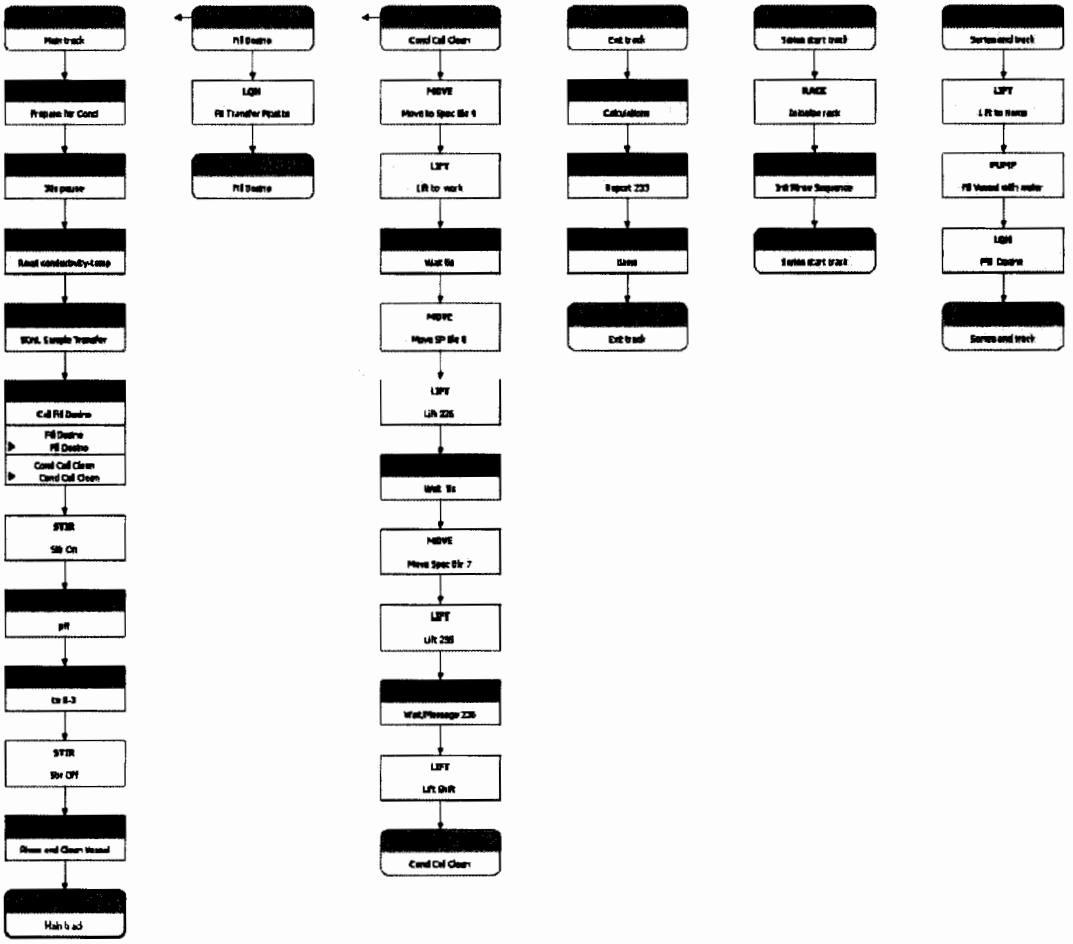
		Poseidon		Burette		%RPD
Sample 1		64.17		63		1.84%
Sample 2		424.12		433		2.07%
Sample 3		419.32		433		3.21%
Sample 4		346.12		374		7.74%
Sample 5		536.48		554		3.21%
Sample 6		521.73		542		3.81%
Sample 7		132.02		136		2.97%
Sample 8		148.47		153		3.01%
Sample 9		29.63		32		7.69%
Sample 10		14.82		15		1.21%
Sample 11		119.83		128		6.59%
Sample 12		120.41		127		5.33%
Sample 13		699.30		742		5.93%
Sample 14		246.61		258		4.51%
Sample 15		170.92		180		5.17%
Sample 16		10.12		12		17.00%
Sample 17		64.20		56		13.64%
Sample 18		91.00		95		4.30%
Sample 19		185.53		190		2.38%
Sample 20		493.20		544		9.80%

Appendix C.

Software Method Flow Chart
Software Method Parameters
Software Method Calculations

Method run

Method name: Cond.-pH-Alk- 50 mL updated
Method version: 5
Method group: Main group



Method parameters

Method ..... Cond.-pH-Alk- 50 mL updated
Method saving date 2009-02-03 14:48:03 UTC-7
Method version 5
Method group ..... Main group
Method status ..... original
Method saved by (full name) ..... Benjamin Derek Harmon
Method saved by (short name) ..... bdh

START **Main track**

General

Workplace view

Current view ..... on

Track view for live window

Live display 1 ..... Main track

Live display 2 ..... Main track

Statistics ..... off

Conditioning

Automatic conditioning ..... off

Application note

Method variables

Name	Type	Assignment	Value	Comment	Monitoring
Sample size	Number	Sample size		Sample size	off
Sample size unit	Text	Sample size unit		Sample size unit	off
Sample position	Number	Sample position		Sample position number	off
ID1	Text	ID1		Sample identification 1	off
ID2	Text	ID2		Sample identification 2	off
ID3	Text	ID3		Sample identification 3	off
cond	Text				off
temp	Text				off

Name ..... **Sample size**
Type ..... Number
Assignment ..... on ..... Sample size
Value ..... off .....
Check at start ..... off
Comment ..... Sample size
Variable monitoring ..... off
Lower limit .....
Upper limit .....
Message .....
Display message ..... on
Record message ..... on

Message by e-mail ..... off
Mail to .....
Subject ..... Message from tiamo - Method 'New method 2' - Command 'Main track'
User .....
Mail from .....
SMTP Server .....
POP3 Server .....
Acoustic signal ..... off
Action ..... off
Stop determination ..... on
Stop determination and series ..... off

Name ..... **Sample position**
Type ..... Number
Assignment ..... on ..... Sample position
Value ..... off .....
Check at start ..... on
Comment ..... Sample position number
Variable monitoring ..... off
Lower limit .....
Upper limit .....
Message .....
Display message ..... on
Record message ..... on
Message by e-mail ..... off
Mail to .....
Subject ..... Message from tiamo - Method 'New method 2' - Command 'Main track'
User .....
Mail from .....
SMTP Server .....
POP3 Server .....
Acoustic signal ..... off
Action ..... off
Stop determination ..... on
Stop determination and series ..... off

Name ..... **Sample size unit**
Type ..... Text
Assignment ..... on ..... Sample size unit
Value ..... off .....
Check at start ..... off
Comment ..... Sample size unit

Name **ID1**
Type ..... Text

Assignment ..... on ID1
Value ..... off .....
Check at start ..... on
Comment ..... Sample identification 1

Name **ID2**
Type ..... Text
Assignment ..... on ID2
Value ..... off .....
Check at start ..... off
Comment ..... Sample identification 2

Name **ID3**
Type ..... Text
Assignment ..... on ID3
Value ..... off .....
Check at start ..... off
Comment ..... Sample identification 3

Name ..... **cond**
Type ..... Text
Assignment ..... off .....
Value ..... on .....
Check at start ..... off
Comment .....

Name ..... **temp**
Type ..... Text
Assignment ..... off .....
Value ..... on .....
Check at start ..... off
Comment .....

SEQUENCE **Prepare for Cond**

MOVE **Move to Sample Pos**

Device
- Device name ..... 855_1

Target
- Tower 1
- Move ..... Sample position

Beaker test
- Display message ..... on
- Stop determination ..... off
- Stop determination and series ..... off

Parameters
- Shift rate ..... 20 °/s
- Shift direction ..... auto

Swing rate . . . . . 55 °/s

LIFT **Lift Down**

Device

Device name . . . . . 855_1

Target

Tower 1

Lift position . . . . . Work position mm

Parameters

Lift rate . . . . . 25 mm/s

LIFT **Lift Up**

Device

Device name . . . . . 855_1

Target

Tower 1

Lift position . . . . . Rinse position mm

Parameters

Lift rate . . . . . 25 mm/s

LIFT **Lift Work**

Device

Device name . . . . . 855_1

Target

Tower 1

Lift position . . . . . Work position mm

Parameters

Lift rate . . . . . 25 mm/s

WAIT **30s pause**

Wait

Stop track and waiting for [Continue] . . . . . off

Stop all tracks and waiting for [Continue] . . . . . off

Waiting time . . . . . on

Time 30

Unit . . . . . s

Message

Record message . . . . . off

Message by e-mail . . . . . off

Acoustic signal . . . . . off

TRANSFER **Read conductivity-temp**

Device

Device name 712

Transfer commands

Name	Action	Command	Variable	Comparison
conductivity	Read	&i.a.m.c$q	MV.cond	(\d+\.\d+E.\d+)

Name	Action	Command	Variable	Comparison
temp	Read	&i.a.m.t$q	MV.temp	(\d+\.\d+)

Timeout ..... off

SEQUENCE **50mL Sample Transfer**

LQH **Fill Dosino with Sample**

General/Hardware

Device

Device name ..... 855_1

Dosing device

Dosing device 1

Solution ..... Transfer Pipette

Parameters

Function ..... Aspirate

Port 1

Volume ..... 50.000 mL

Rate ..... 100.0 mL/min

WAIT **Wait 3**

Wait

Stop track and waiting for [Continue] ..... off

Stop all tracks and waiting for [Continue] ..... off

Waiting time ..... on

Time 3

Unit ..... s

Message

Record message ..... off

Message by e-mail ..... off

Acoustic signal ..... off

LIFT **Lift to Shift**

Device

Device name ..... 855_1

Target

Tower 1

Lift position ..... Shift position mm

Parameters

Lift rate ..... 25 mm/s

LQH **Dispense Sample**

General/Hardware

Device

Device name ..... 855_1

Dosing device

Dosing device 1

Solution ..... Transfer Pipette

Parameters

Function . . . . . Eject to end volume
Port 3
Rate . . . . . maximum mL/min

LQH **Get remaining sample**

General/Hardware

Device
Device name . . . . . 855_1
Dosing device
Dosing device 1
Solution . . . . . Transfer Pipette

Parameters

Function . . . . . Aspirate
Port 1
Volume . . . . . 15.00 mL
Rate . . . . . 100.0 mL/min

LQH **Dispense remaining sample**

General/Hardware

Device
Device name . . . . . 855_1
Dosing device
Dosing device 1
Solution . . . . . Transfer Pipette

Parameters

Function . . . . . Eject to end volume
Port 3
Rate . . . . . maximum mL/min

CALL **Call Fill Dosino**

Call text	Track name	Condition	Condition
Fill Dosino	Fill Dosino	off	
Cond Cell Clean	Cond Cell Clean	off	

STIR **Stir On**

Device
Device name . . . . . 855_1
Stirrer
Stirrer 3
Stirrer type . . . . . 801 Magnetic stirrer
Stirring rate 15
Action
Switch on . . . . . on
Switch off . . . . . off
Duration . . . . . off

MEAS pH **pH**

General/Hardware

Device

Device name ..... 855_1

Sensor

Measuring input 1

Sensor ..... pH electrode

Temperature measurement ..... automatic

Stirrer

Stirrer ..... off

Measuring parameters

Measurement

Measurement with drift control ..... on

Signal drift ..... 2.0 mV/min

Min. waiting time ..... 20 s

Max. waiting time ..... 60 s

Measuring interval ..... 1 s

Stop measured value pH ..... off

Measurement without drift control ..... off

Temperature

Temperature ..... 25.0 °C

Evaluations

Fix end point evaluation ..... off

Minimum evaluation ..... off

Maximum evaluation ..... off

Break point evaluation ..... off

Additional measured values

Additional calculated measured values ..... off

Additional external measured values ..... off

SET pH **to 8-3**

General/Hardware

Device

Device name ..... 855_1

Dosing device

Dosing device 2

Solution H2SO4

Sensor

Measuring input 1

Sensor ..... pH electrode

Temperature measurement ..... automatic

Stirrer

Stirrer ..... off

Start conditions

Initial measured value

Signal drift ..... off mV/min

Min. waiting time . . . . . 0 s
Max. waiting time . . . . . 1 s
Pause 1
Pause 1 . . . . . 0 s
Start volume
Start volume . . . . . 0 mL
Dosing rate . . . . . maximum mL/min
Pause 2
Pause 2 . . . . . 0 s

Control parameters

End point 1
EP1 at pH 8.3
Titration rate
Titration rate . . . . . user
Control
Dynamics pH 4.000
Max. rate . . . . . 8.0 mL/min
Min. rate . . . . . 5.0 µL/min
Stop criterion
Stop criterion . . . . . drift
Stop drift . . . . . 20 µL/min
End point 2 . . . . . on
EP2 at pH 4.2
Titration rate
Titration rate . . . . . user
Control
Dynamics pH 4.0
Max. rate . . . . . 10.00 mL/min
Min. rate . . . . . 25.0 µL/min
Stop criterion
Stop criterion . . . . . drift
Stop drift . . . . . 20 µL/min

Titration parameters

Titration direction . . . . . -
Extraction time . . . . . 0 s
Temperature . . . . . 25.0 °C
Time interval measuring point . . . . . 1 s

Stop conditions

Stop volume . . . . . 60.00 mL
Stop time . . . . . off s
Filling rate . . . . . maximum mL/min

Conditioning

Conditioning . . . . . off

Additional evaluations

Fix end point evaluation . . . . . on
Fixed quantity . . . . . Measured value

Fix EP1 at pH 4.5
Fix EP2 at pH ..... off
Fix EP3 at pH ..... off
Minimum evaluation ..... off
Maximum evaluation ..... off
Additional measured values
Additional calculated measured values ..... off
Additional external measured values ..... off

STIR **Stir Off**
Device
Device name ..... 855_1
Stirrer
Stirrer 3
Stirrer type ..... 801 Magnetic stirrer
Stirring rate 8
Action
Switch on ..... off
Switch off ..... on
Duration ..... off

SEQUENCE **Rinse and Clean Vessel**

PUMP **Asp Pump On**
Device
Device name ..... 855_1
Pumps
Tower 1
Pump(s) 2
Action
Switch on ..... on
Switch off ..... off
Duration ..... off

WAIT **Wait 12s**
Wait
Stop track and waiting for [Continue] ..... off
Stop all tracks and waiting for [Continue] ..... off
Waiting time ..... on
Time 12
Unit ..... s
Message
Record message ..... off
Message by e-mail ..... off
Acoustic signal ..... off

MOVE **Move Spec 5**
Device
Device name ..... 855_1

Target
- Tower 1
- Move . . . . . Special beaker
- Number 5

Beaker test
- Display message . . . . . on
- Stop determination . . . . . off
- Stop determination and series . . . . . off

Parameters
- Shift rate . . . . . 20 °/s
- Shift direction . . . . . auto
- Swing rate . . . . . 55 °/s

LQH **Clean Port 1 Tubing**

General/Hardware

Device
- Device name . . . . . 855_1

Dosing device
- Dosing device 1
- Solution . . . . . Transfer Pipette

Parameters
- Function . . . . . Dosing
- Port 1
- Volume . . . . . 3.25 mL
- Rate . . . . . maximum mL/min

LQH **Clean port 3 tubing**

General/Hardware

Device
- Device name . . . . . 855_1

Dosing device
- Dosing device 1
- Solution . . . . . Transfer Pipette

Parameters
- Function . . . . . Eject to end volume
- Port 3
- Rate . . . . . maximum mL/min

LIFT **Shift Pos**

Device
- Device name . . . . . 855_1

Target
- Tower 1
- Lift position . . . . . Shift position mm

Parameters
- Lift rate . . . . . 25 mm/s

LQH **Fill with air**

General/Hardware

Device

Device name ..... 855_1

Dosing device

Dosing device 1

Solution ..... Transfer Pipette

Parameters

Function ..... Aspirate

Port 1

Volume ..... 15.00 mL

Rate ..... maximum mL/min

LQH **Dispense Air**

General/Hardware

Device

Device name ..... 855_1

Dosing device

Dosing device 1

Solution ..... Transfer Pipette

Parameters

Function ..... Eject to end volume

Port 3

Rate ..... maximum mL/min

PUMP **Rinse Pump-On**

Device

Device name ..... 855_1

Pumps

Tower 1

Pump(s) 1

Action

Switch on ..... off

Switch off ..... off

Duration ..... on

Time ..... 4.0 s

WAIT **Wait 7s**

Wait

Stop track and waiting for [Continue] ..... off

Stop all tracks and waiting for [Continue] ..... off

Waiting time ..... on

Time 7

Unit ..... s

Message

Record message ..... off

Message by e-mail ..... off

Acoustic signal ..... off

PUMP **Pump on**

Device
- Device name ..... 855_1

Pumps
- Tower 1
- Pump(s) 1

Action
- Switch on ..... off
- Switch off ..... off
- Duration ..... on
- Time ..... 4.0 s

WAIT **Wait 10s**

Wait
- Stop track and waiting for [Continue] ..... off
- Stop all tracks and waiting for [Continue] ..... off
- Waiting time ..... on
  - Time 10
  - Unit ..... s

Message
- Record message ..... off
- Message by e-mail ..... off
- Acoustic signal ..... off

PUMP **Pump Off**

Device
- Device name ..... 855_1

Pumps
- Tower 1
- Pump(s) 2

Action
- Switch on ..... off
- Switch off ..... on
- Duration ..... off

TRACK **Fill Dosino**
- Return immediately ..... on
- Delete all data ..... off

LQH **Fill Transfer Pipette**

General/Hardware

Device
- Device name ..... 855_1

Dosing device
- Dosing device 1
- Solution ..... Transfer Pipette

Parameters
- Function ..... Fill

Port 2
Rate . . . . . maximum mL/min

TRACK **Cond Cell Clean**
Return immediately . . . . . on
Delete all data . . . . . off

MOVE **Move to Spec Bkr 9**
Device
Device name . . . . . 855_1
Target
Tower 1
Move . . . . . Special beaker
Number 9
Beaker test
Display message . . . . . on
Stop determination . . . . . off
Stop determination and series . . . . . off
Parameters
Shift rate . . . . . 20 °/s
Shift direction . . . . . auto
Swing rate . . . . . 55 °/s

LIFT **Lift to work**
Device
Device name . . . . . 855_1
Target
Tower 1
Lift position . . . . . 218 mm
Parameters
Lift rate . . . . . 25 mm/s

WAIT **Wait 5s**
Wait
Stop track and waiting for [Continue] . . . . . off
Stop all tracks and waiting for [Continue] . . . . . off
Waiting time . . . . . on
Time 5
Unit . . . . . s
Message
Record message . . . . . off
Message by e-mail . . . . . off
Acoustic signal . . . . . off

MOVE **Move SP Bkr 8**
Device
Device name . . . . . 855_1
Target
Tower 1

Move . . . . . Special beaker
Number 8
Beaker test
Display message . . . . . on
Stop determination . . . . . off
Stop determination and series . . . . . off
Parameters
Shift rate . . . . . 20 °/s
Shift direction . . . . . auto
Swing rate . . . . . 55 °/s

LIFT **Lift 226**
Device
Device name . . . . . 855_1
Target
Tower 1
Lift position . . . . . 218 mm
Parameters
Lift rate . . . . . 25 mm/s

WAIT **Wait 5s**
Wait
Stop track and waiting for [Continue] . . . . . off
Stop all tracks and waiting for [Continue] . . . . . off
Waiting time . . . . . on
Time 5
Unit . . . . . s
Message
Record message . . . . . off
Message by e-mail . . . . . off
Acoustic signal . . . . . off

MOVE **Move Spec Bkr 7**
Device
Device name . . . . . 855_1
Target
Tower 1
Move . . . . . Special beaker
Number 7
Beaker test
Display message . . . . . on
Stop determination . . . . . off
Stop determination and series . . . . . off
Parameters
Shift rate . . . . . 20 °/s
Shift direction . . . . . auto
Swing rate . . . . . 55 °/s

LIFT **Lift 235**

Device
- Device name . . . . . 855_1

Target
- Tower 1
- Lift position . . . . . 218 mm

Parameters
- Lift rate . . . . . 25 mm/s

WAIT **Wait/Message 236**

Wait
- Stop track and waiting for [Continue] . . . . . off
- Stop all tracks and waiting for [Continue] . . . . . off
- Waiting time . . . . . on
  - Time 5
  - Unit . . . . . s

Message
- Record message . . . . . off
- Message by e-mail . . . . . off
- Acoustic signal . . . . . off

LIFT **Lift Shift**

Device
- Device name . . . . . 855_1

Target
- Tower 1
- Lift position . . . . . Shift position mm

Parameters
- Lift rate . . . . . 25 mm/s

EXIT **Exit track**

CALC **Calculations**

Result name	Formula	Unit	Decimal places	Assignment	Statistics
Cond	=1000000 * TextToNumber('MV.cond')	µS/cm	2	RS01	off
Cond Temp	=TextToNumber('MV.temp')	°C	1	RS02	off
pH	= 'pH.EME'	pH units	2	RS03	off
pH Temp	= 'pH.ETE'	°C	1	RS04	off
mLs to 8-3	= Case('to 8-3.EP{1}.VOL' ; 'to 8-3.EP{1}.VOL' ;0;0)	mL	2	RS05	off
mLs to 4-5	= 'to 8-3.FP{1}.VOL'	mL	2	RS06	off
mLs to 4-2	= 'to 8-3.EP{2}.VOL'	mL	2	RS11	off
Total Alk	= ('to 8-3.FP{1}.VOL' * 0.02 * 50000) / 50	mg/L	2	RS07	off
OH raw	= (((2 * Case('to 8-3.EP{1}.VOL' ; 'to 8-3.EP{1}.VOL' ;0;0)) - 'to 8-3.FP{1}.VOL') * 50000 * .02) / 50	mg/L	2	none	off
OH	=Case('RS.OH raw'< 0;0;'RS.OH raw')	mg/L	2	RS08	off

Result name	Formula	Unit	Decimal places	Assignment	Statistics
CO3	= (2 * Case('to 8-3.EP{1}.VOL' : 'to 8-3.EP {1}.VOL' ;0;0) * 50000 * 0.02) / 50	mg/L	2	RS09	off
HCO3 raw	= (('to 8-3.FP{1}.VOL' - (2 * Case('to 8-3. EP{1}.VOL' : 'to 8-3.EP{1}.VOL' ;0;0))) * 50000 * 0.02) / 50	mg/L	2	none	off
HCO3	=Case('RS.HCO3 raw'<0;0;'RS.HCO3 raw')	mg/L	2	RS10	off
Low Level Titration	= (((2 * 'to 8-3.FP{1}.VOL') - 'to 8-3. EP{2}.VOL') * 0.02 * 50000) / 50	mg/L	2	RS12	off
est TDS	=0.66 * 'RS.Cond'	mg/L	0	RS13	off
Free CO2/L	= 2.0 * 'RS.HCO3' * (10 ^ (6- 'RS.pH'))	mg/L	2	RS19	off
Total CO2/L	= 'RS.Free CO2/L' + (.44 * ((2 * 'RS. HCO3') + 'RS.CO3'))	mg/L	2	RS20	off
mLs 8-3 to 4-5 (mLs 4-5 excel)	= 'RS.mLs to 4-5' - 'RS.mLs to 8-3'	mL	2	RS21	off

Result name . . . . . **Cond**
Formula . . . . . =1000000 * TextToNumber('MV.cond')
Unit . . . . . µS/cm
Decimal places 2
Assignment RS01
Statistics . . . . . off
Description . . . . . .
Result monitoring . . . . . off
Save result as common variable . . . . . off
Name . . . . . .
Save result as titer . . . . . off
Solution name . . . . . .

Result name . . . . . **Cond Temp**
Formula . . . . . =TextToNumber('MV.temp')
Unit . . . . . °C
Decimal places 1
Assignment RS02
Statistics . . . . . off
Description . . . . . .
Result monitoring . . . . . off
Save result as common variable . . . . . off
Name . . . . . .
Save result as titer . . . . . off
Solution name . . . . . .

Result name . . . . . **pH**
Formula . . . . . = 'pH.EME'
Unit . . . . . pH units
Decimal places 2

Assignment RS03
Statistics . . . . . off
Description . . . . . RS.'Result name'[.VAL] Result value.
Result monitoring . . . . . off
Save result as common variable . . . . . off
Name . . . . . -
Save result as titer . . . . . off
Solution name . . . . . -

Result name . . . . . **pH Temp**
Formula . . . . . = 'pH.ETE'
Unit . . . . . °C
Decimal places 1
Assignment RS04
Statistics . . . . . off
Description . . . . . -
Result monitoring . . . . . off
Save result as common variable . . . . . off
Name . . . . . -
Save result as titer . . . . . off
Solution name . . . . . -

Result name . . . . . **mLs to 8-3**
Formula . . . . . = Case('to 8-3.EP{1}.VOL' ; 'to 8-3.EP{1}.VOL' ;0;0)
Unit . . . . . mL
Decimal places 2
Assignment RS05
Statistics . . . . . off
Description . . . . . -
Result monitoring . . . . . off
Save result as common variable . . . . . off
Name . . . . . -
Save result as titer . . . . . off
Solution name . . . . . -

Result name . . . . . **mLs to 4-5**
Formula . . . . . = 'to 8-3.FP{1}.VOL'
Unit . . . . . mL
Decimal places 2
Assignment RS06
Statistics . . . . . off
Description . . . . . -
Result monitoring . . . . . off
Save result as common variable . . . . . off
Name . . . . . -
Save result as titer . . . . . off

Solution name . . . . . .

Result name . . . . . **mLs to 4-2**
Formula . . . . . = 'to 8-3.EP{2}.VOL'
Unit . . . . . mL
Decimal places 2
Assignment RS11
Statistics . . . . . off
Description . . . . . .
Result monitoring . . . . . off
Save result as common variable . . . . . off
Name . . . . . .
Save result as titer . . . . . off
Solution name . . . . . .

Result name . . . . . **Total Alk**
Formula . . . . . = ('to 8-3.FP{1}.VOL' * 0.02 * 50000) / 50
Unit . . . . . mg/L
Decimal places 2
Assignment RS07
Statistics . . . . . off
Description . . . . . RS.'Result name'[.VAL] Result value.
Result monitoring . . . . . off
Save result as common variable . . . . . off
Name . . . . . .
Save result as titer . . . . . off
Solution name . . . . . .

Result name . . . . . **OH raw**
Formula . . . . . = (((2 * Case('to 8-3.EP{1}.VOL' ; 'to 8-3.EP{1}.VOL' ;0;0)) - 'to 8-3.FP{1}.VOL') * 50000 * .02) / 50
Unit . . . . . mg/L
Decimal places 2
Assignment . . . . . none
Statistics . . . . . off
Description . . . . . Calculates OH value from (2P-t)*50000*N/V for any P not equal to zero.
Result monitoring . . . . . off
Save result as common variable . . . . . off
Name . . . . . .
Save result as titer . . . . . off
Solution name . . . . . .

Result name . . . . . **OH**
Formula . . . . . =Case('RS.OH raw'< 0;0;'RS.OH raw')
Unit . . . . . mg/L

Decimal places 2
Assignment RS08
Statistics . . . . . off
Description . . . . . If 'OH raw' is less than zero (negative number), then 'OH' equals 0. If 'OH raw' is zero or positive, then 'OH' result is equal to "OH raw' value.
Result monitoring . . . . . off
Save result as common variable . . . . . off
Name . . . . . -
Save result as titer . . . . . off
Solution name . . . . . -

Result name CO3
Formula . . . . . = (2 * Case('to 8-3.EP{1}.VOL' ; 'to 8-3.EP{1}.VOL' ;0;0) * 50000 * 0.02) / 50
Unit . . . . . mg/L
Decimal places 2
Assignment RS09
Statistics . . . . . off
Description . . . . . -
Result monitoring . . . . . off
Save result as common variable . . . . . off
Name . . . . . -
Save result as titer . . . . . off
Solution name . . . . . -

Result name . . . . . HCO3 raw
Formula . . . . . = (('to 8-3.FP{1}.VOL' - (2 * Case('to 8-3.EP{1}.VOL' ; 'to 8-3.EP{1}.VOL' ;0;0))) * 50000 * 0.02) / 50
Unit . . . . . mg/L
Decimal places 2
Assignment . . . . . none
Statistics . . . . . off
Description . . . . . Calculates HCO3 value from (T-2P)*50000*N/V for any P not equal to zero.
Result monitoring . . . . . off
Save result as common variable . . . . . off
Name . . . . . -
Save result as titer . . . . . off
Solution name . . . . . -

Result name HCO3
Formula . . . . . =Case('RS.HCO3 raw'<0;0;'RS.HCO3 raw')
Unit . . . . . mg/L
Decimal places 2
Assignment RS10
Statistics . . . . . off

Description . . . . . If 'HCO3 raw' is less than zero (negative number), then 'HCO3' equals 0. If 'HCO3 raw' is zero or positive, then 'HCO3' result is equal to 'HCO3 raw'.

Result monitoring . . . . . off

Save result as common variable . . . . . off

Name . . . . . .

Save result as titer . . . . . off

Solution name . . . . . .

Result name . . . . . **Low Level Titration**

Formula . . . . . = (((2 * 'to 8-3.FP{1}.VOL') - 'to 8-3.EP{2}.VOL') * 0.02 * 50000) / 50

Unit . . . . . mg/L

Decimal places 2

Assignment RS12

Statistics . . . . . off

Description . . . . . .

Result monitoring . . . . . off

Save result as common variable . . . . . off

Name . . . . . .

Save result as titer . . . . . off

Solution name . . . . . .

Result name . . . . . **est TDS**

Formula . . . . . =0.66 * 'RS.Cond'

Unit . . . . . mg/L

Decimal places 0

Assignment RS13

Statistics . . . . . off

Description . . . . . estimated Total Dissolved Solids (TDS) from Electical Conductivity (EC)

Result monitoring . . . . . off

Save result as common variable . . . . . off

Name . . . . . .

Save result as titer . . . . . off

Solution name . . . . . .

Result name . . . . . **Free CO2/L**

Formula . . . . . = 2.0 * 'RS.HCO3' * (10 ^ (6- 'RS.pH'))

Unit . . . . . mg/L

Decimal places 2

Assignment RS19

Statistics . . . . . off

Description . . . . . .

Result monitoring . . . . . off

Save result as common variable . . . . . off

Name . . . . . .

Save result as titer ..... off
Solution name .....

Result name ..... **Total CO2/L**
Formula ..... = 'RS.Free CO2/L' + (.44 * ((2 * 'RS.HCO3') + 'RS.CO3'))
Unit ..... mg/L
Decimal places 2
Assignment RS20
Statistics ..... off
Description .....
Result monitoring ..... off
Save result as common variable ..... off
Name .....
Save result as titer ..... off
Solution name .....

Result name ..... **mLs 8-3 to 4-5 (mLs 4-5 excel)**
Formula ..... = 'RS.mLs to 4-5' - 'RS.mLs to 8-3'
Unit ..... mL
Decimal places 2
Assignment RS21
Statistics ..... off
Description ..... total mL to 4.5 - mL to 8.3. This gives you the amt. of titrant used to get from 8.3 to 4.5 pH units. This value can be plugged into the excel sheet as a double check.
Result monitoring ..... off
Save result as common variable ..... off
Name .....
Save result as titer ..... off
Solution name .....

REPORT **Report 233**

Report template
Report template ..... New Results Report
Report output
Printer ..... on
Printer ..... Default printer
PDF file ..... off

DATABASE **tiamo**

Database

Results

SERIES START **Series start track**

RACK **Initialize rack**

Device
Device name ..... 855_1
Rack test ..... off

SEQUENCE **Init Rinse Sequence**

PUMP **Pump On**
Device
Device name ..... 855_1
Pumps
Tower 1
Pump(s) 2
Action
Switch on ..... on
Switch off ..... off
Duration ..... off

WAIT **Wait 8s**
Wait
Stop track and waiting for [Continue] ..... off
Stop all tracks and waiting for [Continue] ..... off
Waiting time ..... on
Time 8
Unit ..... s
Message
Record message ..... off
Message by e-mail ..... off
Acoustic signal ..... off

LQH **Make room in Dosino**
General/Hardware
Device
Device name ..... 855_1
Dosing device
Dosing device 1
Solution ..... Transfer Pipette
Parameters
Function ..... Eject to end volume
Port 3
Rate ..... maximum mL/min

LQH **Fill w- air**
General/Hardware
Device
Device name ..... 855_1
Dosing device
Dosing device 1
Solution ..... Transfer Pipette
Parameters

Function . . . . . Aspirate
Port 1
Volume . . . . . 15.00 mL
Rate . . . . . maximum mL/min

LQH **Eject to EV**

General/Hardware

Device

Device name . . . . . 865_1

Dosing device

Dosing device 1

Solution . . . . . Transfer Pipette

Parameters

Function . . . . . Eject to end volume
Port 3
Rate . . . . . maximum mL/min

WAIT **Wait 3s**

Wait

Stop track and waiting for [Continue] . . . . . off
Stop all tracks and waiting for [Continue] . . . . . off
Waiting time . . . . . on
Time 3
Unit . . . . . s

Message

Record message . . . . . off
Message by e-mail . . . . . off
Acoustic signal . . . . . off

PUMP **Rinse 3s**

Device

Device name . . . . . 865_1

Pumps

Tower 1
Pump(s) 1

Action

Switch on . . . . . off
Switch off . . . . . off
Duration . . . . . on
Time . . . . . 3.0 s

WAIT **Wait 8 s**

Wait

Stop track and waiting for [Continue] . . . . . off
Stop all tracks and waiting for [Continue] . . . . . off
Waiting time . . . . . on
Time 8
Unit . . . . . s

Message

Record message ..... off
Message by e-mail ..... off
Acoustic signal ..... off

PUMP **Asp Pump Off**

Device

Device name ..... 855_1

Pumps

Tower 1
Pump(s) 2

Action

Switch on ..... off
Switch off ..... on
Duration ..... off

SERIES END **Series end track**

LIFT **Lift to Home**

Device

Device name ..... 855_1

Target

Tower 1
Lift position ..... Home position mm

Parameters

Lift rate ..... 25 mm/s

PUMP **Fill Vessel with water**

Device

Device name ..... 855_1

Pumps

Tower 1
Pump(s) 1

Action

Switch on ..... off
Switch off ..... off
Duration ..... on
Time ..... 4.0 s

LQH **Fill Dosino**

General/Hardware

Device

Device name ..... 855_1

Dosing device

Dosing device 1
Solution ..... Transfer Pipette

Parameters

Function ..... Fill

Port 2
Rate. . . . . maximum mL/min

Hall Environmental Analysis Laboratory

SOP Acknowledgement Sheet
SOP Name-SM2540C
SOP Number- S-SM2540C-5

I state that I have read the above referenced SOP. I understand all portions of the document as they apply to me and my responsibilities at Hall Environmental Analysis Laboratory. I further recognize this as the most current application of the SOP and that it supersedes any previous documents that I may have utilized.

Effective 3/4/11 - 9/14/11 [illegible] 10/20/11

Name	Signature	Date	Supervisor Initials	QA Officer Initials
Tiffany Shaw	[illegible]	3/9/11	[illegible]	[illegible]
Tom Francish	[illegible]	3/8/11	[illegible]	[illegible]
Ian Cameron	[illegible]	3/1/11		[illegible]
Kevin Sangster	[illegible]	3/1/11	[illegible]	[illegible]
Shelley Vogler	[illegible]	3/15/11	[illegible]	[illegible]
Lavonda Begay	[illegible]	6/~~22~~ 23/11 UD 6/23/11	[illegible]	[illegible]
Shelley Vogler	[illegible]	7/5/11		[illegible]

Hall Environmental Analysis Laboratory

Common Name: Total Dissolved Solids	S.O.P. Number: S-SM2540C-5
Method: SM2540C	
Effective Date: March 4, 2011	Supersedes: S-SM2540C-4
Laboratory Manager:	Quality Assurance/Quality Control Officer:

1. Scope and Application
   1.1. The method is applicable to drinking, surface and saline waters, and domestic and industrial wastes.
   1.2. The practical range of the determination is 10mg/L to 20,000mg/L.
2. Summary of Method
   2.1. A well-mixed sample, usually 100mL, is filtered through a glass fiber filter. The filtrate is evaporated and dried to constant weight at 180° C.
3. Detection Limits
   3.1. The PQL for TDS is 20 mg/L
   3.2. An MDL study is conducted annually. MDLs are updated in LIMS.
4. Definitions
   4.1. Practical Quantitation Limit (PQL) - Minimum detectable amount that can be reported with the appropriate precision.
   4.2. Matrix blank (MB) - DI water taken through the filtration and drying process.
   4.3. Laboratory Control Spike (LCS) - DI spiked with NaCl spike solution and taken through the filtration and drying process.
   4.4. Matrix Spike (MS) - Unknown sample spiked with NaCl spike solution and taken through the filtration and drying process.
   4.5. Matrix Spike Duplicate (MSD) – Duplicate of an unknown sample spiked with NaCl solution and taken through the filtration and drying process.
   4.6. Method detection limit (MDL) - The constituent concentration that when processed through the complete method, produces an analyte concentration with a 99% probability that it is different from the blank.
   4.7. DI Water (DI) – De-ionized water/reagent free water. Water that is passed through a purification system and is free from interferences.
5. Interferences

5.1. If the samples contain high levels of suspended solids it may be difficult to filter. Dilute the sample into DI water. 5X, 10X, or 20X are generally most appropriate. However extremely difficult samples may require a higher dilution. Always dry a few extra beakers in case a difficult sample is encountered.

5.2. Highly mineralized waters containing significant concentrations of calcium, magnesium, chloride, and /or sulfate may be hygroscopic and may require prolonged drying, desiccation, and rapid weighing.

5.3. Samples containing high concentrations of bicarbonate will require careful, and possibly prolonged, drying at 180° C to insure that all the bicarbonate is converted to carbonate.

5.4. Too much residue in the evaporating dish will crust over and entrap water that will not be driven off during the drying. When practical, total residue should be limited to about 2000mg. Most samples will require dilution to make it easier to dry the samples to a constant weight. Appropriate dilution factors can be determined by looking up historical data in LIMS or by calculating an estimated TDS value from the EC values. The auto-titrator will calculate the estimated TDS value for you from EC results. Est. TDS value = EC value in $\mu S \times 0.66$.

5.5. To gauge the effectiveness of the filtration in removing suspended solids, the spike standard is prepared using a soluble salt (NaCl) and an insoluble material (diatomaceous earth).

6. Safety

6.1. For handling procedures and exposure information, always refer to the MSDS. To keep exposure to reagents and standards to a minimum, analysts must wear appropriate laboratory safety attire when working in the laboratory.

7. Apparatus and Materials

7.1. VWR glass microfiber filter, 696. 4.7 cm

7.2. Filtration Apparatus

7.3. Drying oven, for 104 °C and 180°C $\pm$ 2°C. Calibration checked monthly.

7.4. Desiccator with color indicating desiccant or an Auto-dessicator.

7.5. Analytical balance, capable of weighing to 0.0001g.

7.6. Vacuum Pump

7.7. 100mL graduated cylinder

7.8. Forceps for handling filters

8. Reagents and Standards

8.1. DI water

8.2. TDS Spike

8.2.1. A combined 100,000mg/L NaCl/50,000mg/L diatomaceous earth standard is used as a spike.

8.2.1.1. Prepare standard by dissolving 10.00 g of NaCl salt and 5.0g diatomaceous earth into 100mL DI. Mix standard thoroughly. Diatomaceous earth will not go into

solution. A 1mL spike is equivalent to 1,000mg/L dissolved solids and 500mg/L suspended solids.

9. Sample Collection, Preservation, and Storage

9.1. Samples are to be collected in unpreserved plastic containers. Samples are stored at above freezing to 6°C until analysis.

9.2. Holding time until sample filtration is seven days.

10. Quality Control

10.1. A QC batch consists of an MB, LCS, MS, and MSD per 20 samples.

11. Procedure

11.1. See **Appendix A** for a copy of the bench sheet.

11.2. Preparing Beakers

11.2.1. Beaker's IDs are re-written if necessary after being cleaned and before being dried.

11.2.2. Clean beakers are placed in aluminum trays (holds 12 beakers) and are dried for 1 hour at 180°C±2°C.

11.2.3. Once dried, beakers are placed in a dessicator to cool. Warm beakers will drop in mass rapidly as they are being weighed. Should this occur, additional time in the dessicator is required.

11.3. Batch Preparation

11.3.1. Remove samples from the refrigerator and allow them to come to room temperature (a minimum of 2 hours).

11.3.2. A "2540C_PREP" prep batch is set up in LIMS. All pertinent information is included (reagents, spike, samples, and QC etc.)

11.3.3. The Solids Bench Sheet is filled out using the sample IDs from the prep batch in LIMS

11.3.4. Beakers are weighed with clean, dry gloves, and their IDs and masses recorded (to 0.0001 g) on the Solids Bench Sheet.

11.4. Filtration

11.4.1. The filtration apparatus is cleaned prior to every sample. The filter may be reused until it is clogged or poses a risk of tearing.

11.4.2. Assemble the filtration apparatus by placing the glass fiber filter on the apparatus with the smooth side facing down.

11.4.3. Condition a new filter by assembling the filtration apparatus, applying vacuum, and washing the filter with three successive 10 to 20mL volumes of DI water. Apply vacuum until no traces of DI are visible on filter disc. Disassemble and discard washings. Rinse vessel thoroughly with DI.

11.4.4. For MB, measure 100mL DI water in a graduated cylinder, pour into filtration apparatus, and start vacuum.

11.4.5. For LCS sample, measure 100mL DI in a graduated cylinder, spike with 1ml of the well-mixed Spike solution, pour into filtration apparatus, and start vacuum.

11.4.6. For client samples, shake vigorously until all sediment is suspended. Using a graduated cylinder, decant 100mL of sample, or appropriate amount of sample for your dilution. Dilute sample to a final volume of 100mL (with DI), pour into filtration apparatus, and start vacuum.

11.4.7. For MS/MSD samples, pour 100mL (or a dilution there of) of a well-mixed sample into a graduated cylinder. Spike with 1mL of spike solution and pour into filtration apparatus. Start vaccuum.

11.4.8. Record volumes used, if other than 100mL, on the Solids Bench Sheet.

11.4.9. Rinse graduated cylinder 3 times with DI from a squirt bottle and filter rinsate. Try to use approximately 10mL of DI.

11.4.10. Apply vacuum until the entire sample (including rinsate from graduated cylinder) has been filtered.

11.4.11. Rinse the filter with a small amount of DI from squirt bottle. Apply vacuum until no traces of moisture are visible.

11.4.12. Pour filtrate from the filtration apparatus into the appropriately labeled beaker. Rinse the filtration apparatus with a small amount of DI and pour rinsate into beaker.

11.4.13. Rinse the filtration apparatus and graduated cylinder thoroughly with DI water at the sink.

11.4.14. If filter is clean enough to filter another sample, re-assemble the apparatus and filter another sample. There is no need to use a new filter. If, however, the filter is heavily laden with suspended solids, a new conditioned filter (see 11.6) must be used.

11.4.15. Place beakers in the oven at 104° C ± 1°C overnight.

11.5. Drying of Samples

11.5.1. The following morning, check the beakers to make sure all the moisture has evaporated. If not, continue drying at 104 C° ± 1°C until all moisture has evaporated.

11.5.2. Once the samples are dry, adjust the oven temperature to 180° C ± 2° C and heat the residue for 2 hours (allow additional drying time if there is a large amount of residue present).

11.5.3. Using the cotton autoclave gloves, remove samples from oven and allow them to cool for a few seconds (≈ 15-30 seconds- only long enough so they do not melt/warp the shelves of the desiccator). Allow beakers to cool in desiccator for **at least** 2 hours They must be **cool** to the touch before you can weigh them.

11.5.4. Using a fresh pair of dry gloves, remove the samples from the desiccator and promptly weigh them on the analytical balance to the nearest 0.0001g. Check the beaker number to make sure the beaker plus residue weight is being recorded in the correct place on the Solids Bench Sheet.

11.5.5. Re-dry the samples at 180° ± 2° for 1-2 hours. Cool in the desiccator for at least 2 hours. They must be **cool** to the touch before you can weigh them. The weights must be within

0.0020g since the PQL for TDS is 20 mg/L. If not, repeat the drying, cooling, and weighing. Limit re-drying to 3 times.

11.5.6. Fill out tare weights, sample volumes, and final tare + residue weights in Excel spreadsheet. Key final calculated results into LIMS.

12. Calculation

12.1. Calculate filterable residue as follows:

$$\text{Total Solids residue in mg/L} = (x_{1(g)} - x_{2(g)}) \times \frac{1000(mg)}{1(g)} \times \frac{1}{x_{3(mL)}} \times \frac{1000(mL)}{1(L)}$$

Where:

X_1= Final mass of dried beaker and sample (g)
X_2= Mass of clean dry beaker (g)
X_3= volume of sample used in mL

13. Method Performance

13.1. Method Performance

13.1.1. PT or "blind study" samples are routinely analyzed to check the accuracy of the TDS procedure.

13.2. Estimation of Uncertainty

13.2.1. Estimation of uncertainty is to be performed on an annual basis.

13.2.1.1. Collect the most recent 25 LCS percent recoveries. See HEAL Quality Assurance Plan for details on the calculation of uncertainty.

13.3. Demonstration of Capability (DOC)

13.3.1. Demonstration of Capability (DOC) samples are analyzed before an analyst is able to independently generate data.

13.3.2. DOCs for TDS consist of four replicate LCS analyses. Average percent recovery and relative standard deviation are then calculated and must pass within ±20% and 20%, respectively. A detailed description of DOCs can be found in the HEAL QA Plan.

13.3.3. ADOCP are done to ensure the analyst is continually proficient. This is outlined in the HEAL QA Plan.

14. Pollution Prevention

14.1. To minimize waste, the filter may be reused if the filter appears to be clean enough to allow another sample to pass through it.

14.2. Beakers used for drying samples are cleaned and reused.

14.3. Sample volume is limited to 100mL of sample.

15. Data Assessment and Acceptance Criteria for Quality Control Measures
    15.1. See **Appendix B** for a summary of the batch QC requirements.
    15.2. MB not to exceed 20mg/L.
    15.3. LCS recovery must be between 80-120%.
    15.4. MS and MSD recoveries must be between 80-120%.
    15.5. The RPD between MS and MSD samples must be less than 20%.
    15.6. The difference between consecutive dry weights must be within 0.0020g.
16. Corrective Actions For Out Of Control Data
    16.1. If MB exceeds acceptable limit, the samples must be re-filtered, if possible.
    16.2. If LCS fails high or low, all samples must be re-filtered, if possible.
    16.3. If dried samples do not come within the 0.0020g after the 3 dryings at 180°C flag data and report.
    16.4. If the MS or MSD fails high or low, re-prep, if possible. Otherwise flag data and report.
    16.5. If the RPD between MS and MSD is greater the 20%, re-prep, if possible. Otherwise flag data and report.
17. Contingencies for Handling Out-of-Control Data
    17.1. All out-of-control data is flagged and reported to the client.
    17.2. Re-prepare samples out of holding time, compare results, and notify client.
18. Waste Management
    18.1. All TDS residues are disposed of down the drain.
19. References
    19.1. Standard Methods for the Examination of Water and Wastewater, 20th Edition, Method 2540C, (1999).
    19.2. EPA Method 160.1 – Residue, Filterable (Gravimetric, Dried at 180° C). NPDES Editorial Revision 1971.
    19.3. Hall Environmental Quality Assurance Plan

Hall Environmental Analysis Laboratory

Appendix A

Solids Bench Sheet

Solids Bench Sheet

Analysis: SM2540D (TSS) SM2540C (TDS) SM2540B (TS) ***PREP:*** _____

		Estimated TDS	mls Sample	Beaker ID	Beaker Mass (g)	Beaker +Sample Mass (1st Final Mass)	2nd Final Mass (g)	3rd Final Mass (g)	4th Final Mass (g)
1.						Date/Time:			
2.									
3.									
4.									
5.									
6.									
7.									
8.									
9.									
10.									
11.									
12.									
13.									
14.									
15.									
16.									
17.									
18.									
19.									
20.									
21.									
22.									
23.									
24.									

Pipet Information

Pipet No.: _____

Spike Volume: _____

Weight DI (g): _____

Room Temp.: _____ °C

Date: _____

Oven ID: _____

Thermometer ID: _____

Temp: _____

Start Time: _____

Stop Time: _____

Balance ID: _____

Analyst/Date: _____

QA'd/Date: _____

Appendix B

SM2540C: Total Dissolved Solids
Corrective Action Procedures

Analytical Method	Parameter	QC Element	Frequency	Acceptance Criteria	Corrective Action
SM2540C	Wet Chemistry	Method Blank	1 per preperation/analysis batch (≤ 20 samples)	< PQL	1. Reprep all samples in batch (if enough sample is available) 2. All samples must be flagged
		Laboratory Control Sample (LCS)	1 per preperation/analysis batch (≤ 20 samples)	% Recovery +/- 20% of actual value	1. Reprep all samples in batch (if enough sample is available) 2. All samples must be flagged
		Matrix Spike (MS)	1 per preperation/analysis batch (≤ 20 samples)	% Recovery +/- 20% of actual value	1. Flag failures and report.
		Matrix Spike Duplicate (MSD)	1 per preperation/analysis batch (≤ 20 samples)	% Recovery +/- 20% of actual value Relative percent difference (RPD) ≤20%	1. Flag failures and report.
		Dried to a constant weight within 3 re-dryings	All samples and QC samples	Weights must be within +/- 0.0020 g	1. If unable to get weights within range after 3 re-dryings, flag data and report.

Hall Environmental Analysis Laboratory

Common Name: Total Dissolved Solids	S.O.P. Number: S-SM2540C-6
Method: SM2540C	
Effective Date: September 14, 2011	Supersedes: S-SM2540C-5
Laboratory Manager:	Quality Assurance/Quality Control Officer:

1. Scope and Application
   1.1. The method is applicable to drinking, surface and saline waters, and domestic and industrial wastes.
   1.2. The practical range of the determination is 10mg/L to 20,000mg/L.
2. Summary of Method
   2.1. A well-mixed sample, usually 100mL, is filtered through a glass fiber filter. The filtrate is evaporated and dried to constant weight at 180° C.
3. Detection Limits
   3.1. The PQL for TDS is 20 mg/L
   3.2. An MDL study is conducted annually. MDLs are updated in the LIMS.
4. Definitions
   4.1. Practical Quantitation Limit (PQL) - Minimum detectable amount that can be reported with the appropriate precision.
   4.2. Matrix blank (MB) - DI water taken through the filtration and drying process.
   4.3. Laboratory Control Spike (LCS) - DI spiked with NaCl spike solution and taken through the filtration and drying process.
   4.4. Matrix Spike (MS) - Unknown sample spiked with NaCl spike solution and taken through the filtration and drying process.
   4.5. Matrix Spike Duplicate (MSD) – Duplicate of an unknown sample spiked with NaCl solution and taken through the filtration and drying process.
   4.6. Method detection limit (MDL) - The constituent concentration that when processed through the complete method, produces an analyte concentration with a 99% probability that it is different from the blank.
   4.7. DI Water (DI) – De-ionized water/reagent free water. Water that is passed through a purification system and is free from interferences.
5. Interferences

5.1. If the samples contain high levels of suspended solids it may be difficult to filter. Dilute the sample into DI water. 5X, 10X, or 20X are generally most appropriate. However extremely difficult samples may require a higher dilution. Always dry a few extra beakers in case a difficult sample is encountered.

5.2. Highly mineralized waters containing significant concentrations of calcium, magnesium, chloride, and /or sulfate may be hygroscopic and may require prolonged drying, desiccation, and rapid weighing.

5.3. Samples containing high concentrations of bicarbonate will require careful, and possibly prolonged, drying at 180° C to insure that all the bicarbonate is converted to carbonate.

5.4. Too much residue in the evaporating dish will crust over and entrap water that will not be driven off during the drying. When practical, total residue should be limited to about 2000mg. Most samples will require dilution to make it easier to dry the samples to a constant weight. Appropriate dilution factors can be determined by looking up historical data in LIMS or by calculating an estimated TDS value from the EC values. The auto-titrator will calculate the estimated TDS value for you from EC results. Est. TDS value = EC value in µS × 0.66.

5.5. To gauge the effectiveness of the filtration in removing suspended solids, the spike standard is prepared using a soluble salt (NaCl) and an insoluble material (diatomaceous earth).

6. Safety

6.1. For handling procedures and exposure information, always refer to the MSDS. To keep exposure to reagents and standards to a minimum, analysts must wear appropriate laboratory safety attire when working in the laboratory.

7. Apparatus and Materials

7.1. VWR glass microfiber filter, 696. 4.7 cm

7.2. Filtration Apparatus

7.3. Drying oven, for 104 ° C and 180°C ± 2°C. Calibration checked monthly.

7.4. Desiccator with color indicating desiccant or an Auto-dessicator.

7.5. Analytical balance, capable of weighing to 0.0001g.

7.6. Vacuum Pump

7.7. 100mL graduated cylinder

7.8. Forceps for handling filters

8. Reagents and Standards

8.1. DI water

8.2. TDS Spike

8.2.1. A combined 100,000mg/L NaCl/50,000mg/L diatomaceous earth standard is used as a spike.

8.2.1.1. Prepare standard by dissolving 10.00 g of NaCl salt and 5.0g diatomaceous earth into 100mL DI. Mix standard thoroughly. Diatomaceous earth will not go into solution. A 1mL spike is equivalent to 1,000mg/L dissolved solids and 500mg/L suspended solids.

9. Sample Collection, Preservation, and Storage

   9.1. Samples are to be collected in unpreserved plastic containers. Samples are stored at above freezing to 6°C until analysis.

   9.2. Holding time until sample filtration is seven days.

10. Quality Control

   10.1. A QC batch consists of an MB, LCS, MS, and MSD per 20 samples.

11. Procedure

   11.1. See **Appendix A** for a copy of the bench sheet.

   11.2. Preparing Beakers

      11.2.1. Beaker's IDs are re-written if necessary after being cleaned and before being dried.

      11.2.2. Clean beakers are placed in aluminum trays (holds 12 beakers) and are dried for 1 hour at 180°C±2°C.

      11.2.3. Once dried, beakers are placed in a dessicator to cool. Warm beakers will drop in mass rapidly as they are being weighed. Should this occur, additional time in the dessicator is required.

   11.3. Batch Preparation

      11.3.1. Remove samples from the refrigerator and allow them to come to room temperature (a minimum of 2 hours).

      11.3.2. A "2540C_PREP" prep batch is set up in LIMS. All pertinent information is included (reagents, spike, samples, and QC etc.)

      11.3.3. The Solids Bench Sheet is filled out using the sample IDs from the prep batch in LIMS

      11.3.4. Beakers are weighed with clean, dry gloves, and their IDs and masses recorded (to 0.0001 g) on the Solids Bench Sheet.

   11.4. Filtration

      11.4.1. The filtration apparatus is cleaned prior to every sample. The filter may be reused until it is clogged or poses a risk of tearing.

      11.4.2. Assemble the filtration apparatus by placing the glass fiber filter on the apparatus with the smooth side facing down.

      11.4.3. Condition a new filter by assembling the filtration apparatus, applying vacuum, and washing the filter with three successive 10 to 20mL volumes of DI water. Apply vacuum until no traces of DI are visible on filter disc. Disassemble and discard washings. Rinse vessel thoroughly with DI.

      11.4.4. For MB, measure 100mL DI water in a graduated cylinder, pour into filtration apparatus, and start vacuum.

      11.4.5. For LCS sample, measure 100mL DI in a graduated cylinder, spike with 1ml of the well-mixed Spike solution, pour into filtration apparatus, and start vacuum.

      11.4.6. For client samples, shake vigorously until all sediment is suspended. Using a graduated cylinder, decant 100mL of sample, or appropriate amount of sample for your dilution.

Dilute sample to a final volume of 100mL (with DI), pour into filtration apparatus, and start vacuum.

11.4.7. For MS/MSD samples, pour 100mL (or a dilution there of) of a well-mixed sample into a graduated cylinder. Spike with 1mL of spike solution and pour into filtration apparatus. Start vaccuum.

11.4.8. Record volumes used, if other than 100mL, on the Solids Bench Sheet.

11.4.9. Rinse graduated cylinder 3 times with DI from a squirt bottle and filter rinsate. Try to use approximately 10mL of DI.

11.4.10. Apply vacuum until the entire sample (including rinsate from graduated cylinder) has been filtered.

11.4.11. Rinse the filter with a small amount of DI from squirt bottle. Apply vacuum until no traces of moisture are visible.

11.4.12. Pour filtrate from the filtration apparatus into the appropriately labeled beaker. Rinse the filtration apparatus with a small amount of DI and pour rinsate into beaker.

11.4.13. Rinse the filtration apparatus and graduated cylinder thoroughly with DI water at the sink.

11.4.14. If filter is clean enough to filter another sample, re-assemble the apparatus and filter another sample. There is no need to use a new filter. If, however, the filter is heavily laden with suspended solids, a new conditioned filter (see 11.6) must be used.

11.4.15. Place beakers in the oven at 104° C ± 1°C overnight.

11.5. Drying of Samples

11.5.1. The following morning, check the beakers to make sure all the moisture has evaporated. If not, continue drying at 104 C° ± 1°C until all moisture has evaporated.

11.5.2. Once the samples are dry, adjust the oven temperature to 180° C ± 2° C and heat the residue for 2 hours (allow additional drying time if there is a large amount of residue present).

11.5.3. Using the cotton autoclave gloves, remove samples from oven and allow them to cool for a few seconds (≈ 15-30 seconds- only long enough so they do not melt/warp the shelves of the desiccator). Allow beakers to cool in desiccator for **at least** 2 hours They must be **cool** to the touch before you can weigh them.

11.5.4. Using a fresh pair of dry gloves, remove the samples from the desiccator and promptly weigh them on the analytical balance to the nearest 0.0001g. Check the beaker number to make sure the beaker plus residue weight is being recorded in the correct place on the Solids Bench Sheet.

11.5.5. Re-dry the samples at 180° ± 2° for 1-2 hours. Cool in the desiccator for at least 2 hours. They must be **cool** to the touch before you can weigh them. The weights must be within 0.0020g since the PQL for TDS is 20 mg/L. If not, repeat the drying, cooling, and weighing. Limit re-drying to 3 times.

11.5.6. Fill out tare weights, sample volumes, and final tare + residue weights in Excel spreadsheet. Key final calculated results into LIMS.

12. Calculation

12.1. Calculate filterable residue as follows:

$$\text{Total Solids residue in mg/L} = (x_{1(g)} - x_{2(g)}) \times \frac{1000(mg)}{1(g)} \times \frac{1}{x_{3(mL)}} \times \frac{1000(mL)}{1(L)}$$

Where:

X_1= Final mass of dried beaker and sample (g)
X_2= Mass of clean dry beaker (g)
X_3= volume of sample used in mL

13. Method Performance

13.1. Proficiency Testing Sample (PT)

13.1.1. PT or "blind study" samples are routinely analyzed to check the accuracy of the TDS procedure.

13.2.Method Detection Limits (MDLs)

13.2.1. MDLs are conducted annually in accordance with the procedure and calculations outlined in the current QAP

13.2.2. MDLs must be analyzed over a minimum of three days.

13.3. Estimation of Uncertainty

13.3.1. Estimation of uncertainty is to be performed on an annual basis.

13.3.1.1. Collect the most recent 25 LCS percent recoveries. See HEAL Quality Assurance Plan for details on the calculation of uncertainty.

13.4. Demonstration of Capability (DOC)

13.4.1. Demonstration of Capability (DOC) samples are analyzed before an analyst is able to independently generate data.

13.4.2. DOCs for TDS consist of four replicate LCS analyses. Average percent recovery and relative standard deviation are then calculated and must pass within ±20% and 20%, respectively. A detailed description of DOCs can be found in the HEAL QA Plan.

13.4.3. ADOCP are done to ensure the analyst is continually proficient. This is outlined in the HEAL QA Plan.

14. Pollution Prevention

14.1. To minimize waste, the filter may be reused if the filter appears to be clean enough to allow another sample to pass through it.

14.2. Beakers used for drying samples are cleaned and reused.

14.3. Sample volume is limited to 100mL of sample.

15. Data Assessment and Acceptance Criteria for Quality Control Measures

15.1. See **Appendix B** for a summary of the batch QC requirements.

15.2. MB not to exceed 20mg/L.

15.3. LCS recovery must be between 80-120%.

15.4. MS and MSD recoveries must be between 80-120%.

15.5. The RPD between MS and MSD samples must be less than or equal to 5%.

15.6. The difference between consecutive dry weights must be within 0.0020g.

16. Corrective Actions For Out Of Control Data

16.1. If MB exceeds acceptable limit, the samples must be re-filtered, if possible.

16.2. If LCS fails high or low, all samples must be re-filtered, if possible.

16.3. If dried samples do not come within the 0.0020g after the 3 dryings at 180°C flag data and report.

16.4. If the MS or MSD fails high or low, re-prep, if possible. Otherwise flag data and report.

16.5. If the RPD between MS and MSD is greater the 5%, re-prep, if possible. Otherwise flag data and report.

17. Contingencies for Handling Out-of-Control Data

17.1. All out-of-control data is flagged and reported to the client.

17.2. Re-prepare samples out of holding time, compare results, and notify client.

18. Waste Management

18.1. All TDS residues are disposed of down the drain.

19. References

19.1. Standard Methods for the Examination of Water and Wastewater, 20th Edition, Method 2540C, (1999).

19.2. EPA Method 160.1 – Residue, Filterable (Gravimetric, Dried at 180° C). NPDES Editorial Revision 1971.

19.3. Hall Environmental Quality Assurance Plan

Hall Environmental Analysis Laboratory

Appendix A

Solids Bench Sheet

Solids Bench Sheet

Analysis: SM2540D (TSS) SM2540C (TDS) SM2540B (TS) ***PREP:*** _____

	[redacted]	Estimated TDS	mls Sample	Beaker ID	Beaker Mass (g)	Beaker +Sample Mass (1st Final Mass)	2nd Final Mass (g)	3rd Final Mass (g)	4th Final Mass (g)
1.						Date/Time:			
2.									
3.									
4.									
5.									
6.									
7.									
8.									
9.									
10.									
11.									
12.									
13.									
14.									
15.									
16.									
17.									
18.									
19.									
20.									
21.									
22.									
23.									
24.									

Pipet Information

Pipet No.: _____

Spike Volume: _____

Weight DI (g): _____

Room Temp.: _____ °C

Analyst/Date: _____

QA'd/Date: _____

Date:	_____	_____	_____
Oven ID:		_____	_____
Thermometer ID:		_____	_____
Temp:		_____	_____
Start Time:		_____	_____
Stop Time:		_____	_____
Balance ID:	_____	_____	_____

Appendix B

SM2540C: Total Dissolved Solids
Corrective Action Procedures

Analytical Method	Parameter	QC Element	Frequency	Acceptance Criteria	Corrective Action
SM2540C	Wet Chemistry	Method Blank	1 per preperation/analysis batch (≤ 20 samples)	< PQL	1. Reprep all samples in batch (if enough sample is available) 2. All samples must be flagged
		Laboratory Control Sample (LCS)	1 per preperation/analysis batch (≤ 20 samples)	% Recovery +/- 20% of actual value	1. Reprep all samples in batch (if enough sample is available) 2. All samples must be flagged
		Matrix Spike (MS)	1 per preperation/analysis batch (≤ 20 samples)	% Recovery +/- 20% of actual value	1. Flag failures and report.
		Matrix Spike Duplicate (MSD)	1 per preperation/analysis batch (≤ 20 samples)	% Recovery +/- 20% of actual value Relative percent difference (RPD) ≤5%	1. Flag failures and report.
		Dried to a constant weight within 3 re-dryings	All samples and QC samples	Weights must be within +/- 0.0020 g	1. If unable to get weights within range after 3 re-dryings, flag data and report.

8-2

Hall Environmental Analysis Laboratory

Common Name: SM2540D	S.O.P. Number: S-SM2540D-3
Method: Total Suspended Solids	
Effective Date: March 4, 2011	Supersedes: S-SM2540D-2
Laboratory Manager:	Quality Assurance/Quality Control Officer:

1. Scope and Application

   1.1. This SOP. covers the determination of non-filterable residue in groundwater, domestic, and industrial wastes. Non-filterable residue is defined as those solids that are retained by a glass fiber filter and dried to a constant weight.

   1.2. The practical range of the determination is 10 mg/L to 2,000 mg/L.

2. Summary of Method

   2.1. A well-mixed sample, usually 100mL, is filtered through a glass fiber filter. The residue retained on the filter is dried to a constant weight at 103-105°C

   2.2. If Filterable Residue is to be determined, the filtrate may be dried according to SM2540C.

3. Detection Limits

   3.1. The PQL for TSS is 10 mg/L.

   3.2. A MDL study is not performed.

4. Definitions

   4.1. Practical Quantitation Limit (PQL) - Minimum detectable amount that can be reported with the appropriate precision.

   4.2. Matrix blank (MB) - DI water taken through the filtration and drying process.

   4.3. Sample Duplicate (DUP) – A preparation and analysis of a client sample to assess precision.

   4.4. Laboratory Controlled Sample- 100 mL aliquot of NSI ™ Mid-Level Solids Standard used to assess recovery and precision.

   4.5. Method detection limit (MDL) - The constituent concentration, that when processed through the complete method, produces an analyte concentration with a 99% probability that it is different from the blank.

   4.6. DI Water (DI) – De-ionized water/reagent free water. Water that is passed through a purification system and is free from interferences.

5. Interferences

   5.1. If the samples contain high levels of suspended solids it may be difficult to filter. Dilute the sample into DI water. 5x or 10x dilution are generally most appropriate. However, extremely

difficult samples may require a higher dilution. Always dry a few extra filters in case a difficult sample is encountered.

5.2. Samples containing suspended solids greater than 2000 mg/L must be re-prepared at dilution.

6. Safety

6.1. To keep exposure to reagents and standards to a minimum, analysts must wear appropriate laboratory safety attire when working in the laboratory. For handling procedures and exposure information always refer to the MSDS.

7. Apparatus and Materials

7.1. Pre-weighed 47 mm glass fiber filters and aluminum dish, Environmental Express Pro-Weigh™ Filters.

7.2. Filtration Apparatus

7.3. Drying oven, for 104 °C. Calibration checked monthly.

7.4. Desiccator with color indicating desiccant or an Auto-Desiccator

7.5. Analytical balance, capable of weighing to 0.0001g

7.6. Vacuum Pump

7.7. 100mL graduated cylinder.

7.8. Tweezers

8. Reagents and Standards

8.1. DI water

8.2. NSI Mid-Level Solids Standard

9. Sample Collection, Preservation, and Storage

9.1. Samples are to be collected in unpreserved plastic containers. Samples are stored at above freezing to 6°C until analysis.

9.2. HEAL holding time till sample filtration is seven days.

10. Quality Control

10.1. A QC batch consists of an MB, DUP, and a Laboratory Controlled Sample for every 10 samples.

11. Calibration

11.1. Calibrate the Analytical Balance as per the HEAL Analytical Balance Calibration SOP.

11.2. The Drying Oven is to be calibrated quarterly and recorded in the Drying Oven Calibration Log.

12. Procedure

12.1. See **Appendix A** for a copy of the Solids Bench Sheet.

12.2. Filter Preparation

12.2.1. Label aluminum filter dishes with a marker.

12.2.2. A filter must be prepared for each sample, MB, DUP, and Laboratory Controlled Sample. (Prepare additional filters in case a difficult sample is encountered).

12.2.3. Dry filters in their aluminum dishes in oven for one hour at 104° ±1°C

12.2.4. Place filters in desiccator and allow them to cool for at least 15 minutes. Avoid putting hot beakers or other glassware in desiccator with filters as it will increase the time it takes the filters to cool.

12.3. Sample Filtration

12.3.1. Remove samples from the refrigerator and allow them to come to room temperature (a minimum of 2 hours).

12.3.2. With fresh DRY gloves, remove the filters from the desiccator and promptly weigh them to the nearest 0.0001g. Record the ID and tare weight of the filter and aluminum dish on your Solids Bench Sheet.

12.3.3. Thoroughly rinse the filtration apparatus with DI water. Gently remove the filter from the aluminum dish and place on the filtration apparatus (smooth side facing down). GENTLY twist on the receiving cup. Be careful. Twisting it too tightly will tear the filter and result in inaccurate weights.

12.3.4. Start vacuum. Wet filter with a small amount of DI from the squirt bottle. Turn off vacuum. Filter is now ready for sample.

12.3.5. For MB, measure 100mL DI water in a graduated cylinder, pour into filtration apparatus and start the vacuum.

12.3.6. For client samples, shake vigorously until all sediment is suspended. Using a graduated cylinder, decant 100mL of sample (dilute sample if the amount of suspended solids may hinder filtration). Dilute sample to a final volume of 100mL (with DI) and pour into filtration apparatus and start vacuum.

12.3.7. For DUP sample, pour 100mL (or a dilution there of) of a well-mixed sample into a graduated cylinder, pour into filtration apparatus, and start vacuum.

12.3.8. For your Laboratory Controlled Sample, vigorously shake the Mid-Level Solids Standard bottle and invert several times. Pour 100 mL into filtration apparatus and start vacuum.

12.3.9. Record the volume used on TSS Bench Sheet. You will also need to enter these into LIMS after you complete your batch.

12.3.10. Rinse graduated cylinder 3 times with DI from a squirt bottle and filter rinsate. Try to use approximately 10mL of DI.

12.3.11. Apply vacuum until the entire sample (including rinsate from graduated cylinder) has been filtered. (NOTE: if filtration time exceeds 10 minutes discard filter and re-filter sample at dilution).

12.3.12. Rinse the sides of the filtration apparatus and the filter with a small amount of DI from squirt bottle. Apply vacuum until no traces of moisture are visible.

12.3.13. Gently remove filter from filtration apparatus with tweezers and return to the aluminum dish.

12.3.14. BE CAREFUL! Transfer the entire filter and residue from the filtration apparatus to the aluminum dish. Filter material left on the filtration apparatus will significantly affect the results.

12.3.15. Filtrate may be used for SM2540C (TDS) if appropriate glassware has been prepared. Otherwise discard filtrate.

12.3.16. Rinse the filtration apparatus thoroughly with DI water.

12.4. Drying the samples

12.4.1. Place aluminum dishes with filters in the oven at 104° C ±1°C. Dry for one hour. Record the oven temperature and the start and stop times on the Solids Bench Sheet.

12.4.2. Remove samples from oven and allow them to cool for at least 15 minutes in the desiccator.

12.5. Using a fresh pair of dry gloves, remove the samples from the desiccator and promptly weigh them on the analytical balance to the nearest 0.0001g. Check the ID number to make sure the weight is being recorded in the correct place on your Solids Bench Sheet.

12.6. Place the filters back in the oven for 1 hour and then back in the desiccator for at least 15 minutes to cool. Record the oven temperature and the start and stop times on the Solids Drying Temperature Cover Sheet.

12.7. Re-weigh the filters and record the weights on the bench sheet. The difference between the weights must be within 0.0005 gm. If not, repeat the drying and weighing process until the weights are within range. Limit re-drying to 3 times.

12.8. Record your final weights on your Solids Bench sheet and enter them into your Excel sheet. Hand key result into LIMS.

13. Calculation

13.1. Calculate non-filterable residue as follows:

$$\textit{Non-filterable residue, mg/L} = \frac{(A-B) \times 1{,}000{,}000}{C}$$

Where:
A= weight of residue + filter + aluminum dish (g)
B= weight of filter + aluminum dish (g)
C= volume of sample used in mL

14. Method Performance

14.1. Method Performance

14.1.1. PT, or "blind study", samples are routinely analyzed to check the accuracy of the TSS procedure.

14.2. Estimation of Uncertainty

14.2.1. Estimation of uncertainty is to be preformed on an annual basis.

14.2.2. Collect the most recent 25 LCS percent recoveries. Calculate the average percent recovery and standard deviation *s*. Calculate 95% confidence interval (2 times the standard deviation). Report the estimated uncertainty as +/- 2*s*. For details on estimation of uncertainty See HEAL QAP.

14.3. Demonstration of Capability (DOC)

14.3.1. Demonstration of Capability (DOC) consists of four replicate samples of Mid-Level Solids Standard. The acceptance range is given on the current Certificate of Analysis located in the Wet Chemistry Spikes and Standard Notebook. (The acceptance range may change each time new standards are ordered.)

14.3.2. Analyst is considered proficient after close supervision by a senior analyst and by examination of RPD results from DOC samples (RPD <15).

14.3.3. ADOCP are done to ensure analyst is considered to be continually proficient. This is outlined in the HEAL QAP.

15. Pollution Prevention

15.1. Filters are thrown away and aluminum dishes are recycled. Sample size is limited to 100 mL.

16. Data Assessment and Acceptance Criteria for Quality Control Measures

16.1. See **Appendix B** for a summary of the batch QC requirements.

16.2. MB not to exceed 10mg/L or be less than -5mg/L.

16.3. The RPD between sample and DUP samples must be less than 15%.

16.4. All samples must not have a concentration less than -5mg/L.

16.5. The difference between consecutive dry weights must be within 0.0005g.

17. Corrective Actions For Out Of Control Data

17.1. Matrix Blank

17.1.1. If MB has a concentration less than -5mg/L, all samples in the batch must be re-prepared (out of holding time if necessary). If there is not enough sample, flag data and report.

17.1.2. If the MB concentration is greater than 10mg/L, first re-dry the samples in the oven at 104°C 1 for an hour, allow them to cool in the desiccator, and re-weigh. If still greater than 10mg/L, re-prep all samples (out of holding time if necessary). If there is not enough sample, flag data and report.

17.2. If the RPD between sample and DUP is greater than 15%, assess data in the following manner:

17.2.1. If the sample or DUP concentration is less than 2 times the PQL, the concentrations must agree to within 3 mg/L. If they do, generate corrective action addressing RPD failure. If they do not, re-prep all samples from the same set of HEAL numbers. Due to sample volume limitations, it may be necessary to select a different sample to DUP. Re-prep out of holding time if necessary. If re-prep is not an option or if re-prepped samples do not agree to within 3mg/L, generate corrective action, flag, and report failure.

17.2.2. If the sample and DUP concentrations are greater than 2 times the PQL, re-prep all samples from the same set of HEAL numbers. Due to sample volume limitations, it may be necessary to select a different sample to DUP. Re-prep out of holding time if necessary. If re-prep is not an option, generate corrective action, flag, and report failure.

17.3. If the Laboratory Control Sample concentration does not fall between the acceptance range specified in the corresponding Certificate of Analysis for the lot # being used, re-prep the entire batch including the MB, DUP, and Laboratory Controlled Sample. Due to sample volume limitations, it may be necessary to select a different sample to DUP. Re-prep out of holding time if necessary.

17.4. If consecutive dry weights are not within 0.0005 g after 3 re-dryings, generate corrective action, flag data, and report.

18. Contingencies for Handling Out-of-control Data

18.1. All out-of-control data is flagged and reported to the client.

18.2. Re-prepare samples out-of-holding time, compare results, and notify client.

19. Waste Management

19.1. All TSS filters are disposed of in the trash. The Aluminum dishes are recycled.

20. References

20.1. Standard Methods for the Examination of Water and Wastewater, 20th Edition, Method 2540D, (1999).

20.2. HEAL Quality Assurance Plan

20.3. HEAL Balance Calibration SOP

Appendix A

Solids Bench Sheet

Solids Bench Sheet

Analysis: SM2540D (TSS) SM2540C (TDS) SM2540B (TS) ***PREP:*** _____

		Estimated TDS	mls Sample	Beaker ID	Beaker Mass (g)	Beaker +Sample Mass (1st Final Mass)	2nd Final Mass (g)	3rd Final Mass (g)	4th Final Mass (g)
1.						Date/Time:			
2.									
3.									
4.									
5.									
6.									
7.									
8.									
9.									
10.									
11.									
12.									
13.									
14.									
15.									
16.									
17.									
18.									
19.									
20.									
21.									
22.									
23.									
24.									

Pipet Information

Pipet No.: _____

Spike Volume: _____

Weight DI (g): _____

Room Temp.: _____ °C

Analyst/Date: _____

QA'd/Date: _____

Date: _____ _____ _____

Oven ID: _____ _____

Thermometer ID: _____ _____

Temp: _____ _____

Start Time: _____ _____

Stop Time: _____ _____

Balance ID: _____ _____ _____

Appendix B

SM2540D: Total Suspended Solids
Corrective Action Procedures

Analytical Method	Parameter	QC Element	Frequency	Acceptance Criteria	Corrective Action
SM2540D	Wet Chemistry	Method Blank	1 per preparation/analysis batch (≤ 10 samples)	>-5mg/L and <PQL	1. If <-5mg/L reprep all samples in batch (if enough sample is available) 2. If >PQL dry samples for an additional hour in oven, cool in desiccator, and reweigh. Reprep if still out. 3. All samples must be flagged and a corrective action generated.
		Sample Duplicate (DUP)	1 per preparation/analysis batch (≤ 10 samples)	Relative percent difference (RPD) ≤15%	1. If DUP concentration is greater than the PQL but is less than 2x the PQL the results must be within 3mg/L of each other. If they are generate corrective action addressing failure, do not reprep. If however they are not, all of the samples from the same HEAL number set and DUP must be re-prepared. (Due to sample volume limitations it may be necessary to select a different DUP sample for reprep). 2. If greater than 2x PQL reprep all samples from the same HEAL number set. (Due to sample volume limitations it may be necessary to select a different DUP sample for reprep).
		Laboratory Contolled Sample	1 per preparation/analysis batch (≤ 10 samples)	See the current Ceritficate of Analysis located in the Spikes and Standards Logbook for the Acceptance Limits.	1. If Laboratory Conrtolled Sample falls outside current Acceptance Limits for the lot # being used re-prepare all samples including the Laboratory Contolled Sample, MB, and DUP. (Due to sample volume limitations it may be necessary to select a different DUP sample for re-prep).

9

Hall Environmental Analysis Laboratory

SOP Acknowledgement Sheet
SOP Name-SM5310B/9060
SOP Number- S-TOC-4

I state that I have read the above referenced SOP. I understand all portions of the document as they apply to me and my responsibilities at Hall Environmental Analysis Laboratory. I further recognize this as the most current application of the SOP and that it supersedes any previous documents that I may have utilized.

Effective 12/14/10- 6/3/11 SC

Name	Signature	Date	Supervisor Initials	QA Officer Initials
John Caldwell	John Caldwell	12-8-10		
Steve Crandell		12-8-10		
Lynda Walls	Lynda Walls	12-8-10		
Michael Williams		12/10/10		

Hall Environmental Analysis Laboratory

Common Name: SM5310B Total Organic Carbon (Combustion Infrared Method) EPA9060 Total Organic Carbon	S.O.P. Number: S-TOC-4
Method: Total Organic Carbon	
Effective Date: 14 December 2010	Supersedes: S-TOC-3
Laboratory Manager:	Quality Assurance/Quality Control Officer:

1. Scope and Application

   1.1. This S.O.P. covers the determination of the concentration of Non-purgeable Organic Carbon (NPOC) in ground water, surface and saline waters, drinking water, and domestic and industrial wastes.

2. Summary of Method

   2.1. A 5-ml sample aliquot is drawn out of a 40mL, thin septa amber vial, acidified by HCl or H_2SO_4 to a pH<2, and sparged for 1.5 minutes. A 50ul aliquot of the sample is then injected onto a high sensitivity oxidation catalyst inside of the combustion chamber at 680°C. The sample is burned in the combustion chamber to produce CO_2 and other combustion products which are then swept through a dehumidifier and halogen scrubber to a non-dispersive infrared (NDIR) gas analyzer, where the CO_2 is detected. The analog detection signal of the NDIR forms a peak, and the area of this peak is measured by a data processor.

3. Detection Limits

   3.1. MDLs shall be empirically established annually. MDL is subject to change. See LIMS for current MDL.

   3.2. The current PQL is 1.0mg/L

4. Definitions

   4.1. Calibration Blank (CalibBlank) – Blank Solution of DI water to "zero" the instrument.

   4.2. Initial Calibration Verification 5 mg/L (ICV5) – Second source standard @ 5 mg/L analyzed after the initial calibration of the Low-level curve. It is also analyzed every 15 samples to check the stability of the instrument during analysis.

   4.3. Initial Calibration Verification 50 mg/L (ICV50) – Second source standard @ 50 mg/L analyzed after the initial calibration of the Med-level curve. It is also analyzed every 15 samples to check the stability of the instrument during analysis.

   4.4. Initial Calibration Blank (ICB) – Analysis of Calibration blank solution after initial calibration to check

the cleanliness of blank. Should report below PQL.

4.5. Laboratory Control Spike (LCS) – Laboratory fortified blank.

4.6. Matrix Spike (MS) – Laboratory fortified sample.

4.7. Matrix Spike Duplicate (MSD) – Laboratory fortified sample duplicate.

4.8. Matrix Blank (MBLK) – Laboratory blank.

4.9. Method Detection Limit (MDL) – The constituent concentration that when processed through the complete method, produces a signal with a 99% probability that it is different than the blank.

4.10. Practical Quantitation Limit (PQL)/Reporting Limit – The minimum limit to which an analyte can be routinely reported.

5. Interference

5.1. Carbonate and bicarbonate carbon (inorganic carbon) represent interferences and are removed by acidification and purging of the sample. An LCS fortified with Na_2CO_3 is analyzed to show that the inorganic carbon is removed.

5.2. Method interference may be caused by contaminants in the reagent water, sample processing chemicals or analysis apparatus.

5.3. This procedure is applicable only to homogeneous samples which can be injected into the apparatus reproducibly by means of a syringe. The opening of this syringe limits the maximum size of particle which may be included in the sample. If necessary, allow the sample to settle, decant liquid into a fresh VOA. Label with sample ID and HEAL number. Retain decanted sample with original sample containers.

5.4. Samples that are not drawn into the syringe easily may require dilution into DI water. This is evidenced by a large variance of replicate combustion readings during sample analysis.

6. Safety

6.1 At minimum, latex gloves, a lab coat, and safety glasses must be worn to avoid any unnecessary contact with samples and standards. Refer to the MSDS before handling any standards for safety concerns.

6.2 When the analysts are done with standard vials they need to be disposed of in the appropriate manner. All standards, analytical vials, samples, and other laboratory-generated waste will be disposed in accordance with the HEAL Chemical Hygiene Plan.

6.3 Refer to the safety/warning section of the instrument manual before running or maintaining the instrument to avoid burns or electrical shock.

7. Apparatus, Materials

7.1. Shimadzu TOC-V – Total Organic Carbon Analyzer.

7.1.1. NDIR: non-dispersive infrared gas analyzer detector.

7.2. Operating Conditions

7.2.1. Furnace Temperature: 680°C

7.2.2. Carrier gas set to 150 psi.

7.2.3. Purge time set to 1.5 minutes

7.2.4. Acid addition to 1.5% of 2.0 N HCl

7.2.5. Injection volume set to 50uL.

7.2.6. Method programmed to accept best 4 of 5 injections.

7.2.7. Shimadzu combustion tube (part no. 638-41323) with platinum mesh.

7.2.8. Shimadzu high sensitivity catalyst (part no. 630-00996).

7.3. Shimadzu ASI-V Automated Sampler with 40mL VOA auto sampler rack.

7.3.1. The ASI-V Automated Sampler powers on and off through the instrument. It is important to note that the auto sampler will not operate while the cover is off; and that removal of the cover during analysis will result in the halting of the analysis.

7.4. Analytical balance capable of weighing to the nearest 0.0001g

7.5. pH strips that measure to the nearest 0.5 of a pH unit.

7.6. Sample vials (40mL) thin septa VOA vials.

7.7. Hand-held re-pipettors of various dispensing volumes.

7.8. Volumetric flasks – 50mL, 100mL, 200mL, 250mL, and 500mL volumes.

7.9. Micro-syringes – 250mL and 500mL.

7.10. Variable volume 1mL handheld pipette.

7.11. Environmental Express Digestion Tubes – SC475.

8. Reagents and Standards

8.1. De-ionized water, free from organic carbons.

8.2. 2N HCl – Made from high purity OmniTrace Concentrated HCl. Add 41.25mL Conc. HCL to 100mL DI water in a 250mL volumetric flask. Bring to 250mL final volume with DI water. Reagent must be logged

into LIMS and recorded in TOC maintenance logbook.

8.3. Stock Standards

8.3.1. Primary source standard is a certified TOC standard purchased from Accustandard @ 1000mg/L.

8.3.2. Secondary source standard (ICV) is purchased from Ultra Scientific at a concentration of 1000mg/L.

8.3.3. Low Cal Stock (10mg/L): Pipette 5mL of primary source standard (1000mg/L) into a 500mL volumetric flask. Fill to the mark (QS) with reagent water and invert 3 times.

8.3.4. Med Cal Stock (100mg/L): Pipette 50mL of primary source standard (1000mg/L) into a 500mL volumetric flask. Fill to the mark (QS) with reagent water and invert 3 times.

8.3.5. Inorganic carbon spiking standard (480mg/L C): Prepare using 0.848g Na_2CO_3 (dried for 2hrs @ 250°C and cooled in a desiccator) into 200mL DI. 1mL of standard will be used for LCS. This is equivalent to 11.7mg/L C after spiking into 40mL VOA.

8.4. Calibration Standards

8.4.1. Calibration standards are generated by the auto-sampler. Calibration concentrations are programmed into a calibration template in the software and the instrument performs the appropriate dilutions from either the Low Cal or Med Cal Stock solutions.

8.5. Second Source Standards

8.5.1. ICV5 and ICV50 secondary source standards are diluted from 1000mg/L second source stock solution (ICV).

8.5.2. ICV5 – pipette 2.5mL of stock solution (ICV) into 500mL volumetric flask containing DI water.

8.5.3. ICV50 – pipette 25mL of stock solution (ICV) into 500mL volumetric flask containing DI water.

9. Sample Preservation and Handling

9.1. Field samples are taken in amber vials containing 1:1 HCl (H2SO4 can also be used). Samples should be acidified to a pH < 2 and stored at <6°C.

9.1.1. The pH of the samples is taken after analysis. All samples with a pH >2 will be H flagged and reported out of holding time.

9.2. Holding time for preserved samples is 28 days.

10. Quality Control

10.1. Initial Calibration as needed (Dictated when major maintenance occurs or through QC failures):

10.1.1. A five point low level and med level curve.

10.1.2. An ICV at 50mg/L.

10.1.3. An ICV at 5mg/L.

10.1.4. An ICB.

10.2. Continuing Calibration

10.2.1. An ICV5 (used to validate the low level curve), ICV50 (used to validate the med. Level curve), and ICB are run every 15 samples.

10.2.1.1. It is not necessary to analyze both ICVs if the analytes of interest are going to be found in only one curve range. (If the concentration of a known sample is going to be 5mg/L then only the ICV5 needs to be analyzed as a continuing standard).

10.3. Batch QC Requirements

10.3.1. A MB and LCS are analyzed every 20 samples.

10.3.2. A MS and MSD are analyzed every 10 samples.

10.3.3. A MB is prepared in a 40mL VOA that contains DI water only.

10.3.4. Prepare LCS sample in the following manner:

10.3.4.1. Measure 40mL DI and pour into a new VOA.

10.3.4.2. Using 250uL syringe, spike 200uL of the 1000mg/L primary source standard into the VOA. Spike concentration 4.85mg/L.

10.3.4.3. Be sure to submerge the needle in the DI water.

10.3.4.4. Using calibrated pipette, spike 1000uL of Inorganic Carbon Spiking Standard into the VOA. Fortifying the LCS with this spike shows that the inorganic carbon is being purged.

10.3.4.5. Cap and invert VOA 3 times.

10.3.4.6. Clean syringe 3 times with DI water.

10.3.5. Prepare MS/MSD samples in the following manner.

10.3.5.1. A single sample VOA, analyzed twice is used for both the MS/MSD.

10.3.5.2. If possible use a VOA without any headspace.

10.3.5.3. Prepare MS/MSD VOA without headspace in the following manner.

10.3.5.3.1. Carefully open VOA.

10.3.5.3.2. Using 250uL syringe, spike 200uL of the 1000mg/L primary source standard.

10.3.5.4. Be sure to submerge the syringe needle below the surface of the sample.

10.3.5.5. Spike concentration is 4.65mg/L.

10.3.5.6. Do not add inorganic carbon spiking standard to MS/MSD.

10.3.5.7. Cap and invert 3 times.

10.3.5.8. Clean syringe 3 times with DI water.

10.3.6. Prepare MS/MSD with headspace in the following manner.

10.3.6.1. Open sample VOA and measure 40mL of sample into a clean 50mL beaker.

10.3.6.2. Transfer the 40mL sample from the beaker into a clean labeled VOA vial.

10.3.6.3. Spike 200ul of the 1000mg/L primary source standard into the VOA vial.

10.3.6.4. Be sure to submerge the syringe needle below the surface of the sample.

10.3.6.5. The spike concentration will be 4.98mg/L.

10.3.6.6. Swirl to mix and transfer to a fresh 40mL VOA.

10.3.6.7. Cap and mix three times.

10.3.6.8. Clean syringe 3 times with DI water.

11. Calibration

11.1. Initial calibration

11.1.1. Initial calibration consists of a Low Cal Curve, Med Cal Curve, ICV5, ICV 50, and an ICB.

11.1.2. The calibration is based upon the peak area vs. the analyte concentration. The calibration equations are linear including the calibration blank response (zero point). See current HEAL Quality Assurance plan for details on the calibration equation.

11.1.3. At the start of every analytical run the ICV 5.0ppm and 50ppm are analyzed to verify the low and med calibration curve. They must pass within 10% of the expected concentration.

11.1.3.1. If either ICV fails to meet these criteria they may be reanalyzed one more time. If they pass continue running samples. If both ICVs fail a new calibration must be analyzed for both the low and medium curve. If only one ICV fails to confirm

the curve, then reanalyze the curve which corresponds to the failed ICV.

11.1.4. Each curve is based on a minimum of five levels.

11.1.5. The calibration concentrations in mg/L for the Low Cal Curve are as follows: 0, 0.5, 1.0, 5.0 and 10.0. The instrument dilutes down to these concentrations using the 10 mg/L Low Cal Curve stock.

11.1.6. The calibration concentrations in ppm for the Med Cal Curve are as follows: 0, 10, 25, 50 and 100. The instrument dilutes down to these concentrations using the 100mg/L Med Cal Curve stock.

11.1.7. Two second source standards are analyzed to validate both the Low and Med-level curves and an ICB to verify the cleanliness of the system.

11.1.7.1. ICV5 – 5mg/L TOC.

11.1.7.2. ICV50 – 50mg/L TOC.

11.1.7.3. ICB – Initial Calibration Blank

11.2. Continuing Calibration

11.2.1. An ICV5, ICV50, and an ICB must be run every 15 samples and at the end of the analysis.

11.2.1.1. It is not necessary to analyze both ICVs if the analytes of interest are going to be found in only one curve range. (If the concentration of a known sample is going to be 5mg/L then only the ICV5 needs to be analyzed as a continuing standard).

12. Sample Analysis

12.1. Check that the humidifier, drain container, rinse and dilution bottles are full with DI water, and HCl bottle adequately full of 2.0N HCl.

12.2. Prepare standards by filling sample vials with varying standard solutions.

12.3. Samples are received from the field in amber 40mL VOAs. These are placed directly on the instrument.

12.4. Power on instrument via the power button on lower right front of instrument. Allow the instrument about 45 minutes to heat the furnace and stabilize.

12.5. While the instrument stabilizes, open the TOC-Control V software, choose the sample table editor, and log in.

12.6. Connect the instrument to the computer by right clicking the TOCV icon in the left column of the

screen. Choose connect.

12.7. Open a new sample run and auto-generate your run for the day, making sure to include your current method, cal curves and QC. Fill in the appropriate columns with the sample name and vial position. The instrument can be programmed to shut down at the end of analysis.

12.8. The instrument must be reconnected via the yellow Icon before the run can be started.

12.9. Note: The auto sampler will not operate while the cover is off; and that removal of the cover during analysis will result in the halting of the analysis.

12.10. Arrange sample vials in the auto sampler tray according to their vial positions on the sample run list. The MB, LCS and samples should follow the primary ICV(5)/ICV(50)/ICB.

12.11. After the instrument has stabilized, start the run with the stoplight icon. Once started the analytical batch can not be altered or added to.

12.12. Be sure to save the batch file with the analysis date.

12.13. After successful analysis open sample vials and check pH using pH strip. Record pH on run log.

13. Calculations

13.1. The result is taken directly from the data sheet. If any of the samples are off-scale the auto-sampler auto-dilutes them and brings them on scale. The NPOC result from the data sheet is the final result unless the analyst manually diluted the sample. Then the sample result from the data sheet needs to be multiplied by the dilution made by the analyst to correctly calculate the total TOC result. See HEAL QAP for formula.

14. Method Performance/Measurement of Uncertainty

14.1. Method Performance

14.1.1. Proficiency testing is conducted bi-annually.

14.1.2. IDOCs (Initial Demonstration of Capability) are to be analyzed by all new analysts before handling client samples. ADOCPs (Annual Documentation of Continued Proficiency) are to be analyzed every year by all analysts working on the instrument. See HEAL QAP for more information.

14.2.Estimation of Measurement Uncertainty

14.2.1. Estimation of measurement uncertainty is to be conducted on an annual basis.

14.2.2. Collect the most recent 20 LCS results, calculate the standard deviation (s) and the 95% confidence interval (2 times the standard deviation). Report the uncertainty as ±2(s). See current HEAL QAP for details on uncertainty calculation.

14.2.3. See current HEAL QAP for details on calculating uncertainty.

14.3. MDLs must be empirically established annually. Refer to the HEAL QAP for more details.

14.3.1. MDLs must be established for both the low and med. level curves.

14.3.1.1. To accurately establish that the PQL of 10 based on the med. level curve can be achieved the analyst will reprocess the MDLs from the low level curve using the med. level curve. The calculated MDL from the med. level curve is not updated in LIMS and will not be reported to the client.

15. Pollution Prevention

15.1. This technique only requires a small amount of sample. Traditional wet chemistry methods for TOC require 500mL of sample. This method generally requires only 20mLs.

15.2. Less glass is required for the bottle kit.

16. Data Assessment and Acceptance Criteria for Quality Control Measures

16.1. The instrument is set up to take 4-5 replicate reading per sample. A fifth reading is only required if there is an outlier reading.

16.1.1. Initial Calibration

16.1.1.1. Both a low and mid-level five point calibration curve is analyzed. The r^2 (correlation coefficient) must be >/= 0.995

16.1.1.2. Both the ICV5 and ICV50 must agree within 10% of the expected value.

16.1.1.3. An ICB must be analyzed and quantitate to less than the PQL.

16.1.2. Continuing Calibration

16.1.2.1. An ICV5, ICV50, and ICB must be analyzed every 15 samples. The ICV5 and ICV50 must agree to within 10% of the expected value. The ICB must quantitate to less than the PQL.

16.1.3. Batch QC

16.1.3.1. An MB and LCS must be analyzed every 20 client samples.

16.1.3.2. An MS/MSD must be analyzed every 10 client samples.

16.1.3.3. The MB must quantitate to less than the PQL.

16.1.3.4. The LCS must agree to within 10% of the expected value.

16.1.3.5. The MS and MSD samples must agree to within 25% of the expected value. The RPD between the MS and MSD must be less than 20%.

16.1.4. Client Samples

16.1.4.1. Client samples must be preserved to a pH <2.

16.1.4.2. If the sample is ≥2 then the analyst must notate in LIMS that the sample is out of holding time by typing an "H" in the qualifier TAB of data entry.

17. Corrective Actions for Out-of-Control Data

17.1. See **Appendix A** for a summery of the Calibration, Continuing Calibration, and Batch QC requirements, acceptance criterion, and corrective actions.

17.2. Initial Calibration

17.2.1. If the r^2 for either of the curves is unacceptable, recalibrate or prepare new standards and recalibrate the curve that fails the r^2 criteria.

17.2.2. If the ICV5, ICV50, or ICB are unacceptable the standard may be re-prepped and analyzed one additional time. If the standard is still out-of-control conduct instrument maintenance and re-calibrate the instrument.

17.3. Continuing Calibration

17.3.1. If the ICV5, ICV50, or ICB is unacceptable, reanalyze affected samples back to the last passing continuing calibration (ICV5, ICV50, and ICB).

17.3.2. For example: If only the ICV50 was failing, then only the samples with hits calculated by the med. level calibration need to be reanalyzed. If the ICV5 fails then only the samples with hits calculated by the low level calibration need to be reanalyzed.

17.4. Batch QC

17.4.1. If the MB contains TOC above the PQL the MB may be analyzed one additional time. If the result is still greater than the PQL all samples in the analytical batch must be re-analyzed.

17.4.2. If the LCS recovery is unacceptable, it may be re-analyzed one additional time. If result is still unacceptable all samples in batch must be re-analyzed.

17.4.3. If the MS or MSD recovery is unacceptable do a 5x dilution of both the sample and the MS/MSD and reanalyze. Narrate all outliers.

17.4.4. If the RPD for MSD sample is unacceptable reanalyze one additional time. Narrate if unacceptable.

18. Contingencies for Handling Out-of-Control Data

18.1. All out-of-control data is flagged with the appropriate qualifier, and documented appropriately.

18.2. The client is notified about the out-of-control data.

19. Waste Management

19.1. Analytical results are used to characterize their respective sample contamination level(s) so that the proper disposal can be performed. These wastes will be disposed of according to their hazard as well as their type and level of contamination. Refer to the Hall Environmental Analysis Laboratory Chemical Hygiene Plan for details regarding waste disposal.

20. References

20.1. USEPA Method 9060 Total Organic Carbon, US EPA Revision 0, (1986).

20.2. Standard Methods for the Examination of Water and Wastewater, 20th Edition, Method 5310B (1999).

20.3. Shimadzu Total Organic Carbon Analyzer TOC-V CSH/CSN, User Manual (2001).

20.4. Quality Assurance Plan (QAP), HEAL.

20.5. Chemical Hygiene Plan, HEAL.

Appendix A

Total Organic Carbon Combustion Procedure Using Methods SM5310B/EPA9060
Calibration Specification And Corrective Action Procedures

Analytical Method	Parameter	QC Element	Frequency	Acceptance Criteria	Corrective Action
SM5310B	Wet Chemistry	Intial Calibration Low-Level (5 points + blank minimum)	Daily prior to analysis	Correlation coefficient r ≥ 0.995	• Recalibrate
		Intial Calibration Mid-Level (5 points + blank minimum)	Daily prior to analysis	Correlation coefficient r ≥ 0.995	• Recalibrate
		Intial Calibration Verifications (ICV5 and ICV50)	After low-level and mid-level calibration, prior to sample analysis	Results <10% from calibration standard	• Re-prep, recalibrate, and reanalyze
		Initial Calibration Blank (ICB)	After calibration, prior to sample analysis	< Practical quantitation limit (PQL)	• Clean system perform maintenance • Re-prep, recalibrate, and reanalyze
		Method Blank	1 per preperation/analysis batch (≤ 10 samples)	< PQL	• Reprep/reanalyze analytical batch as appropriate
		Laboratory Control Sample (LCS)	1 per preperation/analysis batch (≤ 10 samples)	% Recovery +/- 20% of actual value	• Reprep/reanalyze analytical batch as appropriate
		Matrix Spike (MS)	1 per preperation/analysis batch (≤ 10 samples)	% Recovery +/- 25% of actual value	• Dilute sample, MS/MSD 5x and reanalyze • Narrate all outliers
		Matrix Spike Duplicate (MSD)	1 per preperation/analysis batch (≤ 10 samples)	% Recovery +/- 25% of actual value Relative percent difference (RPD) <20%	• Dilute sample, MS/MSD 5x and reanalyze • Narrate all outliers
		Continuing Calibration Verification (ICV5 and ICV50)	Every 15 samples and the end of the analytical sequence	Results +/-10% from calibration standard	• Reanalyze affected samples back to the last acceptable ICV5/ICV50
		Continuing Calibration Blank (ICB)	Every 15 samples after both the ICV5 and ICV50 and at the end of the run	< Practical quantitation limit (PQL)	• Reanalyze all samples back to last acceptable ICB

Hall Environmental Analysis Laboratory

Common Name: SM5310B Total Organic Carbon (Combustion Infrared Method) EPA9060 Total Organic Carbon	S.O.P. Number: S-TOC-5
Method: Total Organic Carbon	
Effective Date: 3 June 2011	Supersedes: S-TOC-4
Laboratory Manager:	Quality Assurance/Quality Control Officer:

1. Scope and Application

   1.1. This S.O.P. covers the determination of the concentration of Non-purgeable Organic Carbon (NPOC) in ground water, surface and saline waters, drinking water, and domestic and industrial wastes.

2. Summary of Method

   2.1. A 5-ml sample aliquot is drawn out of a 40mL, thin septa amber vial, acidified by HCl or H_2SO_4 to a pH<2, and sparged for 1.5 minutes. A 50ul aliquot of the sample is then injected onto a high sensitivity oxidation catalyst inside of the combustion chamber at 680°C. The sample is burned in the combustion chamber to produce CO_2 and other combustion products which are then swept through a dehumidifier and halogen scrubber to a non-dispersive infrared (NDIR) gas analyzer, where the CO_2 is detected. The analog detection signal of the NDIR forms a peak, and the area of this peak is measured by a data processor.

3. Detection Limits

   3.1. MDLs shall be empirically established annually. MDL is subject to change. See LIMS for current MDL.

   3.2. The current PQL is 1.0mg/L

4. Definitions

   4.1. Calibration Blank (CalibBlank) – Blank Solution of DI water to "zero" the instrument.

   4.2. Initial Calibration Verification 5 mg/L (ICV5) – Second source standard @ 5 mg/L analyzed after the initial calibration of the Low-level curve. It is also analyzed after every tenth analysis to check the stability of the instrument during analysis.

   4.3. Initial Calibration Verification 50 mg/L (ICV50) – Second source standard @ 50 mg/L analyzed after the initial calibration of the Med-level curve. It is also analyzed after every tenth analysis to check the stability of the instrument during analysis.

   4.4. Initial Calibration Blank (ICB) – Analysis of Calibration blank solution after initial calibration to check

the cleanliness of blank. Should report below PQL.

4.5. Laboratory Control Spike (LCS) – Laboratory fortified blank.

4.6. Matrix Spike (MS) – Laboratory fortified sample.

4.7. Matrix Spike Duplicate (MSD) – Laboratory fortified sample duplicate.

4.8. Matrix Blank (MBLK) – Laboratory blank.

4.9. Method Detection Limit (MDL) – The constituent concentration that when processed through the complete method, produces a signal with a 99% probability that it is different than the blank.

4.10. Practical Quantitation Limit (PQL)/Reporting Limit – The minimum limit to which an analyte can be routinely reported.

5. Interference

5.1. Carbonate and bicarbonate carbon (inorganic carbon) represent interferences and are removed by acidification and purging of the sample. An LCS fortified with Na_2CO_3 is analyzed to show that the inorganic carbon is removed.

5.2. Method interference may be caused by contaminants in the reagent water, sample processing chemicals or analysis apparatus.

5.3. This procedure is applicable only to homogeneous samples which can be injected into the apparatus reproducibly by means of a syringe. The opening of this syringe limits the maximum size of particle which may be included in the sample. If necessary, allow the sample to settle, decant liquid into a fresh VOA. Label with sample ID and HEAL number. Retain decanted sample with original sample containers.

5.4. Samples that are not drawn into the syringe easily may require dilution into DI water. This is evidenced by a large variance of replicate combustion readings during sample analysis.

6. Safety

6.1 At minimum, latex gloves, a lab coat, and safety glasses must be worn to avoid any unnecessary contact with samples and standards. Refer to the MSDS before handling any standards for safety concerns.

6.2 When the analysts are done with standard vials they need to be disposed of in the appropriate manner. All standards, analytical vials, samples, and other laboratory-generated waste will be disposed in accordance with the HEAL Chemical Hygiene Plan.

6.3 Refer to the safety/warning section of the instrument manual before running or maintaining the instrument to avoid burns or electrical shock.

7. Apparatus, Materials

7.1. Shimadzu TOC-V – Total Organic Carbon Analyzer.

7.1.1. NDIR: non-dispersive infrared gas analyzer detector.

7.2. Operating Conditions

7.2.1. Furnace Temperature: 680°C

7.2.2. Carrier gas set to 150 psi.

7.2.3. Purge time set to 1.5 minutes

7.2.4. Acid addition to 1.5% of 2.0 N HCl

7.2.5. Injection volume set to 50uL.

7.2.6. Method programmed to accept best 4 of 5 injections.

7.2.7. Shimadzu combustion tube (part no. 638-41323) with platinum mesh.

7.2.8. Shimadzu high sensitivity catalyst (part no. 630-00996).

7.3. Shimadzu ASI-V Automated Sampler with 40mL VOA auto sampler rack.

7.3.1. The ASI-V Automated Sampler powers on and off through the instrument. It is important to note that the auto sampler will not operate while the cover is off; and that removal of the cover during analysis will result in the halting of the analysis.

7.4. Analytical balance capable of weighing to the nearest 0.0001g

7.5. pH strips that measure to the nearest 0.5 of a pH unit.

7.6. Sample vials (40mL) thin septa VOA vials.

7.7. Hand-held re-pipettors of various dispensing volumes

7.8. Volumetric flasks – 50mL, 100mL, 200mL, 250mL, and 500mL volumes.

7.9. Micro-syringes – 250mL and 500mL.

7.10. Variable volume 1mL handheld pipette.

7.11. Environmental Express Digestion Tubes – SC475.

8. Reagents and Standards

8.1. De-ionized water, free from organic carbons.

8.2. 2N HCl – Made from high purity OmniTrace Concentrated HCl. Add 41.25mL Conc. HCL to 100mL DI water in a 250mL volumetric flask. Bring to 250mL final volume with DI water. Reagent must be logged

into LIMS and recorded in TOC maintenance logbook.

8.3. Stock Standards

8.3.1. Primary source standard is a certified TOC standard purchased from Accustandard @ 1000mg/L.

8.3.2. Secondary source standard (ICV) is purchased from Ultra Scientific at a concentration of 1000mg/L.

8.3.3. Low Cal Stock (10mg/L): Pipette 5mL of primary source standard (1000mg/L) into a 500mL volumetric flask. Fill to the mark (QS) with reagent water and invert 3 times.

8.3.4. Med Cal Stock (100mg/L): Pipette 50mL of primary source standard (1000mg/L) into a 500mL volumetric flask. Fill to the mark (QS) with reagent water and invert 3 times.

8.3.5. Inorganic carbon spiking standard (480mg/L C): Prepare using 0.848g Na_2CO_3 (dried for 2hrs @ 250°C and cooled in a desiccator) into 200mL DI. 1mL of standard will be used for LCS. This is equivalent to 11.7mg/L C after spiking into 40mL VOA.

8.4. Calibration Standards

8.4.1. Calibration standards are generated by the auto-sampler. Calibration concentrations are programmed into a calibration template in the software and the instrument performs the appropriate dilutions from either the Low Cal or Med Cal Stock solutions.

8.5. Second Source Standards

8.5.1. ICV5 and ICV50 secondary source standards are diluted from 1000mg/L second source stock solution (ICV).

8.5.2. ICV5 – pipette 2.5mL of stock solution (ICV) into 500mL volumetric flask containing DI water.

8.5.3. ICV50 – pipette 25mL of stock solution (ICV) into 500mL volumetric flask containing DI water.

9. Sample Preservation and Handling

9.1. Field samples are taken in amber vials containing 1:1 HCl (H2SO4 can also be used). Samples should be acidified to a pH < 2 and stored at <6°C.

9.1.1. The pH of the samples is taken after analysis. All samples with a pH >2 will be H flagged and reported out of holding time.

9.2. Holding time for preserved samples is 28 days.

10. Quality Control

10.1. Initial Calibration as needed (Dictated when major maintenance occurs or through QC failures):

10.1.1. A five point low level and med level curve.

10.1.2. An ICV at 50mg/L.

10.1.3. An ICV at 5mg/L.

10.1.4. An ICB.

10.2. Continuing Calibration

10.2.1. An ICV5 (used to validate the low level curve), ICV50 (used to validate the med. Level curve), and ICB are run after every tenth analysis.

10.2.1.1. It is not necessary to analyze both ICVs if the analytes of interest are going to be found in only one curve range. (If the concentration of a known sample is going to be 5mg/L then only the ICV5 needs to be analyzed as a continuing standard).

10.3. Batch QC Requirements

10.3.1. A MB and LCS are analyzed every 20 samples.

10.3.2. A MS and MSD are analyzed every 10 samples.

10.3.3. A MB is prepared in a 40mL VOA that contains DI water only.

10.3.4. Prepare LCS sample in the following manner:

10.3.4.1. Measure 40mL DI and pour into a new VOA.

10.3.4.2. Using 250uL syringe, spike 200uL of the 1000mg/L primary source standard into the VOA. Spike concentration 4.85mg/L.

10.3.4.3. Be sure to submerge the needle in the DI water.

10.3.4.4. Using calibrated pipette, spike 1000uL of Inorganic Carbon Spiking Standard into the VOA. Fortifying the LCS with this spike shows that the inorganic carbon is being purged.

10.3.4.5. Cap and invert VOA 3 times.

10.3.4.6. Clean syringe 3 times with DI water.

10.3.5. Prepare MS/MSD samples in the following manner.

10.3.5.1. A single sample VOA, analyzed twice is used for both the MS/MSD.

10.3.5.2. If possible use a VOA without any headspace.

10.3.5.3. Prepare MS/MSD VOA without headspace in the following manner.

10.3.5.3.1. Carefully open VOA.

10.3.5.3.2. Using 250uL syringe, spike 200uL of the 1000mg/L primary source standard.

10.3.5.4. Be sure to submerge the syringe needle below the surface of the sample.

10.3.5.5. Spike concentration is 4.65mg/L.

10.3.5.6. Do not add inorganic carbon spiking standard to MS/MSD.

10.3.5.7. Cap and invert 3 times.

10.3.5.8. Clean syringe 3 times with DI water.

10.3.6. Prepare MS/MSD with headspace in the following manner.

10.3.6.1. Open sample VOA and measure 40mL of sample into a clean 50mL beaker.

10.3.6.2. Transfer the 40mL sample from the beaker into a clean labeled VOA vial.

10.3.6.3. Spike 200ul of the 1000mg/L primary source standard into the VOA vial.

10.3.6.4. Be sure to submerge the syringe needle below the surface of the sample.

10.3.6.5. The spike concentration will be 4.98mg/L.

10.3.6.6. Swirl to mix and transfer to a fresh 40mL VOA.

10.3.6.7. Cap and mix three times.

10.3.6.8. Clean syringe 3 times with DI water.

11. Calibration

11.1. Initial calibration

11.1.1. Initial calibration consists of a Low Cal Curve, Med Cal Curve, ICV5, ICV 50, and an ICB.

11.1.2. The calibration is based upon the peak area vs. the analyte concentration. The calibration equations are linear including the calibration blank response (zero point). See current HEAL Quality Assurance plan for details on the calibration equation.

11.1.3. At the start of every analytical run the ICV 5.0ppm and 50ppm are analyzed to verify the low and med calibration curve. They must pass within 10% of the expected concentration.

11.1.3.1. If either ICV fails to meet these criteria they may be reanalyzed one more time. If they pass continue running samples. If both ICVs fail a new calibration must be analyzed for both the low and medium curve. If only one ICV fails to confirm

the curve, then reanalyze the curve which corresponds to the failed ICV.

11.1.4. Each curve is based on a minimum of five levels.

11.1.5. The calibration concentrations in mg/L for the Low Cal Curve are as follows: 0, 0.5, 1.0, 5.0 and 10.0. The instrument dilutes down to these concentrations using the 10 mg/L Low Cal Curve stock.

11.1.6. The calibration concentrations in ppm for the Med Cal Curve are as follows: 0, 10, 25, 50 and 100. The instrument dilutes down to these concentrations using the 100mg/L Med Cal Curve stock.

11.1.7. Two second source standards are analyzed to validate both the Low and Med-level curves and an ICB to verify the cleanliness of the system.

11.1.7.1. ICV5 – 5mg/L TOC.

11.1.7.2. ICV50 – 50mg/L TOC.

11.1.7.3. ICB – Initial Calibration Blank

11.2. Continuing Calibration

11.2.1. An ICV5, ICV50, and an ICB must be run after every tenth analysis and at the end of the analysis.

11.2.1.1. It is not necessary to analyze both ICVs if the analytes of interest are going to be found in only one curve range. (If the concentration of a known sample is going to be 5mg/L then only the ICV5 needs to be analyzed as a continuing standard).

12. Sample Analysis

12.1. Check that the humidifier, drain container, rinse and dilution bottles are full with DI water, and HCl bottle adequately full of 2.0N HCl.

12.2. Prepare standards by filling sample vials with varying standard solutions.

12.3. Samples are received from the field in amber 40mL VOAs. These are placed directly on the instrument.

12.4. Power on instrument via the power button on lower right front of instrument. Allow the instrument about 45 minutes to heat the furnace and stabilize.

12.5. While the instrument stabilizes, open the TOC-Control V software, choose the sample table editor, and log in.

12.6. Connect the instrument to the computer by right clicking the TOCV icon in the left column of the

screen. Choose connect.

12.7. Open a new sample run and auto-generate your run for the day, making sure to include your current method, cal curves and QC. Fill in the appropriate columns with the sample name and vial position. The instrument can be programmed to shut down at the end of analysis.

12.8. The instrument must be reconnected via the yellow Icon before the run can be started.

12.9. Note: The auto sampler will not operate while the cover is off; and that removal of the cover during analysis will result in the halting of the analysis.

12.10. Arrange sample vials in the auto sampler tray according to their vial positions on the sample run list. The MB, LCS and samples should follow the primary ICV(5)/ICV(50)/ICB.

12.11. After the instrument has stabilized, start the run with the stoplight icon. Once started the analytical batch can not be altered or added to.

12.12. Be sure to save the batch file with the analysis date.

12.13. After successful analysis open sample vials and check pH using pH strip. Record pH on run log.

13. Calculations

13.1. The result is taken directly from the data sheet. If any of the samples are off-scale the auto-sampler auto-dilutes them and brings them on scale. The NPOC result from the data sheet is the final result unless the analyst manually diluted the sample. Then the sample result from the data sheet needs to be multiplied by the dilution made by the analyst to correctly calculate the total TOC result. See HEAL QAP for formula.

14. Method Performance/Measurement of Uncertainty

14.1. Method Performance

14.1.1. Proficiency testing is conducted bi-annually.

14.1.2. IDOCs (Initial Demonstration of Capability) are to be analyzed by all new analysts before handling client samples. ADOCPs (Annual Documentation of Continued Proficiency) are to be analyzed every year by all analysts working on the instrument. See HEAL QAP for more information.

14.2.Estimation of Measurement Uncertainty

14.2.1. Estimation of measurement uncertainty is to be conducted on an annual basis.

14.2.2. Collect the most recent 20 LCS results, calculate the standard deviation (s) and the 95% confidence interval (2 times the standard deviation). Report the uncertainty as ±2(s). See current HEAL QAP for details on uncertainty calculation.

14.2.3. See current HEAL QAP for details on calculating uncertainty.

14.3. MDLs must be empirically established annually. Refer to the HEAL QAP for more details.

14.3.1. MDLs must be established for both the low and med. level curves.

14.3.1.1. To accurately establish that the PQL of 10 based on the med. level curve can be achieved the analyst will reprocess the MDLs from the low level curve using the med. level curve. The calculated MDL from the med. level curve is not updated in LIMS and will not be reported to the client.

15. Pollution Prevention

15.1. This technique only requires a small amount of sample. Traditional wet chemistry methods for TOC require 500mL of sample. This method generally requires only 20mLs.

15.2. Less glass is required for the bottle kit.

16. Data Assessment and Acceptance Criteria for Quality Control Measures

16.1. The instrument is set up to take 4-5 replicate reading per sample. A fifth reading is only required if there is an outlier reading.

16.1.1. In the NPOC tab of the instrument method the criteria for replicate acceptability is set to a SD max of 0.1 and CV Max of 2.0%. Any reading failing to meet these requirements is flagged as an outlier and excluded by the instrument software.

16.1.2. Initial Calibration

16.1.2.1. Both a low and mid-level five `point calibration curve` is analyzed. The r^2 (correlation coefficient) must be >/= 0.995

16.1.2.2. Both the ICV5 and ICV50 must agree within 10% of the expected value.

16.1.2.3. An ICB must be analyzed and quantitate to less than the PQL. The ICB must not have a negative value reported less than negative the PQL.

16.1.3. Continuing Calibration

16.1.3.1. An ICV5, ICV50, and ICB must be analyzed after every tenth analysis. The ICV5 and ICV50 must agree to within 10% of the expected value. The ICB must quantitate to less than the PQL.

16.1.4. Batch QC

16.1.4.1. An MB and LCS must be analyzed every 20 client samples.

16.1.4.2. An MS/MSD must be analyzed every 10 client samples.

16.1.4.3. The MB must quantitate to less than the PQL. The MB must not have a negative value reported less than negative the PQL.

16.1.4.4. The LCS must agree to within 10% of the expected value.

16.1.4.5. The MS and MSD samples must agree to within 25% of the expected value. The RPD between the MS and MSD must be less than 20%.

16.1.5. Client Samples

16.1.5.1. Client samples must be preserved to a pH <2.

16.1.5.2. If the sample is ≥2 then the analyst must notate in LIMS that the sample is out of holding time by typing an "H" in the qualifier TAB of data entry.

17. Corrective Actions for Out-of-Control Data

17.1. See **Appendix A** for a summery of the Calibration, Continuing Calibration, and Batch QC requirements, acceptance criterion, and corrective actions.

17.2. Initial Calibration

17.2.1. If the r^2 for either of the curves is unacceptable, recalibrate or prepare new standards and recalibrate the curve that fails the r^2 criteria.

17.2.2. If the ICV5, ICV50, or ICB are unacceptable the standard may be re-prepped and analyzed one additional time. If the standard is still out-of-control conduct instrument maintenance and re-calibrate the instrument.

17.3. Continuing Calibration

17.3.1. If the ICV5, ICV50, or ICB is unacceptable, reanalyze affected samples back to the last passing continuing calibration (ICV5, ICV50, and ICB).

17.3.2. For example: If only the ICV50 was failing, then only the samples with hits calculated by the med. level calibration need to be reanalyzed. If the ICV5 fails then only the samples with hits calculated by the low level calibration need to be reanalyzed.

17.4. Batch QC

17.4.1. If the MB contains TOC above the PQL the MB may be analyzed one additional time. If the result is still greater than the PQL all samples in the analytical batch must be re-analyzed.

17.4.2. If the LCS recovery is unacceptable, it may be re-analyzed one additional time. If result is still unacceptable all samples in batch must be re-analyzed.

17.4.3. If the MS or MSD recovery is unacceptable do a 5x dilution of both the sample and the MS/MSD and reanalyze. Narrate all outliers.

17.4.4. If the RPD for MSD sample is unacceptable reanalyze one additional time. Narrate if unacceptable.

18. Contingencies for Handling Out-of-Control Data

18.1. All out-of-control data is flagged with the appropriate qualifier, and documented appropriately.

18.2. The client is notified about the out-of-control data.

19. Waste Management

19.1. Analytical results are used to characterize their respective sample contamination level(s) so that the proper disposal can be performed. These wastes will be disposed of according to their hazard as well as their type and level of contamination. Refer to the Hall Environmental Analysis Laboratory Chemical Hygiene Plan for details regarding waste disposal.

20. References

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20.3. Shimadzu Total Organic Carbon Analyzer TOC-V CSH/CSN, User Manual (2001).

20.4. Quality Assurance Plan (QAP), HEAL.

20.5. Chemical Hygiene Plan, HEAL.

Appendix A

Total Organic Carbon Combustion Procedure Using Methods SM5310B/EPA9060
Calibration Specification And Corrective Action Procedures

Analytical Method	Parameter	QC Element	Frequency	Acceptance Criteria	Corrective Action
SM5310B	Wet Chemistry	Intial Calibration Low-Level (5 points + blank minimum)	When ICVs Fail	Correlation coefficient r ≥ 0.995	• Recalibrate
		Intial Calibration Mid-Level (5 points + blank minimum)	When ICVs Fail	Correlation coefficient r ≥ 0.995	• Recalibrate
		Intial Calibration Verifications (ICV5 and ICV50)	After low-level and mid-level calibration, prior to sample analysis	Results <10% from calibration standard	• Re-prep, recalibrate, and reanalyze
		Initial Calibration Blank (ICB)	After calibration, prior to sample analysis	< Practical quantitation limit (PQL); The ICB must not have a negative value reported less than negative the PQL.	• Clean system perfrom maintenance • Re-prep, recalibrate, and reanalyze
		Method Blank	1 per preperation/analysis batch (≤ 20 samples)	< PQL; The MB must not have a negative value reported less than negative the PQL.	• Reprep/reanalyze analytical batch as appropriate
		Laboratory Control Sample (LCS)	1 per preperation/analysis batch (≤ 20 samples)	% Recovery +/- 20% of actual value	• Reprep/reanalyze analytical batch as appropriate
		Matrix Spike (MS)	1 per preperation/analysis batch (≤ 10 samples)	% Recovery +/- 25% of actual value	• Dilute sample, MS/MSD 5x and reanalyze • Narrate all outliers
		Matrix Spike Duplicate (MSD)	1 per preperation/analysis batch (≤ 10 samples)	% Recovery +/- 25% of actual value Relative percent difference (RPD) <20%	• Dilute sample, MS/MSD 5x and reanalyze • Narrate all outliers
		Continuing Calibration Verification (ICV5 and ICV50)	Every 10 injections and the end of the analytical sequence	Results +/-10% from calibration standard	• Reanalyze affected samples back to the last acceptable ICV5/ICV50
		Continuing Calibration Blank (ICB)	Every 10 injections after both the ICV5 and ICV50 and at the end of the run	< Practical quantitation limit (PQL); The ICB must not have a negative value reported less than negative the PQL.	• Reanalyze all samples back to last acceptable ICB

Hall Environmental Analysis Laboratory, Inc.

Corrective Action Report

Date Initiated: 10-May-11 **Corrective Action Report ID:** 3533

Initiated By: Sarah F. Edwards **Department:** ad

Corrective Action Description

CAR Summary: As a result of the 2011 ORELAP Audit: analysts are now required to link calibration information to each run.

Description of Nonconformance: Quality Systems; 2011 ORELAP Audit Finding: Although the laboratory does keep their calibration data in the run that is calibrated, the individual runs only reference where the calibration records are kept and do not include pertinent information for accurately analyzing the data. Documentation of the relevant calibration, such as a checklist, or the calibration itself must be included in data packets in methods that do not calibrate for every run.

Description of Corrective Action: All staff has been retrained to link the calibration information to each run. Each Technical Director or Section Supervisor will select the most appropriate way to do this. A training document has been read and signed by all analysts and a copy is attached. Calibration summaries have been created for 8260, 8270, 9060/SM5310B, 504.1/8011, and 8310. Copies of these summaries will be attached to this CAR and scanned into the current SOPs. Effective 6/6/2011, these sheets will be generated and reviewed after each calibration event and analyst will place a copy of the approved sheet in the data folder with each subsequent batch of data. The sheets show a summary of Pass/Fail for the ICV and whether or not there were any Calibration Points excluded. There is also a section for additional comments to be made that may be pertinent to the calibration. At the next due revisions the calibration summaries will be added to the SOP appendices and discussed in the text. The 8260 SOP will be updated on or before 6/29/2011, the 8270 on or before 11/01/2011, the SM5310B/9060 on or before 6/3/2012, the 504.1/8011 on or before 11/17/2011, and the 8310 on or before 12/8/2011.

Performed By: Andy Freeman **Completion Date:** 03-Jun-11

Client Notification

Client Notification Required: No **Notified By:**

Comment:

Quality Assurance Review

Approval and Closure

Technical Director / Deputy Tech. Dir.: Andy **Close Date:** 6/6

QA Officer Approval: Carolyn Swanson **QA Date:** 06-Jun-11

Hall Environmental Analysis Laboratory, Inc.

Corrective Action Report

Date Initiated: 10-May-11 **Corrective Action Report ID:** 3533

Initiated By: Sarah F. Edwards **Department:** ad

Nonconformance Type: Deficiency

Further Action required by QA:

Approval and Closure

Technical Director / Deputy Tech. Dir.: _____ Close Date:

QA Officer Approval: _____ QA Date: 06-Jun-11

Carolyn Swanson

Last Updated BY sarah Updated: 06-Jun-2011 11:32 AM Reported: 06-Jun-2011 11:33 A

9060/SM5310B
Calibration Summary

Instrument: _____ **Date:** _____

Method: 9060/SM5310B

Calibration Name: _____

Analyte	ICV Pass	ICV Fail	Calibration Curve Points Removed Yes	Calibration Curve Points Removed No	
Total Organic Carbon(LOW)	☐	☐	☐	☐	_____
Total Organic Carbon(MED)	☐	☐	☐	☐	_____

Additional Notes:

_____

Analyst: _____

QA: _____

Hall Environmental
Analysis Laboratory

Common Name: SM 4500-H+	S.O.P. Number: S-SM4500-H+-6
Method: The pH of Aqueous Samples and Aqueous Extracts of Soil/Waste Samples	
Effective Date: August 29, 2011	Supersedes: S-SM4500-H+-5
Laboratory Manager:	Quality Assurance/Quality Control Officer:

1. Scope and Application

   1.1. This Standard Operating Procedure (SOP) is applicable to the analysis of pH for aqueous samples, including domestic, surface, and saline water, and acid rain. It is also applicable to aqueous extracts of soil and waste samples.

2. Summary of Method

   2.1. pH of a sample is measured electrometrically by use of a glass electrode. Aqueous samples are analyzed directly. Soil and waste samples are analyzed by immersing the pH probe directly into a 1:1 (DI water: sample) sample slurry.

      2.1.1. The process for pH determination using the automated analysis is discussed herein as a single entity. However, analysis can be done conjointly with eC and alkalinity determination without any procedural deviation based on this SOP.

      2.1.2. Samples for pH determination can also be analyzed using the SympHony SB70P. All soils should be analyzed using this instrument.

      2.1.3. Appendix A contains a method flow chart and method parameters for Poseidon.

3. Detection Limits

   3.1. The pH should be reported to within 0.01 pH units for all samples.

4. Definitions

4.1. Analytical batch – a group of samples not to exceed 20 plus a sample duplicate that are analyzed consecutively.

4.2. DI Water (DI) – De-ionized water/reagent-free water. Water that is passed through a purification system and is free from interferences.

4.3. Continuing Calibration Verification (CCV) – A standard analyzed from the same source as the calibration to check instrument stability.

4.4. Initial Calibration Verification (ICV) – A pH buffer from a different manufacturer than those standards used for calibration. Used as a Second Source verification.

4.5. Sample Duplicate (DUP) – Identical split of a sample that is analyzed to determine reproducibility of a method in a given matrix.

5. Interferences

5.1. Oily or particulate matter may impair electrode response and can be eliminated by mild detergent rinsing followed by rinsing with copious amounts of DI.

5.2. Suspended solids may interfere with the electrode responsiveness and should be allowed to settle if time permits. Suspended solids in soil sample extracts can be remedied by centrifuging the sample.

5.3. Temperature fluctuations will increase stabilization time. Make certain to allow the electrode to stabilize. Samples shall be allowed to equilibrate to room temperature prior to analysis.

5.4. Magnetic stir bars should not be used for the analysis of acid rain samples. The sample should be gently swirled.

5.5. Samples with very low or very high pH may give incorrect readings. Samples with a pH>10 may give a result that is biased low. Samples with a pH<1, may give a result that is biased high.

6. Safety

6.1. For handling procedures and exposure information always refer to the MSDS. To keep exposure to reagents and standards to a minimum, analysts must wear appropriate personal protective equipment (PPE) when working in the laboratory. Gloves, lab coats, and safety glasses must be worn when handling samples, standards, or reagents.

7. Apparatus and Materials

7.1. Poseidon - 855 Robotic Titrosampler equipped with a pH glass electrode using 3mol/L KCL as electrolyte.

7.1.1. Temperature is determined using the combination electrode.

7.1.2. Electrode storage solution – 3M KCL.

7.1.3. Electrode filling solutions - use 3M KCl. Do not use storage solution as filling solution.

7.1.4. 60mL disposable plastic cups for *Titrosampler.*

7.1.5. DI dispenser – squirt bottle

7.2. Memphis - VWR SympHony pH/Temp Meter with a SympHony Ag/AgCl pH Gel Electrode and a separate ATC temperature probe.

7.2.1. Electrode filling solutions - SympHony Ag/AgCl pH Gel Electrode use VWR Cat. no 14002-824.

7.2.2. Electrode storage solution – SympHony Storage Solution – VWR Cat. No. 14002-828.

7.2.3. Magnetic stirrer and Teflon stir bars.

7.2.4. Plastic 4oz disposable solo cups.

7.2.5. Plastic squirt bottle w/ DI water.

7.2.6. Spatulas.

7.2.7. Kim-wipes.

8. Reagents and Standards

8.1. Certified calibration pH buffer solutions: 1.68, 4.00, 7.00, 10.00, and 12.45.

8.2. Certified second source (ICV) pH buffer solution: 7.00.

9. Sample Collection, Preservation, and Storage

9.1. Aqueous samples are to be collected in unpreserved plastic containers. Aqueous samples are stored at above freezing to 6°C until analysis. All Drinking Water complicate samples have a holding time of 15 minutes from the time of collection. All non-drinking water compliant samples have a holding time of 24 hours.

9.2. Soil/waste samples are to be collected in glass jars, plastic containers, or Ziploc bags. Soil samples do not require refrigeration.

9.3. Soil/Waste samples must be analyzed as soon as possible. Current HEAL hold time is 180 days. Soil extracts are analyzed immediately after preparation with DI water.

10. Quality Control

10.1. Initial Calibration

10.1.1. Poseidon

10.1.1.1. The pH meter is calibrated at least daily prior to sample analysis or more frequently as needed due to interferences or changes in ambient conditions.

10.1.1.2. Calibration consists of pH buffers 1.68, 4.00, 7.00, 10.00, 12.45.

10.1.1.3. A certified ICV pH buffer of 7.00 is used to verify calibration.

10.1.1.3.1. The calibration slope must be 96-102%.

10.1.1.3.2. The ICV must be within ±0.1pH units.

10.1.2. Memphis

10.1.2.1. The pH meter is calibrated at least once daily prior to sample analysis or more frequently if needed due to interferences or changes in ambient conditions.

10.1.2.2. Routine calibration consists of pH buffers 4.00, 7.00, and 10.00. All samples reported from the routine calibration must have pH values between pH buffers 4 and 10. Otherwise a separate non-routine calibration must be performed.

10.1.2.3. Non-routine low pH calibration is required when sample pH values fall below the pH 4 buffer. Low pH calibration consists of pH buffers 1.68, 4.00, and 7.00.

10.1.2.4. Non-routine high pH calibration is required when sample pH values fall above the pH 10 buffer. High pH calibration consists of pH buffers 7, 10, and 12.45.

10.1.2.5. A certified ICV pH buffer 7.00 is used to check meter calibration.

10.1.2.5.1. The calibration slope must be 92-102%.

10.1.2.5.2. The ICV must be within ±0.1pH units.

10.2. Continuing Calibration

10.2.1. A CCV is analyzed at a minimum of one per analytical batch.

10.2.1.1. The CCV must be within ±0.1pH units of its known value to be acceptable.

10.2.1.2. For Poseidon, CCV analysis rotates between the 4.00, 7.00, and 10.00 buffers.

10.2.1.3. For Memphis, CCV analysis rotates between the 4.00, 7.00, and 10.00 buffers.

10.2.1.3.1. For non-routine low calibrations using Memphis, rotate between 1.68, 4.00, and 7.00 buffers. For non-routine high calibrations using the Memphis, rotate between 7.00, 10.00, and 12.45 buffers.

10.2.2. The CCV analysis should be more frequent when analyzing samples that are viscous. A CCV should be analyzed following any samples which are suspected to cause interference with the pH electrode.

10.2.3. If the pH meter has previously been calibrated on the day of use, an ICV is used to check calibration before sample analysis begins. The ICV must pass acceptance criteria prior to sample analysis

10.2.3.1. The ICV must be within ±0.1pH units to be acceptable.

10.3. Batch QC requirements

10.3.1. A sample duplicate must be analyzed every 20 samples.

10.3.1.1. The value of the DUP must be within ±0.1pH units of the sample

11. Calibration Procedures

11.1. Poseidon

11.1.1. Empty the waste container into the sink if container is more than half full. Dilute waste with copious amounts of tap water.

11.1.2. Fill DI reagent container with DI water.

11.1.3. Remove pH probe for storage container and rinse with DI water.

11.1.4. Remove plug in electrode fill hole and add 3mol/L KCL filling solution.

11.1.5. Place pH probe into glass beaker on instrument.

11.1.6. Log into the *Tiamo* software.

11.1.7. Select Workplace icon.

11.1.7.1. Select the *Determination Series* tab.

11.1.7.2. Click the *Sample Tab* and select *New*.

11.1.7.2.1. This will reset the table (sequence list).

11.1.7.3. Double click on the first line and choose the *Prep System* method from the scroll down menu. Click *Apply*, and then *Close*.

11.1.7.4. Double click on the next line and select **pH Calibration** method from the scroll down menu.

11.1.7.4.1. On ID1 line, enter the unique standard identification numbers of the standards being used for the calibration. The system is setup to sample specific positions on the auto-sampler rack. The calibration will be sampled starting with the lowest buffer, 1.68, and finish with the highest buffer, 12.45.

11.1.7.5. Double click on the next line and select pH only method from the scroll down menu.

11.1.7.5.1. Type the sample position number for the location of the sample.

11.1.7.5.2. On ID1 line, enter *DI H_2O*. This will serve as a blank to further clean the system after analysis of the 12.45 buffer.

11.1.7.6. Double click on the next line and select the *pH only* method to program the ICV. The pH ICV must be run by the pH only method and never by a titration method.

11.1.7.6.1. Type the sample position number for the location of the sample.

11.1.7.6.2. On ID1 line, enter the unique standard identification number and 7.00pH ICV.

11.1.7.6.3. Click *Apply*, and then *Close*.

11.1.7.7. Place pH buffers used for calibration in the appropriate, labeled positions on the auto-sampler rack.

11.1.7.8. Place DI H_2O and 7.00 ICV in correct positions on auto-sampler rack.

11.1.7.9. Click the *Start* button.

11.2. Memphis

11.2.1. Check the condition of the probe. Fill probe with the appropriate filling solution if necessary. Check to be sure there is not excessive crystal build up on the inside of the probe (clean if necessary). Record any maintenance in the maintenance logbook.

11.2.2. Check buffer solutions. Periodically change all buffer solutions (approximately every 4 weeks). Or change if they show any signs of degradation.

11.2.3. Check the calibration logbook for the Beckman to see if the meter has been calibrated on the day of use. Calibrate if necessary. If calibration has been performed earlier during the day, it is possible to skip meter calibration by analyzing the ICV.

11.2.4. If necessary turn on the pH Meter and allow it to warm up.

11.2.5. Using the pH Meter Daily Calibration Log record the date of calibration, meter name, and ID numbers of the calibration and second source buffers.

11.2.6. Remove the electrode from the storage solution and rinse thoroughly with DI-water. Gently shake, using a vertical motion to remove any excess water.

11.2.7. Follow instructions in the instrument manual on page 27 for appropriate steps to be used in pH electrode calibration.

11.2.8. After calibration, the slope will be displayed. Record slope in the Daily Calibration Log.

11.2.8.1. The slope must be 92-102% to meet acceptance criteria.

11.2.9. Analyze the ICV and record the pH of the solution in the Daily Calibration Log.

11.2.9.1. The resultant value must be 7.00 ±0.1pH units.

12. Sample Analysis

12.1. If necessary calibrate the pH meter according to the outlined procedures dependant on instrument of choice. Otherwise continue analysis by analyzing an ICV.

12.2. Aqueous Samples

12.2.1. Poseidon

12.2.1.1. Remove samples from refrigerator and set them out on the counter for at least two hours before analysis.

12.2.1.2. Obtain clean 60mL auto-sampler cups.

12.2.1.3. Select the Workplace icon. Click on the *Determination Series* tab.

12.2.1.3.1. Double click on the first line and choose the *pH ONLY* method from the scroll down menu.

12.2.1.3.2. Enter the HEAL sample ID on ID1 line.

12.2.1.3.3. Enter additional notes for sample on ID2 line.

12.2.1.3.4. Enter the sample position number.

12.2.1.3.5. Click the right scroll arrow on the bottom left of the window to advance to the next sample position.

12.2.1.3.5.1. Repeat 12.2.1.3.1 to load additional samples into list.

12.2.1.3.6. Click *Apply*, and then *Close* when finished entering IDs for all desired analyses.

12.2.1.4. Sample Reports will print after each determination is complete.

12.2.1.5. After sample list has been filled appropriately, and prior to analysis, the list should be printed. The list shall be used to ensure that samples are located in appropriate positions on the auto-sampler rack.

12.2.1.5.1. Click on *Sample table* and select *Print (PDF)*. This will generate a temporary PDF file.

12.2.1.5.2. Print the PDF file. Save the PDF file to the Sample list directory located on the desktop with the file name corresponding to the analysis date.

12.2.1.6. Once sample list is complete and has been printed, prepare and load samples according to the sample list. Using this sequence should reduce analyst error of loading the auto-sampler incorrectly.

12.2.1.7. Pour sample into beaker to ¾ volume. Place samples in the auto-sampler tray in the appropriate sample positions.

12.2.1.7.1. Avoid floating particulates as they can cause interferences with the electrode.

12.2.1.7.2. Never load samples that have large amounts of suspended solids onto Poseidon. This could lead to fouled plumbing and extensive repairs.

12.2.1.8. Remember to load and analyze a sample duplicate for up to every twenty samples.

12.2.1.9. Click *Start* to begin analysis.

12.2.1.10. A passing CCV must also be analyzed at the end of each analytical batch.

12.2.1.10.1. Resultant value must be ±0.1pH units of expected value to be acceptable.

12.2.1.11. The auto-sampler rinses between each sample.

12.2.1.12. The aspiration line and conductivity probe on the robotic arm are rinsed three times between sample analyses.

12.2.1.13. When the analysis of aqueous samples that have less than 20mL of volume, use the instrument method, *pH manual*. Any analyst using this method must be trained on the use of this method and refer to the appropriate training document.

12.2.1.13.1. The samples, when analyzed using *pH manual*, shall be analyzed individually, not using a sequence table. This allows time for analyst to thoroughly clean the electrode prior to the following sample. No sample is aspirated into the titration vessel. The electrode is attached to a "stand" and used similarly to Memphis.

12.2.2. Memphis

12.2.2.1. Pour enough of the sample into a disposable Solo cup to fully immerse the tip of the electrode (approximately 50mL).

12.2.2.2. Add a Teflon stir bar and place sample on stirrer. Gently stir samples.

12.2.2.3. Do not use a stir bar for acid rain samples. Gently swirl the sample.

12.2.2.4. Immerse probe and allow the meter to stabilize and record the pH and temperature. The glass bulb on the probe must be completely immersed in the sample (Memphis has an external thermocouple, be sure it is immersed in the sample as well).

12.2.2.5. Rinse with DI and blot the electrode with a Kim-wipe between samples.

12.3. Soil/Waste Samples

12.3.1. Soils should not be analyzed on Poseidon when other options are viable.

12.3.1.1. When it is necessary to analyze soils using Poseidon, follow the appropriate training document. Use the appropriate instrument method, *pH manual*. This method has been created to analyze soil pH and aqueous matrices where sample volume is less than 20mL. Only analysts that have received training using this instrument method can analyze soil pH on Poseidon.

12.3.2. Place 25g of soil sample and 25mL DI water into a disposable Solo cup.

12.3.3. Add a Teflon stir bar and place on stirrer. Gently stir samples for 5 minutes

12.3.4. Allow any suspended soil or waste to settle for 1 hour if time permits or centrifuge if necessary.

12.3.5. Immerse probe, making certain it is only in the aqueous phase, and allow the meter to stabilize and record the pH and temperature to the nearest 0.01. The glass bulb on the probe must be completely immersed in the sample (Mephis has an external thermocouple, be sure it is immersed in the sample as well).

12.3.6. Rinse with DI-water. Gently shake, using a vertical motion to remove excess water.

13. Method Performance

13.1. Method Performance

13.1.1. PT or "blind study" aqueous samples are analyzed semi-annually to check the accuracy of the pH meters.

13.2. Estimation of Uncertainty

13.2.1. Estimation of Uncertainty is to be performed on an annual basis.

13.2.2. Collect at least 20 of the most recent LCS recoveries. Calculate the average percent recovery and standard deviation *s*. Calculate 95% confidence interval (2 times the standard deviation). Report the estimated uncertainty as +/- 2*s*.

13.2.3. Calculate standard deviation (s) and 95% confidence interval according to the formulas outlined in the HEAL QAP.

13.3. Demonstration of Capability (DOC)

13.3.1. After every new analyst has been trained, and before he/she is able to report pH results, an acceptable DOC must be completed.

13.3.2. DOC for pH consists of four replicate analyses of a pH buffer; 4, 7 or 10. Each buffer must fall between ±0.1 pH units of the true value to be deemed acceptable.

13.3.3. Annual Documentation of Continued Proficiency (ADOCP) is performed annually after acceptable DOC has been established. See HEAL Quality Assurance Plan for the appropriate procedure and calculations

13.3.4. IDOCs are required for the two procedures.

14. Pollution Prevention

14.1. Small amounts of standards are used during calibration of Memphis. Standards are used until degradation is observed or they reach their designated expiration (approximately 4 weeks).

14.2. Buffers with longer expiration dates are ordered in bulk. Buffers with short expiration dates are only ordered as needed to reduce waste.

14.3. Using the automated system requires the use of higher volumes of buffer, however up to three determinations can be done using a single aliquot of sample.

15. Data Assessment and Acceptance Criteria for Quality Control Measures

15.1. Initial Calibration

15.1.1. Poseidon

15.1.1.1. The resultant slope must be 96% - 101%.

15.1.1.2. The ICV result must be ±0.1 pH units of the assigned value.

15.1.1.3. Sample analysis cannot proceed until these requirements have been meet.

15.1.2. Memphis

15.1.2.1. The slope of the calibration must be 92-102 % to be acceptable.

15.1.2.2. ICV must read to within ±0.1 pH units of assigned value.

15.2. Continuing Calibration

15.2.1. The CCV must read to within ±0.1 pH units of assigned value.

15.3. Batch QC

15.3.1. DUP must read to within ±0.1 pH units from original sample.

16. Corrective Actions For Out of Control Data

16.1. Initial Calibration

16.1.1. Poseidon

16.1.1.1. If the slope is not within the specified range, reattempt calibration. Next replace buffer solutions and recalibrate. Finally, perform probe maintenance and recalibrate.

16.1.1.2. If the ICV standard fails, replace ICV solution and reanalyze. Perform probe maintenance, recalibrate and reanalyze if ICV continues to fail. Analysis cannot be performed if the ICV is unacceptable.

16.1.2. Memphis

16.1.2.1. If the slope of the calibration curve is not between 92-102% the probe icon will be flashing and samples may not be analyzed. First attempt recalibration. Next replace buffer solutions and recalibrate. Finally perform probe maintenance and recalibrate.

16.1.2.2. If the ICV standard fails, replace ICV solution and reanalyze. Perform probe maintenance, recalibrate and reanalyze if ICV continues to fail. Analysis cannot be performed if the ICV is unacceptable.

16.2. Continuing Calibration

16.2.1. If the CCV is unacceptable, reanalyze. If CCV continues to fail perform probe maintenance recalibrate meter and reanalyze all samples to the previous acceptable CCV.

16.3. Batch QC

16.3.1. If sample DUP is unacceptable, reanalyze. If DUP continues to fail, reanalyze both sample and DUP, and report reanalysis. If DUP is still unacceptable, report as is and flag duplicate sample.

17. Contingencies For Handling Out of Control Data

17.1. All out-of-control data is flagged, a corrective action generated, and reported to the client.

18. Waste Management

18.1. After analysis all samples, sample rinsates, and standards are disposed of in accordance with the HEAL Chemical Hygiene Plan.

19. References

19.1. Standard Methods for the Examination of Water and Wastewater, 20th Edition, Method 4500H+ B, 1999.

19.2. EPA Method150.1 – pH (Electrometric). NPDES Editorial Revision 1978; 1982.

19.3. SW-846 Method 9045C – Soil and Waste pH. Revision 3, 1995.

19.4. HEAL Quality Assurance Plan – Current Version.

19.5. HEAL Chemical Hygiene Plan – Current Version

19.6. *Tiamo* Software Help section.

Appendix A

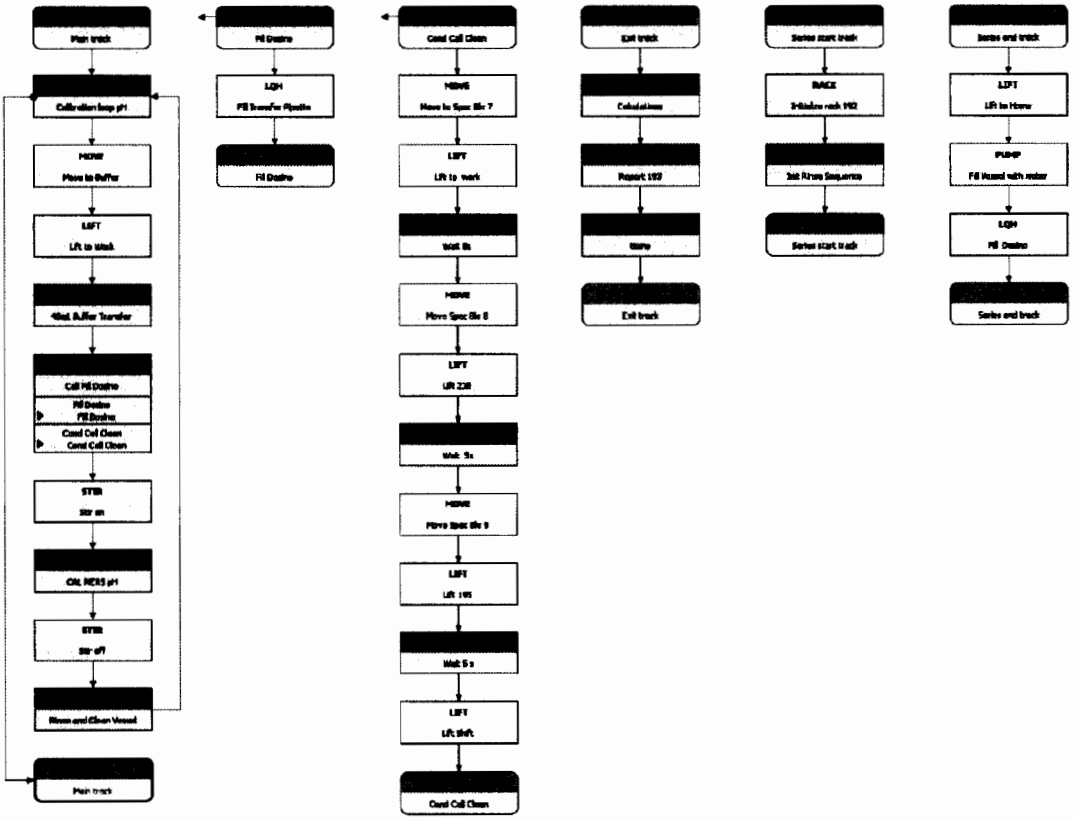
Method Flow Charts and Method Parameters

The method flow chart and method parameters for pH sample analysis are included in the current SOP for alkalinity determination. These two tests can be done conjointly.

tiamo 1.2.1 Build 43 | PC designation: HALLXP | User (short name): bdh
License ID: 6369515 | Printed: 2009-02-03 15:23:17 UTC-7

Method run

Method name: pH Calibration
Method version: 67
Method group: Main group



Method parameters

Method ..... pH Calibration
Method saving date 2008-11-19 16:11:18 UTC-7
Method version 67
Method group ..... Main group
Method status ..... original
Method saved by (full name) ..... Karen Stasiunas
Method saved by (short name) ..... kms

START **Main track**

General

Workplace view

Current view ..... on

Track view for live window

Live display 1 ..... Main track

Live display 2 ..... Main track

Statistics ..... off

Conditioning

Automatic conditioning ..... off

Application note

pH CAL takes place in Special Beakers 1 - 3 Special Beaker 1 = Position 53 = Buffer 4
Special Beaker 2 = Position 54 = Buffer 7 Special Beaker 3 = Position 55 = Buffer 10

Method variables

Name	Type	Assignment	Value	Comment	Monitoring
Sample size	Number	Sample size		Sample size	off
Sample size unit	Text	Sample size unit		Sample size unit	off
Sample position	Number	Sample position		Sample position number	off
ID1	Text	ID1		Sample identification 1	off
ID2	Text	ID2		Sample identification 2	off
ID3	Text	ID3		Sample identification 3	off

Name ..... **Sample size**
Type ..... Number
Assignment ..... on ..... Sample size
Value ..... off .....
Check at start ..... off
Comment ..... Sample size
Variable monitoring ..... off
Lower limit .....
Upper limit .....
Message .....
Display message ..... on
Record message ..... on
Message by e-mail ..... off

Mail to .....
Subject ..... Message from tiamo - Method 'New method 2' - Command 'Main track'
User .....
Mail from .....
SMTP Server .....
POP3 Server .....
Acoustic signal ..... off
Action ..... off
Stop determination ..... on
Stop determination and series ..... off

Name ..... **Sample position**
Type ..... Number
Assignment ..... on ..... Sample position
Value ..... off .....
Check at start ..... off
Comment ..... Sample position number
Variable monitoring ..... off
Lower limit .....
Upper limit .....
Message .....
Display message ..... on
Record message ..... on
Message by e-mail ..... off
Mail to .....
Subject ..... Message from tiamo - Method 'New method 2' - Command 'Main track'
User .....
Mail from .....
SMTP Server .....
POP3 Server .....
Acoustic signal ..... off
Action ..... off
Stop determination ..... on
Stop determination and series ..... off

Name ..... **Sample size unit**
Type ..... Text
Assignment ..... on ..... Sample size unit
Value ..... off .....
Check at start ..... off
Comment ..... Sample size unit

Name **ID1**
Type ..... Text
Assignment ..... on ID1

Value ..... off .....
Check at start ..... off
Comment ..... Sample identification 1

Name ID2
Type ..... Text
Assignment ..... on ID2
Value ..... off .....
Check at start ..... off
Comment ..... Sample identification 2

Name ID3
Type ..... Text
Assignment ..... on ID3
Value ..... off .....
Check at start ..... off
Comment ..... Sample identification 3

CAL LOOP pH **Calibration loop pH**

Buffers

Number of buffers 5
Buffer type ..... Special
Buffer 1 pH 1.680
Buffer 2 pH 4.000
Buffer 3 pH 7.000
Buffer 4 pH 10.000
Buffer 5 pH 12.450
Request for buffer exchange ..... off
Subsequent command ..... Main track

MOVE **Move to Buffer**

Device

Device name ..... 855_1

Target

Tower 1
Move ..... Special beaker
Number ..... = 'Calibration loop pH.LCO'

Beaker test

Display message ..... on
Stop determination ..... off
Stop determination and series ..... off

Parameters

Shift rate ..... 20 °/s
Shift direction ..... auto
Swing rate ..... 55 °/s

LIFT **Lift to Work**

Value ..... off
Check at start ..... off
Comment ..... Sample identification 1

Name ID2
Type ..... Text
Assignment ..... on ID2
Value ..... off
Check at start ..... off
Comment ..... Sample identification 2

Name ID3
Type ..... Text
Assignment ..... on ID3
Value ..... off
Check at start ..... off
Comment ..... Sample identification 3

CAL LOOP pH **Calibration loop pH**

Buffers

Number of buffers 5
Buffer type ..... Special
Buffer 1 pH 1.680
Buffer 2 pH 4.000
Buffer 3 pH 7.000
Buffer 4 pH 10.000
Buffer 5 pH 12.450
Request for buffer exchange ..... off
Subsequent command ..... Main track

MOVE **Move to Buffer**

Device

Device name ..... 855_1

Target

Tower 1
Move ..... Special beaker
Number ..... = 'Calibration loop pH.LCO'

Beaker test

Display message ..... on
Stop determination ..... off
Stop determination and series ..... off

Parameters

Shift rate ..... 20 °/s
Shift direction ..... auto
Swing rate ..... 55 °/s

LIFT **Lift to Work**

Dosing device 1
Solution . . . . . Transfer Pipette

Parameters

Function . . . . . Aspirate
Port 1
Volume . . . . . 15.00 mL
Rate . . . . . maximum mL/min

LQH **Dispense remaining sample**

General/Hardware

Device

Device name . . . . . 855_1

Dosing device

Dosing device 1
Solution . . . . . Transfer Pipette

Parameters

Function . . . . . Eject to end volume
Port 3
Rate . . . . . maximum mL/min

CALL **Call Fill Dosino**

Call text	Track name	Condition	Condition
Fill Dosino	Fill Dosino	off	
Cond Cell Clean	Cond Cell Clean	off	

STIR **Stir on**

Device

Device name . . . . . 855_1

Stirrer

Stirrer 3
Stirrer type . . . . . 801 Magnetic stirrer
Stirring rate 5

Action

Switch on . . . . . on
Switch off . . . . . off
Duration . . . . . off

CAL MEAS pH **CAL MEAS pH**

General/Hardware

Device

Device name . . . . . 855_1

Sensor

Measuring input 1
Sensor . . . . . pH electrode
Temperature measurement . . . . . automatic

Stirrer

Stirrer . . . . . off

Measuring parameters

Measurement with drift control

Signal drift ..... 2.0 mV/min
Min. waiting time ..... 30 s
Max. waiting time ..... 60 s
Measuring interval ..... 2.0 s

Temperature

Temperature ..... 25.0 °C

STIR **Stir off**

Device

Device name ..... 855_1

Stirrer

Stirrer 3
Stirrer type ..... unknown
Stirring rate 8

Action

Switch on ..... off
Switch off ..... on
Duration ..... off

SEQUENCE **Rinse and Clean Vessel**

PUMP **Asp Pump On**

Device

Device name ..... 855_1

Pumps

Tower 1
Pump(s) 2

Action

Switch on ..... on
Switch off ..... off
Duration ..... off

WAIT **Wait 12s**

Wait

Stop track and waiting for [Continue] ..... off
Stop all tracks and waiting for [Continue] ..... off
Waiting time ..... on
Time 12
Unit ..... s

Message

Record message ..... off
Message by e-mail ..... off
Acoustic signal ..... off

MOVE **Move Spec 5**

Device

Device name ..... 855_1

Target
- Tower 1
- Move . . . . . Special beaker
- Number 8

Beaker test
- Display message . . . . . on
- Stop determination . . . . . off
- Stop determination and series . . . . . off

Parameters
- Shift rate . . . . . 20 °/s
- Shift direction . . . . . auto
- Swing rate . . . . . 55 °/s

LQH **Clean Port 1 Tubing**

General/Hardware

Device
- Device name . . . . . 855_1

Dosing device
- Dosing device 1
- Solution . . . . . Transfer Pipette

Parameters
- Function . . . . . Dosing
- Port 1
- Volume . . . . . 3.25 mL
- Rate . . . . . maximum mL/min

LQH **Clean port 3 tubing**

General/Hardware

Device
- Device name . . . . . 855_1

Dosing device
- Dosing device 1
- Solution . . . . . Transfer Pipette

Parameters
- Function . . . . . Eject to end volume
- Port 3
- Rate . . . . . maximum mL/min

LIFT **Shift Pos**

Device
- Device name . . . . . 855_1

Target
- Tower 1
- Lift position . . . . . Shift position mm

Parameters
- Lift rate . . . . . 25 mm/s

LQH **Fill with air**

General/Hardware

Device

Device name ..... 855_1

Dosing device

Dosing device 1

Solution ..... Transfer Pipette

Parameters

Function ..... Aspirate

Port 1

Volume ..... 15.00 mL

Rate ..... maximum mL/min

LQH **Dispense Air**

General/Hardware

Device

Device name ..... 855_1

Dosing device

Dosing device 1

Solution ..... Transfer Pipette

Parameters

Function ..... Eject to end volume

Port 3

Rate ..... maximum mL/min

PUMP **Rinse Pump-On**

Device

Device name ..... 855_1

Pumps

Tower 1

Pump(s) 1

Action

Switch on ..... off

Switch off ..... off

Duration ..... on

Time ..... 4.0 s

WAIT **Wait 7s**

Wait

Stop track and waiting for [Continue] ..... off

Stop all tracks and waiting for [Continue] ..... off

Waiting time ..... on

Time 7

Unit ..... s

Message

Record message ..... off

Message by e-mail ..... off

Acoustic signal ..... off

PUMP **Pump on**

Device
- Device name ..... 855_1

Pumps
- Tower 1
- Pump(s) 1

Action
- Switch on ..... off
- Switch off ..... off
- Duration ..... on
- Time ..... 4.0 s

WAIT **Wait 10s**

Wait
- Stop track and waiting for [Continue] ..... off
- Stop all tracks and waiting for [Continue] ..... off
- Waiting time ..... on
  - Time 10
  - Unit ..... s

Message
- Record message ..... off
- Message by e-mail ..... off
- Acoustic signal ..... off

PUMP **Pump Off**

Device
- Device name ..... 855_1

Pumps
- Tower 1
- Pump(s) 2

Action
- Switch on ..... off
- Switch off ..... on
- Duration ..... off

TRACK **Fill Dosino**
- Return immediately ..... on
- Delete all data ..... off

LQH **Fill Transfer Pipette**

General/Hardware
- Device
  - Device name ..... 855_1
- Dosing device
  - Dosing device 1
  - Solution ..... Transfer Pipette

Parameters
- Function ..... Fill

Port 2
Rate . . . . . maximum mL/min

TRACK **Cond Cell Clean**
Return immediately . . . . . on
Delete all data . . . . . off

MOVE **Move to Spec Bkr 7**
Device
Device name . . . . . 855_1
Target
Tower 1
Move . . . . . Special beaker
Number 7
Beaker test
Display message . . . . . on
Stop determination . . . . . off
Stop determination and series . . . . . off
Parameters
Shift rate . . . . . 20 °/s
Shift direction . . . . . auto
Swing rate . . . . . 55 °/s

LIFT **Lift to work**
Device
Device name . . . . . 855_1
Target
Tower 1
Lift position . . . . . 218 mm
Parameters
Lift rate . . . . . 25 mm/s

WAIT **Wait 5s**
Wait
Stop track and waiting for [Continue] . . . . . off
Stop all tracks and waiting for [Continue] . . . . . off
Waiting time . . . . . on
Time 5
Unit . . . . . s
Message
Record message . . . . . off
Message by e-mail . . . . . off
Acoustic signal . . . . . off

MOVE **Move Spec Bkr 8**
Device
Device name . . . . . 855_1
Target
Tower 1

Move ..... Special beaker
Number 8
Beaker test
Display message ..... on
Stop determination ..... off
Stop determination and series ..... off
Parameters
Shift rate ..... 20 °/s
Shift direction ..... auto
Swing rate ..... 55 °/s

LIFT **Lift 226**
Device
Device name ..... 855_1
Target
Tower 1
Lift position ..... 218 mm
Parameters
Lift rate ..... 25 mm/s

WAIT **Wait 5s**
Wait
Stop track and waiting for [Continue] ..... off
Stop all tracks and waiting for [Continue] ..... off
Waiting time ..... on
Time 5
Unit ..... s
Message
Record message ..... off
Message by e-mail ..... off
Acoustic signal ..... off

MOVE **Move Spec Bkr 9**
Device
Device name ..... 855_1
Target
Tower 1
Move ..... Special beaker
Number 9
Beaker test
Display message ..... on
Stop determination ..... off
Stop determination and series ..... off
Parameters
Shift rate ..... 20 °/s
Shift direction ..... auto
Swing rate ..... 55 °/s

LIFT **Lift 195**

Device
- Device name ..... 855_1

Target
- Tower 1
- Lift position ..... 218 mm

Parameters
- Lift rate ..... 25 mm/s

WAIT **Wait 5 s**

Wait
- Stop track and waiting for [Continue] ..... off
- Stop all tracks and waiting for [Continue] ..... off
- Waiting time ..... on
  - Time 5
  - Unit ..... s

Message
- Record message ..... off
- Message by e-mail ..... off
- Acoustic signal ..... off

LIFT **Lift Shift**

Device
- Device name ..... 855_1

Target
- Tower 1
- Lift position ..... Shift position mm

Parameters
- Lift rate ..... 25 mm/s

EXIT **Exit track**

CALC **Calculations**

Result name	Formula	Unit	Decimal places	Assignment	Statistics
pH Slope	= 'Calibration loop pH.SLO'		2	RS24	off
pH(as)	= 'Calibration loop pH.ENP'		2	RS25	off

Result name ..... **pH Slope**
Formula ..... = 'Calibration loop pH.SLO'
Unit .....
Decimal places 2
Assignment RS24
Statistics ..... off
Description ..... RS.'Result name'[.VAL] Result value.
Result monitoring ..... off

Save result as common variable . . . . . off
Name . . . . . .
Save result as titer . . . . . off
Solution name . . . . . .

Result name . . . . . **pH(as)**
Formula . . . . . = 'Calibration loop pH.ENP'
Unit . . . . . .
Decimal places 2
Assignment RS25
Statistics . . . . . off
Description . . . . . RS.'Result name'[.VAL] Result value.
Result monitoring . . . . . off
Save result as common variable . . . . . off
Name . . . . . .
Save result as titer . . . . . off
Solution name . . . . . .

REPORT **Report 193**
Report template
Report template . . . . . pH Calibration Report
Report output
Printer . . . . . on
Printer . . . . . Default printer
PDF file . . . . . off

DATABASE **tiamo**

Database

Results

SERIES START **Series start track**

RACK **Initialize rack 192**
Device
Device name . . . . . 855_1
Rack test . . . . . off

SEQUENCE **Init Rinse Sequence**

PUMP **Pump On**
Device
Device name . . . . . 855_1
Pumps
Tower 1
Pump(s) 2
Action

Switch on ..... on
Switch off ..... off
Duration ..... off

WAIT **Wait 8s**

Wait
- Stop track and waiting for [Continue] ..... off
- Stop all tracks and waiting for [Continue] ..... off
- Waiting time ..... on
  - Time 8
  - Unit ..... s

Message
- Record message ..... off
- Message by e-mail ..... off
- Acoustic signal ..... off

LQH **Make room in Dosino**

General/Hardware
- Device
  - Device name ..... 855_1
- Dosing device
  - Dosing device 1
  - Solution ..... Transfer Pipette

Parameters
- Function ..... Eject to end volume
- Port 3
- Rate ..... maximum mL/min

LQH **Fill w- air**

General/Hardware
- Device
  - Device name ..... 855_1
- Dosing device
  - Dosing device 1
  - Solution ..... Transfer Pipette

Parameters
- Function ..... Aspirate
- Port 1
- Volume ..... 15.00 mL
- Rate ..... maximum mL/min

LQH **Eject to EV**

General/Hardware
- Device
  - Device name ..... 855_1
- Dosing device
  - Dosing device 1
  - Solution ..... Transfer Pipette

Parameters

Function . . . . . Eject to end volume
Port 3
Rate . . . . . maximum mL/min

WAIT **Wait 3s**

Wait
Stop track and waiting for [Continue] . . . . . off
Stop all tracks and waiting for [Continue] . . . . . off
Waiting time . . . . . on
Time 3
Unit . . . . . s

Message
Record message . . . . . off
Message by e-mail . . . . . off
Acoustic signal . . . . . off

PUMP **Rinse 3s**

Device
Device name . . . . . 855_1

Pumps
Tower 1
Pump(s) 1

Action
Switch on . . . . . off
Switch off . . . . . off
Duration . . . . . on
Time . . . . . 3.0 s

WAIT **Wait 8 s**

Wait
Stop track and waiting for [Continue] . . . . . off
Stop all tracks and waiting for [Continue] . . . . . off
Waiting time . . . . . on
Time 8
Unit . . . . . s

Message
Record message . . . . . off
Message by e-mail . . . . . off
Acoustic signal . . . . . off

PUMP **Asp Pump Off**

Device
Device name . . . . . 855_1

Pumps
Tower 1
Pump(s) 2

Action

Switch on ..... off
Switch off ..... on
Duration ..... off

SERIES END **Series end track**

LIFT **Lift to Home**

Device
Device name ..... 855_1
Target
Tower 1
Lift position ..... Home position mm
Parameters
Lift rate ..... 25 mm/s

PUMP **Fill Vessel with water**

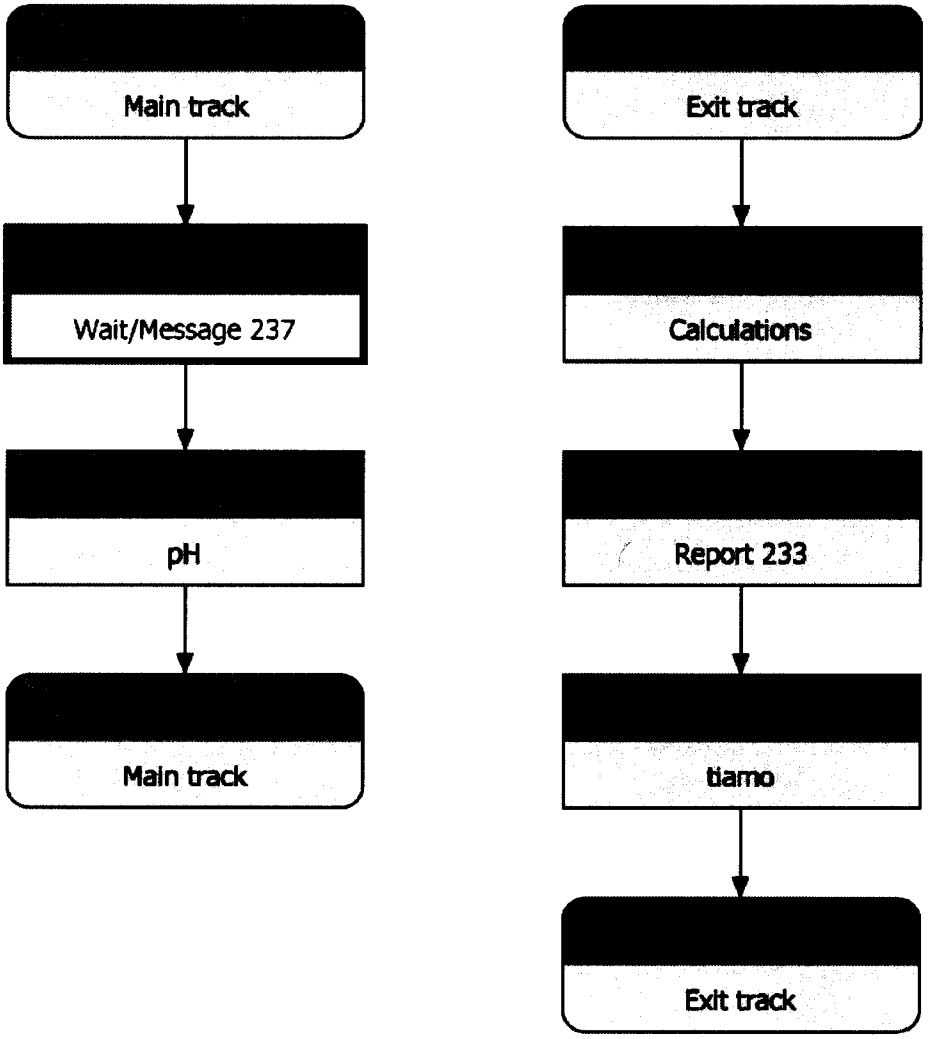
Device
Device name ..... 855_1
Pumps
Tower 1
Pump(s) 1
Action
Switch on ..... off
Switch off ..... off
Duration ..... on
Time ..... 4.0 s

LQH **Fill Dosino**

General/Hardware
Device
Device name ..... 855_1
Dosing device
Dosing device 1
Solution ..... Transfer Pipette
Parameters
Function ..... Fill
Port 2
Rate ..... maximum mL/min

Method run

Method name:	pH manual
Method version:	3
Method group:	Main group



Method parameters

Method . . . . . pH manual
Method saving date 2009-10-05 14:25:36 UTC-6
Method version 3
Method group . . . . . Main group
Method status . . . . . original
Method saved by (full name) . . . . . Benjamin Derek Harmon
Method saved by (short name) . . . . . bdh

START **Main track**

General

Workplace view

Current view . . . . . on

Track view for live window

Live display 1 . . . . . Main track

Live display 2 . . . . . Main track

Statistics . . . . . off

Conditioning

Automatic conditioning . . . . . off

Application note

Method variables

Name	Type	Assignment	Value	Comment	Monitoring
Sample size	Number	Sample size		Sample size	off
Sample size unit	Text	Sample size unit		Sample size unit	off
Sample position	Number	Sample position		Sample position number	off
ID1	Text	ID1		Sample identification 1	off
ID2	Text	ID2		Sample identification 2	off
ID3	Text	ID3		Sample identification 3	off
cond	Text				off
temp	Text				off

Name . . . . . **Sample size**
Type . . . . . Number
Assignment . . . . . on . . . . . Sample size
Value . . . . . off . . . . . .
Check at start . . . . . off
Comment . . . . . Sample size
Variable monitoring . . . . . off
Lower limit . . . . . .
Upper limit . . . . . .
Message . . . . . .
Display message . . . . . on
Record message . . . . . on

Message by e-mail . . . . . off
Mail to . . . . . .
Subject . . . . . Message from tiamo - Method 'New method 2' - Command 'Main track'
User . . . . . .
Mail from . . . . . .
SMTP Server . . . . . .
POP3 Server . . . . . .
Acoustic signal . . . . . off
Action . . . . . off
Stop determination . . . . . on
Stop determination and series . . . . . off

Name . . . . . **Sample position**
Type . . . . . Number
Assignment . . . . . on . . . . . Sample position
Value . . . . . off . . . . . .
Check at start . . . . . on
Comment . . . . . Sample position number
Variable monitoring . . . . . off
Lower limit . . . . . .
Upper limit . . . . . .
Message . . . . . .
Display message . . . . . on
Record message . . . . . on
Message by e-mail . . . . . off
Mail to . . . . . .
Subject . . . . . Message from tiamo - Method 'New method 2' - Command 'Main track'
User . . . . . .
Mail from . . . . . .
SMTP Server . . . . . .
POP3 Server . . . . . .
Acoustic signal . . . . . off
Action . . . . . off
Stop determination . . . . . on
Stop determination and series . . . . . off

Name . . . . . **Sample size unit**
Type . . . . . Text
Assignment . . . . . on . . . . . Sample size unit
Value . . . . . off . . . . . .
Check at start . . . . . off
Comment . . . . . Sample size unit

Name **ID1**
Type . . . . . Text

Assignment ..... on ID1
Value ..... off .....
Check at start ..... on
Comment ..... Sample identification 1

Name **ID2**
Type ..... Text
Assignment ..... on ID2
Value ..... off .....
Check at start ..... off
Comment ..... Sample identification 2

Name **ID3**
Type ..... Text
Assignment ..... on ID3
Value ..... off .....
Check at start ..... off
Comment ..... Sample identification 3

Name ..... **cond**
Type ..... Text
Assignment ..... off .....
Value ..... on .....
Check at start ..... off
Comment .....

Name ..... **temp**
Type ..... Text
Assignment ..... off .....
Value ..... on .....
Check at start ..... off
Comment .....

WAIT **Wait/Message 237**

Wait

Stop track and waiting for [Continue] ..... off
Stop all tracks and waiting for [Continue] ..... off
Waiting time ..... on
Time 60
Unit ..... s

Message

Record message ..... off
Message by e-mail ..... off
Acoustic signal ..... off

MEAS pH **pH**

General/Hardware

Device

Device name . . . . . 855_1

Sensor

Measuring input 1

Sensor . . . . . pH electrode

Temperature measurement . . . . . automatic

Stirrer

Stirrer . . . . . off

Measuring parameters

Measurement

Measurement with drift control . . . . . on

Signal drift . . . . . 1.0 mV/min

Min. waiting time . . . . . 60 s

Max. waiting time . . . . . 180 s

Measuring interval . . . . . 2.0 s

Stop measured value pH . . . . . off

Measurement without drift control . . . . . off

Temperature

Temperature . . . . . 25.0 °C

Evaluations

Fix end point evaluation . . . . . off

Minimum evaluation . . . . . off

Maximum evaluation . . . . . off

Break point evaluation . . . . . off

Additional measured values

Additional calculated measured values . . . . . off

Additional external measured values . . . . . off

EXIT **Exit track**

CALC **Calculations**

Result name	Formula	Unit	Decimal places	Assignment	Statistics
pH	= 'pH.EME'	pH units	2	RS03	off
pH Temp	= 'pH.ETE'	°C	1	RS04	off

Result name . . . . . **pH**

Formula . . . . . = 'pH.EME'

Unit . . . . . pH units

Decimal places 2

Assignment RS03

Statistics . . . . . off

Description . . . . . RS.'Result name'[.VAL] Result value.

Result monitoring . . . . . off

Save result as common variable . . . . . off

Name . . . . . .

Save result as titer . . . . . off

Solution name . . . . . .

Result name . . . . . **pH Temp**
Formula . . . . . = 'pH.ETE'
Unit . . . . . °C
Decimal places 1
Assignment RS04
Statistics . . . . . off
Description . . . . . .
Result monitoring . . . . . off
Save result as common variable . . . . . off
Name . . . . . .
Save result as titer . . . . . off
Solution name . . . . . .

REPORT **Report 233**

Report template
Report template . . . . . New Results Report
Report output
Printer . . . . . on
Printer . . . . . Default printer
PDF file . . . . . off

DATABASE **tiamo**

Database

090309 starting

Trace Metal Analysis by EPA Method 6020A

Applicable Matrices: Non-Potable Water, Sludges, Soils, Paint, Oil, Fertilizers

MDL: The Instrument Detection Limit must be evaluated quarterly (see procedure below). See individual instrument activity logbook and/or MDL file on server for data.

Scope and Application: This method provides procedures for determination of total recoverable element concentrations in soils, sludges, oil, fertilizers and paint.

Summary of test method: Prior to analysis samples that require total (acid-leachable) values must be digested using appropriate sample preparation methods, EPA 1311 or 1312.

EPA 6020A describes the multi-element determination of trace elements by ICP-MS. Sample material in solution is introduced by pneumatic nebulization into a radiofrequency inductively coupled plasma where energy transfer processes cause desolvation, atomization, and ionization. The ions are extracted from the plasma through a differentially pumped vacuum interface and separated on the basis of their mass to charge ratio by a quadrupole mass spectrometer having a minimum resolution capability of 1 amu peak width at 5% peak height. An electron multiplier detects the ions transmitted through the quadrupole and the ion information is processed by a data handling system. Interferences related to the technique must be recognized and corrected for. Such corrections must include compensation for isobaric elemental interferences and interferences from polyatomic ions derived from the plasma gas, reagents, or sample matrix. Instrumental drift as well as suppressions or enhancements of instrument response caused by the sample matrix must be corrected for by the use of internal standards.

Definitions: Refer to QA Manual, 'Internal Quality Control' section.

Interferences: Refer to EPA Method 6020A for more detailed information on interferences. The following is a brief description of some interferences.
Isobaric elemental interferences – Are caused by isotopes of different elements which form singly or doubly charged ions of the same nominal mass-to-charge ratio and which cannot be resolved by the mass spectrometer in use.
Abundance sensitivity – Is a property defining the degree to which the wings of a mass peak contribute to adjacent masses.
Isobaric polyatomic ion interferences – Are caused by ions consisting of more than one atom which have the same nominal mass-to-charge ratio as the isotope of interest, and which cannot be resolved by the mass spectrometer in use.
Physical interferences – Are associated with the physical processes which govern the transport of sample into the plasma, sample conversion process in the plasma and the transmission of ions through the plasma-mass spectrometer interface.
Memory interferences – Result when isotopes of elements in a previous sample contribute to the signals measured in a new sample.

Safety: The toxicity and carcinogenicity of reagents used in this method have not been fully established. Concentrated nitric and hydrochloric acids present various hazards and are moderately toxic and extremely irritating to the skin and mucus membranes. Use these reagents in a fume hood whenever possible and if eye or skin contact occurs, flush with large volumes of water.
The acidification of samples containing reactive materials may result in the release of toxic gases, such as cyanides or sulfides.

Equipment and Supplies:

a. Agilent 7500 ICP-MS with an Integrated Sample Introduction System and CETAC ASX-510 Autosampler.
b. CPI MOD-BLOC digestion apparatus with accompanying graduated digestion vessels and watchglasses.
c. Milli-Q 50 water purification system.

Reagents and Standards:

The listed grade of reagents is the minimum that may be used for metals analysis.

a. Trace Metal Grade Nitric Acid.
b. Trace Metal Grade Hydrochloric Acid.
c. Hydrogen Peroxide 30%.
d. 18 MΩ deionized water.

Calibration Solutions

*Use 1% HNO3 and 0.5% HCL for preservation of all solutions and standards.

Auto Tune Solution – 10 ppb Co, Li, Y, Ce, Tl. Dilute 1 mL 10 ppm Auto Tune Standard and 1 mL Co Stock Standard into 1000 mL volumetric flask and bring to volume with 18 MΩ deionized water.

P/A Factor Solution – 100 ppb multi element standard (Use unused 100 ppb CAL from previous day).

Tune Solution – 100 ppb Be, Mg, Co, In, Pb. Dilute 10 mL 10 ppm Tune Standard into 1000 mL volumetric flask and bring to volume with 18 MΩ deionized water.

Internal Standard – 0.5 ppm Ho, 1 ppm Sc & Rh, 2 ppm Li 6, and 4 ppm Ge. Dilute 0.250 mL 1000ppm Ho, 0.500 mL 1000 ppm Sc & Rh, 1.00 mL Li, and 2 mL 1000 ppm Ge standard in a 500mL volumetric flask and bring to volume with Milli-Q H2O.

Dilution Standard –100 ppb Ce. Dilute 100 uL 1000 ppm Ce in a 1000 mL volumetric flask and bring to volume with 18 MΩ deionized water.

Calibration Blank - 18 MΩ deionized water.

Metals Calibration Standards – Analytes at concentrations of 1, 25, 50, and 100 ppb (Fe & Al at 2.0, 50, 100, 200 and Hg at 0.05, 1.25, 2.50, and 5.00 ppb).

Cation Calibration Standards – Na, Mg, Si, K, S, and Ca at concentrations of 100, 1000, 5000, 10000, 25000 ppb.

ICV – Concentration of 40 ppb (Fe & Al at 80 ppb and Hg at 4 ppb)

Cation ICV 12,500 – Concentration of 12,500 ppb Na, Mg, Si, K, and Ca.

LCS/MS/MSD – For Metals analysis dilute appropriate stock standards to 50 ppb in 10mL. (i.e., 0.050 mL 10ppm solution into 10mL of sample.) Cation LCS/MS/MSD is prepared from a 500 ppm stock standard prepared from certified standards to achieve a 5 ppm spike in 10 mL of sample.

ICS-A Standard – Dilute 5 mL Stock ICS-A standard to a final volume of 50 ml.

ICS-AB Standard – Dilute 5 mL Stock ICS-A standard and 0.05 mL of Stock ICS-B standard to a final volume of 50 ml.

Sample Collection, Preservation, Shipment and Storage:

Soils, oils, paints, fertilizers, and sludges have an established holding time of 180 days, but must be kept refrigerated below 4 degrees Celsius.

Quality Control:

IS - Internal standard areas - Monitor for all analyses, use CALB as reference.

All samples:	70% - 130% of CALB
CALIB	80% - 120% of CALB
All Other	70% - 130% of CALB

TUNE – A tune solution must be run every 8 hours to verify the RSD is < 5.00% and the peak location is +/- 0.1 amu for Be, Mg, Co, In, and Pb.

ICS_A – Analyze ICS_A every 12 hours and assess the data for potential interference with analytes of interest.

ICS_AB - Quantify the following every 12 hours: As, Cd, Co, Cr, Cu, Mn, Ni, Se, Ag, V, and Zn at 16 to 24 ppb.

*** DO NOT run a full metals list with an ICS because the high concentrations of interferences causes instability and can cause a sequence to be aborted due to the inability to recover the calibration to acceptable limits.**

LRB – One blank per 20 samples for each matrix must be prepared and analyzed. For waters the results for each analyte may not exceed 2.2 times the MDL or 10 % of the sample concentration, whichever is greater. For soils, the concentration may not exceed 3 times the MDL.

MS/MSD - Once every 20 samples or 1 per batch, which ever is less. Spike and digest at 50 ppb (2.5 ppb for Hg). All metals that were less than 200 ppb in unspiked digest should give recovery of ±25 %. For those over 200 ppb, dilute five fold and re-analyze. Results for over 200 ppb metals must agree within 10 %.

LCS - Once every 20 samples for each sample matrix. Spike a blank at 50 ppb target metals and 2.5 ppb Hg. Acceptable recovery is 85-115%.

ICV - Once after every CALIB. Sample must be near the mid point of the calibration curve. All target metals must agree to within 10% for analysis to continue.

CCB - Once every 10 samples. CCB values must not exceed 1.0 ppb for any metal. (100 ppb for cations)

CCV - Once every 10 samples. CCV (50 ppb CAL) values must agree to within 10 % or the instrument must be re-calibrated and the samples in the previous analytical block re-analyzed. The concentration analyzed for a CCV must change once quarterly to verify the validity of all points on the curve.

Dilution Standard – The dilution standard must be analyzed every 20 samples to verify it is within 20% of the original dilution standard set just after the calibration and must also be between 8 and 12 fold.

DUP - Perform a duplicate analysis every 20 samples. All metals over 20 ppb in the digest must agree to within 20% RPD, or a cause must be determined.

Instrument Detection Limit (IDL):

Estimate the IDL by calculating the average of the standard deviations of three runs on three non-consecutive days from the analysis of a reagent blank solution with seven consecutive measurements per day. Each measurement must be performed as though it were a separate analytical sample (i.e., each measurement must be followed by a rinse and/or any other procedure normally performed between the analysis of separate samples). IDLs must be determined at least every three months and kept with the instrument log book.

Calibration and Standardization:

Refer to Calibration Standards and procedure for calibration information. Follow recommended operating conditions provided by the manufacturer.

Procedure:

Sample Preparation

a. Solid Sample Hot Plate Digestion

Solid samples (soils, sludges, composts, and biosolids) typically contain moisture and thus the percent solid of all solid samples needs to be computed. Percent solids is performed by drying about 5.00 g of wet solid in an oven set at 110 °C for about two hours and is recorded in the percent solids logbook. The dry weight divided by the wet weight is taken as the % solids. Prepare soils by weighing approximately 5 grams (soil, sludge, biosolid) (2.5 grams compost) into a graduated digestion vessel. Add 10 mL 18 MΩ water, 10 mL conc. Nitric acid, and 5 mL conc. HCL. Gently swirl the samples before placing them on the MOD BLOC digester. Allow the samples to digest for 2 hours (DO NOT LET THE SAMPLES GO DRY) then remove and allow the samples to cool. Add 10 mL 18 MΩ water and add 30% H_2O_2 one mL at a time (CAUTION) until effervescence has ceased or 4 mL has been added, whichever is less. Allow samples to digest for an additional hour then remove and allow the samples to cool. Bring the samples up to 50 mL with 18 MΩ water, cap, and gently shake. Allow the samples to settle on the bench-top overnight before analysis. Dilute soil samples 50 fold from the digestion vessel for analysis. A multiplier must be calculated for these samples as follows.

$$M = \frac{2500}{weight\ of\ sample \times \%soilds\ (decimal\ )}$$

b. Preparation of TCLP Samples

TCLP extracts should be diluted 10 fold in order to overcome interferences from extraction fluid. Refer to Method 1311 TCLP Extraction SOP for extraction.

c. Fertilizers. Special care must be taken when dealing with fertilizers so as to not contaminate the ICP with analytes such as Fe, K, Mg, etc. Consult inorganic supervisor for procedure. In general, dilute liquid fertilizers 500x into a digestion vessel and treat like a waste-water. Solid samples should use about 1 gram and be treated like a soil. Final dilutions should be 5000 for liquids and 10,000 for solids.

Calculations:

Interference Equations:

**V (51) = M (51) – 3.127 × {M (53) –0.113 × M (52)}

**As (75) = M (75) – 3.132 × M (77) +2.736 × M (82) –2.761 × Kr (83)

**Mo (98) = M (98) – 0.146 × M (99)

**Cd (114) = M (114) – 1.628 × M(108) + 1.115 × Cd (106) + 1.934 × Pd (105) – 0.026 ×M (118)

Cd (111) = M (111) – 1.074 × M (108) + 0.736 × M (106) +0.376 × Pd (105)

Pb (208) = M (208) + (207) + (206)

U (238) = M (238) + (234) + (235)

** Equations not needed for 7500 ICP-MS when using reaction mode

Method Performance: Refer to MDL/IDL and EPA Method 6020A

Pollution Prevention: Refer to EPA Method 6020A

Data Assessment and Acceptance Criteria for QC Measures: Refer to QC above.

Corrective Actions for Out-Of-Control or Unacceptable Data: Refer to 'Corrective Action' section of Anatek QA Plan

Waste Management: Refer to EPA 6020A and the 'Waste Disposal' section of QA Plan

References:

1. EPA Method 6020A
2. EPA Method 3051A
3. EPA Method 3050B
4. EPA Method 1311
5. Current Anatek Labs QA Plan (http://www.anateklabs.com/Content/Documents/Current%20QA%20Plan.pdf)

References: Refer to EPA Method 6020A, Method 3051A, Method 3050B, Method 1311

Tables, Diagrams, Flowcharts and Validation Data: Refer to EPA 6020A